

**SYSTEMATIC REVIEW AND META-ANALYSIS OF  
THE ASSOCIATION BETWEEN VITAMIN D LEVELS AND THE  
SINGLE NUCLEOTIDE POLYMORPHISMS, *rs7041*, *rs4588*, AND  
*rs2282679* OF THE GROUP-SPECIFIC COMPONENT GENE (GC)**



**IBRAHIM AFZAL**

**A THESIS SUBMITTED IN PARTIAL FULFILLMENT  
OF THE REQUIREMENTS FOR THE DEGREE OF  
MASTER OF SCIENCE (MEDICAL EPIDEMIOLOGY)  
FACULTY OF GRADUATE STUDIES  
MAHIDOL UNIVERSITY**

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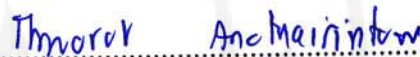
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COMPONENT GENE (GC)**



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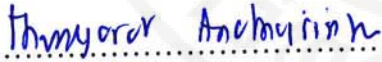
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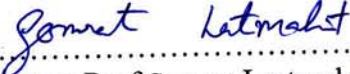
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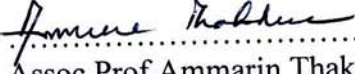
  
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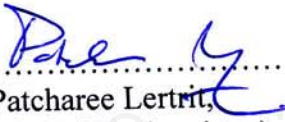
  
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SYSTEMATIC REVIEW AND META-ANALYSIS OF THE ASSOCIATION BETWEEN VITAMIN D LEVELS AND THE SINGLE NUCLEOTIDE POLYMORPHISMS, *rs7041*, *rs4588*, AND *rs2282679* OF THE GROUP-SPECIFIC COMPONENT GENE (GC)

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ABSTRACT

**Background:** Previous literature suggested that certain single nucleotide polymorphisms (SNP) of group-specific component gene (GC) (i.e. *rs7041*, *rs4588*, and *rs2282679*) were associated with serum vitamin D level.

**Objectives:** To assess effects of GC gene including *rs7041*, *rs4588*, and *rs2282679* on serum vitamin D level.

**Methods:** Studies were located from Medline via PubMed and Scopus databases from initiation to 17th January 2015. All observational studies, published in English were selected if they studied the effect of interested SNPs on serum vitamin D level. Mean differences (MD) of serum vitamin D level between genotypes of each SNP were estimated using multivariate meta-analysis.

**Results:** A total of 44 studies met inclusion criteria, 25, 20, and 27 studies had data for *rs4588*, *rs7041*, and *rs2282679*, respectively. For *rs4588*; genotype effect of overall pooled MDs of 25 hydroxy vitamin D (25(OH)D) levels, MD1 (AA versus CC) and MD2 (AC versus CC) for Caucasian adults were, -4.119 (-5.408, -2.831) and -1.871 (95%CI: -2.644, -1.097), and homogenous with the I<sup>2</sup> of 25% and 0%, respectively. For *rs7041*, pooled MD of 25(OH)D level between genotypes TT versus GG (MD1) and TG versus GG (MD2) were, -2.707 (95%CI: -4.91, -0.504) and -1.407 (95%CI: -2.203, -0.612) in Caucasians, with I<sup>2</sup> 71.4% and 0%, respectively. For *rs2282679*, pooled MD of 25(OH)D level between genotypes TT versus GG (MD1) and TG versus GG (MD2) for Caucasians were, -3.598 (95%CI: -5.086, -2.111) and -1.841 (95%CI: -2.743, -0.94), with I<sup>2</sup> of 76% and 80.4%, respectively.

**Conclusions:** The SNPs *rs4588* and *rs2282679* showed negative association with level of serum 25(OH)D.

**KEY WORDS:** VITAMIN D/*rs7041*/ *rs4588*/*rs2282679*/GC

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## LIST OF ABBREVIATIONS

1,25(OH) <sub>2</sub> D	1,25-dihydroxyvitamin D
25(OH)D	25-hydroxyvitamin D
BMI	Body mass index
CI	Confidence interval
GC	Group-specific component
HWE	Hardy-Weinberg equilibrium
MD	Mean difference
n	Number of subjects
OR	Odds ratio
RANK	Receptor Activator of Nuclear Factor Kappa
RANKL	Receptor Activator of Nuclear Factor Kappa Ligand
SD	Standard deviation
SE	Standard error
SNP	Single nucleotide polymorphism

## CHAPTER I INTRODUCTION

### 1.1 Vitamin D synthesis

Vitamin D is a fat soluble vitamin, which can be acquired from various foods (e.g., fish, liver oils, certain plants and fungi) and synthesized in the skin when exposed to the ultra violet blue radiation from sunlight. The synthesis pathway is described in Figure 1.1<sup>1</sup>.

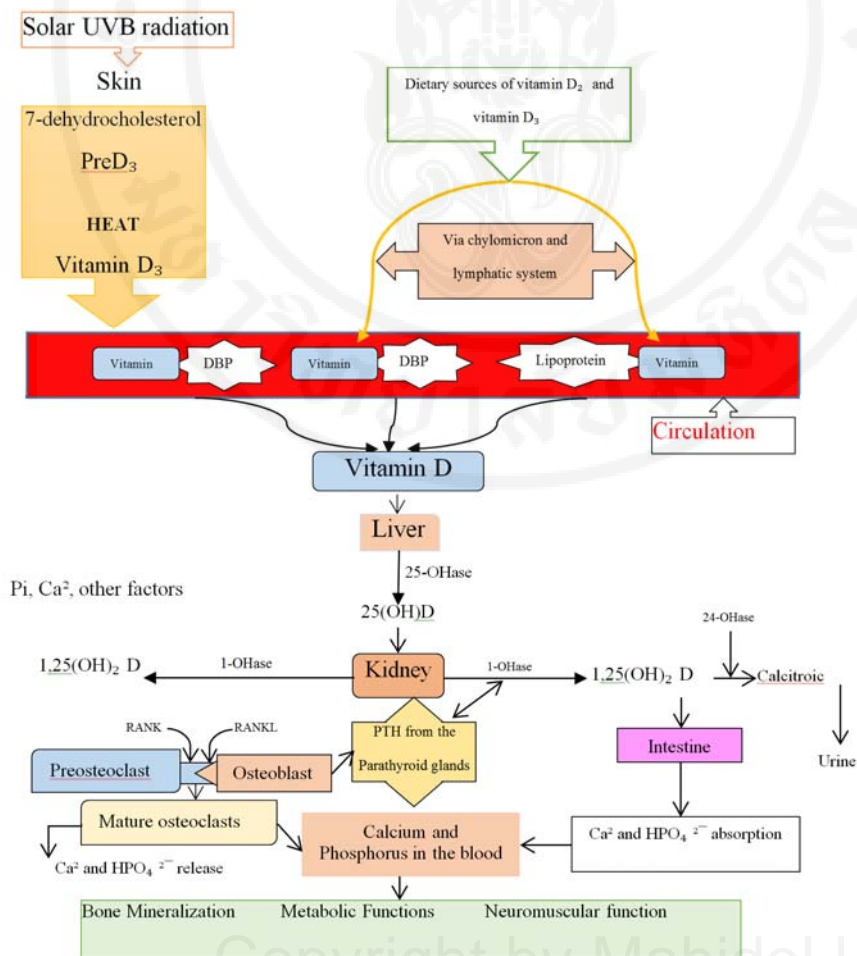


Figure 1.1 Vitamin D synthesis

Two types of vitamin D (i.e., vitamin D<sub>2</sub> (cholecalciferol) and vitamin D<sub>3</sub> (ergocalciferol)) are transported in the blood stream, where D<sub>2</sub> is bounded to chylomicrons and lipoproteins whereas vitamin D<sub>3</sub>, from the skin, is bounded to a group-specific component (GC), which is also known as vitamin D-binding protein<sup>1</sup>. Vitamin D<sub>3</sub> and vitamin D<sub>2</sub> are converted to 25-hydroxyvitamin D (25(OH)D) in the liver by the enzyme 25-hydroxylase (CYP2R1). 25(OH)D is transformed in the kidneys to the active form of vitamin D known as 1,25-dihydroxyvitamin D (1,25(OH)<sub>2</sub>D), which binds to the vitamin D receptor to exert its biological effect. It also plays a role in the absorption of calcium and phosphorus from the intestines, as well as stimulating the expression of Receptor Activator of Nuclear Factor Kappa Ligand (RANKL) on the osteoblasts. This causes it to interact with the Receptor Activator of Nuclear Factor Kappa (RANK) on osteoclasts and subsequently induces the mature osteoclastic activity, resulting in the release of calcium and phosphorus<sup>1</sup>. Also 1,25(OH)<sub>2</sub>D acts as its own inhibitor and causes 24-hydroxylase (CYP24A1) to convert it to its biologically inactive form as calcitroic acid which is water-soluble that is excreted in the bile. 25(OH)D is the main form of vitamin D found in the blood, which makes it a good biomarker of a body's vitamin D level.

Besides sun exposure and diet, serum levels of 25(OH)D can also be affected by other factors such as supplementary vitamin D intake, race, age, sex, obesity, skin color, and tea intake<sup>2</sup>. Genetic factors may also contribute to the serum levels of vitamin D<sup>29, 63, 68</sup>.

## 1.2 Functions of Vitamin D

Vitamin D plays a role in several functions of the body, such as regulating calcium and phosphate homeostasis and playing a maintenance role in the smooth functioning of the musculoskeletal system. Vitamin D has also been implicated in the sustenance of a healthy immune system, cardiovascular function, respiratory function, and in the development of the brain as well as having anti-cancer effects<sup>3</sup>.

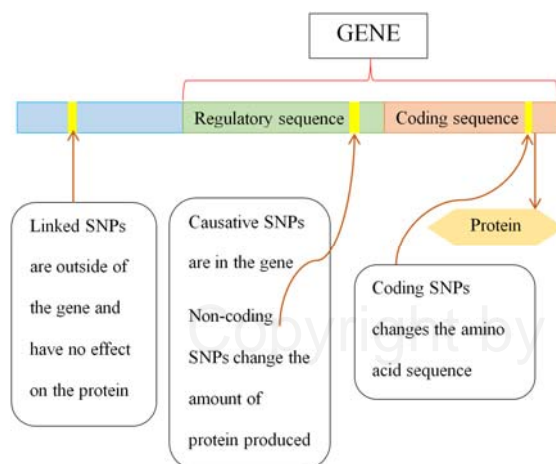
Vitamin D deficiency can increase risk of developing osteoporosis<sup>4</sup>, multiple sclerosis<sup>5</sup>, rickets, metabolic syndrome, hypertension<sup>6</sup>, autoimmune diseases<sup>7</sup>, cardiovascular diseases<sup>8</sup>, type 1 and type 2 diabetes mellitus<sup>9 10 11</sup> as well as several

types of cancer <sup>12 13 14</sup>. Vitamin D deficiency is also commonly found in patients with rheumatoid arthritis <sup>4</sup>, and liver fibrosis. High vitamin D levels may be favorably associated with chronic inflammatory diseases such as multiple sclerosis, and psoriasis <sup>15</sup>.

### 1.3 Genes involved with the vitamin D synthesis pathway

Single nucleotide polymorphisms (SNP, pronounced as “snips), are the most common type of genetic variations among people. Each SNP represents a difference in a single DNA building block, called a nucleotide. For example, a SNP may replace the nucleotide cytosine (C) with the nucleotide thymine (T) in a certain stretch of DNA <sup>16</sup>. A SNP is somewhat like a point mutation, but much more frequent in its occurrence <sup>17</sup>.

SNPs occur in at least 1% of the population for it to be called a SNP. SNPs can be classified into 2 main categories, i.e., linked and causative/disease SNPs, see Figure 2. Linked SNPs (also called indicative SNPs) are not on the disease genes, but they are found nearby <sup>17</sup>. They do not affect the gene expression of proteins, but they do correspond to a particular drug response or the risk of a disease. Causative/disease SNPs can be coding or non-coding SNPs. Coding SNPs are found within the coding region that affect the amino-acid sequence of the protein whereas non-coding SNPs are found in the regulatory sequences of the gene and can change the timing, location or level of the gene expression <sup>17</sup>.



Some of the following genes are involved in the vitamin D synthesis pathway. First, the vitamin D-binding protein is encoded by the GC that transports vitamin D3 in the blood <sup>18</sup>. Vitamin D3 and vitamin D2 are also converted to 25(OH)D in the liver by the enzyme 25-

Figure 1.2 Types of SNPs diagram

hydroxylase which is encoded by the gene CYP2R1<sup>19</sup>.

There have been several studies which assessed the association between genes and serum 25(OH)D levels. These SNPs included GC genes (e.g., rs7041<sup>20-22</sup>, rs4588<sup>23-27</sup>, and rs2282679<sup>28, 29</sup>), and other genes (e.g., rs1155563<sup>30</sup>, rs222016, rs222020, and rs222029<sup>31</sup>, rs2298849<sup>18</sup>, rs842999, rs12512631, rs16846876 and rs17467825)<sup>32</sup> which found to significantly associate with serum levels of 25(OH)D. Among these SNPs, the GC gene at rs7041, rs4588, and rs2282679 were the most common SNPs that had been studied. Genetic risk score analysis also suggested that carriers with no risk alleles of rs4588 and rs842999 had significantly higher serum 25(OH)D concentrations compared to carriers of all risk alleles<sup>32</sup>, or in another word, having the risk genotype of rs7041 was associated with lower levels of 25(OH)D<sup>20, 29</sup>. In addition, subjects who carried risk allele of GC at rs2282679 were increased risk of developing vitamin D deficiency<sup>33, 34</sup>.

#### **1.4 Rationale for conducting a systematic review**

Associations between GC genes and levels of serum 25(OH)D were controversial, i.e., some studies<sup>29</sup> found significant associations whereas some other studies<sup>22, 35</sup> did not. This might be because some of those individual studies had small sample sizes<sup>20, 22, 24, 33, 34, 36-38</sup>, and thus could not detect the gene effects whereas some studies with larger sample sizes could. Increasing the sample size by conducting a systematic review to identify these relevant studies as many as possible, and then pool them together using meta-analysis should be able to increase power to detect gene effects. Therefore, a systematic review and meta-analysis was performed with following research questions.

#### **1.5 Research question**

Is there an association between the vitamin D level and the SNPs, including, rs7041, rs4588, and rs2282679 of the GC gene?

## 1.6 Research objectives

1.6.1 To estimate minor allele effects on serum 25(OH)D levels by estimating the pooled mean differences (MDs) between

- T vs G for rs7041
- A vs C for rs4588
- G vs T for rs2282679

1.6.2 To estimate genotype effects on serum 25(OH)D levels by estimating the pooled MDs between

- TT vs GG and TG vs GG for rs7041
- AA vs CC and AC vs CC for rs4588
- GG vs TT and GT vs TT for rs2282679

1.6.3 To estimate allele effects on vitamin D deficiency or /insufficiency by estimating the pooled odds ratios (OR) of

- T vs G for rs7041
- A vs C for rs4588
- G vs T for rs2282679

1.6.4 To estimate the genotype effects on vitamin D deficiency/insufficiency by estimating the pooled ORs of

- TT vs GG and TG vs GG for rs7041
- AA vs CC and AC vs CC for rs4588
- GG vs TT and GT vs TT for rs2282679

## CHAPTER II

### MATERIALS AND METHODS

#### 2.1 Search strategy

Relevant studies were located from Medline via PubMed and Scopus databases from initiation to January 2015. Reference lists of included studies were explored for additional relevant studies. The search terms were constructed according to P, I, and O domains of PICO, as described in Appendix A. Search strategies were then constructed by combining search terms with ‘OR’ within the same domain, and ‘AND’ for between the domains, as described below for PubMed and Scopus:

PubMed:

The 3 domains were constructed using the search terms:

- (("allele") OR "genotype") OR "polymorphism") OR "polymorphisms") OR "SNPs") OR "single nucleotide polymorphisms"))
- (("rs4588") OR "rs7041") OR "rs2282679") OR "Thr420Lys") OR "Glu416Asp") OR "group specific component") OR "GC gene") OR "GC loci") OR "GC locus"))
- (("1,25(OH)2D") OR "1,25-dihydroxyvitamin D") OR "25(OH)D") OR "25 hydroxyvitamin D") OR "vitamin D"))

The 3 terms were then combined with ‘AND’ as follows:

- (("allele") OR "genotype") OR "polymorphism") OR "polymorphisms") OR "SNPs") OR "single nucleotide polymorphisms")) AND (("rs4588") OR "rs7041") OR "rs2282679") OR "Thr420Lys") OR "Glu416Asp") OR "group specific component") OR "GC gene") OR "GC loci") OR "GC locus")) AND (("1,25(OH)2D") OR "1,25-dihydroxyvitamin D") OR "25(OH)D") OR "25 hydroxyvitamin D") OR "vitamin D"))

Scopus:

The 3 domains were constructed using the search terms:

- ( "allele" OR "genotype" OR "polymorphism" OR "polymorphisms" OR "SNPs" OR "single nucleotide polymorphisms" )
- ( "rs4588" OR "rs7041" OR "rs2282679" OR "Thr420Lys" OR "Glu416Asp" OR "group specific component" OR "GC gene" OR "GC loci" OR "GC locus" )
- ( "1, 25(OH)2D" OR "1, 25-dihydroxyvitamin D" OR "25(OH)D" OR "25 hydroxyvitamin D" OR "vitamin D" )

The 3 terms were then combined with 'AND' as follows:

- ( "allele" OR "genotype" OR "polymorphism" OR "polymorphisms" OR "SNPs" OR "single nucleotide polymorphisms" ) AND ( "rs4588" OR "rs7041" OR "rs2282679" OR "Thr420Lys" OR "Glu416Asp" OR "group specific component" OR "GC gene" OR "GC loci" OR "GC locus" ) AND ( "1, 25(OH)2D" OR "1, 25-dihydroxyvitamin D" OR "25(OH)D" OR "25 hydroxyvitamin D" OR "vitamin D" )

## 2.2 Selection of studies

### Inclusion criteria

All observational studies published in English were included, if they met all of the following criteria.

- Studied in human.
- Studied the effect of genotypes or alleles of any of the following SNPs: rs2282679, rs7041, or rs4588.
- Measured any of the following outcomes: serum 25(OH)D, or serum 1,25(OH)2D levels for continuous outcomes; or vitamin D insufficiency or deficiency for dichotomous outcomes.
- Have sufficient data for pooling, i.e. mean and standard deviation (SD) or the beta coefficients and its standard error for continuous outcome (e.g., serum 25(OH)D levels) between allele/genotype groups; reported the contingency table of

interested allele/genotypes and vitamin D insufficiency or deficiency; or odds ratio (OR) and the 95% CI the for dichotomous outcomes).

Studies that did not meet with our inclusion criteria were coded, with an ineligibility criterion coding, see Appendix B.

### **2.3 Study selection**

Initial studies were identified from Medline via PubMed and Scopus and then managed using the EndNote software X7 by remove duplicates. The studies were independently selected based on the title and abstracts by two reviewers (IA and TA), studies that did not meet the eligibility criteria were eliminated with providing the reasons. The full text articles were retrieved and read to make decisions to include in the review. Results of study selection between the two reviewers were then validated. Any disagreements were solved by discussion and consensus. A kappa test was applied to assess agreements between the two reviewers.

### **2.4 Data extraction**

Summary data were extracted independently by two reviewers (IA and TA) using a standardized data extraction form, see Appendix C. The data extraction forms consisted of general characteristics of studies and studied subjects, The general characteristics of the studies included the author, journal, year of publication, type of study design, types of SNP, and whether the HWE was checked or not. The characteristics of the studied subjects were those that may be associated with the vitamin D level. These included, age, gender, obesity, physical activity, kidney disease, liver disease, sun exposure, alcohol intake, and vitamin D supplement.

Data for pooling were continuous and dichotomous outcomes. For continuous outcome, total number of studied subjects, mean, and SD of the serum vitamin D levels by allele/genotype groups were extracted. For those studies did not report mean and SD but instead reported the beta coefficients and the SE, these data

were extracted and combined with estimated MDs from those studies reported mean and SD.

For dichotomous outcome, the frequencies of the alleles and the genotypes between outcome groups were collected. ORs and along with 95% CI were used for those studies did not report frequency data. Any disagreement was resolved by consensus. Data cleaning and checking were performed separately for each study.

## **2.5 Risk of bias assessment**

The risk of bias was assessed by modifying the form constructed by Thakkinstian et al<sup>39</sup>. The forms were used separately by type of study design (i.e., the case-control, cohort, and cross-sectional study), see Appendix D. The forms consisted of 6 domains as follows:

For case control studies

### **2.5.1 Selection bias**

#### **2.5.1.1 Representativeness of cases and controls assessed**

whether the cases and controls were consecutive/randomly selected from their population with a clearly defined random frame.

### **2.5.2 Information bias**

#### **2.5.2.1 Ascertainment of vitamin D level assessed whether the**

method of vitamin D measurement was clearly given, and also whether definition of vitamin D deficiency or insufficiency was clearly defined.

#### **2.5.2.2 Ascertainment of genotyping examination assessed**

whether the genotyping was done with blinding, whether the error rate was reported and whether any quality control measures were taken.

### **2.5.3 Confounding bias.**

#### **2.5.3.1 Population stratification assessed whether any**

population stratification was done.

#### **2.5.3.2 Confounding variables assessed whether any**

confounding variables were adjusted in analysis.

#### 2.5.4 Multiple testing.

2.5.4.1 Assessed whether any adjustment was done for multiple testing.

#### 2.5.5 Selective reporting.

2.5.5.1 Assessed whether the study reported the results of all polymorphisms mentioned in objectives, regardless significant or not.

#### 2.5.6 HWE.

2.5.6.1 Assessed whether Hardy-Weinberg equilibrium (HWE) was checked.

For cross sectional and cohort studies

#### 2.5.7 Selection bias.

2.5.7.1 Representativeness assessed whether the study subjects were consecutive/randomly selected from the target population with a clearly defined random frame.

#### 2.5.8 Information bias.

2.5.8.1 Ascertainment of vitamin D level assessed whether the method of vitamin D measurement was clearly described.

2.5.8.2 Ascertainment of genotyping examination assessed whether the genotyping was done with blinding, whether the genotyping error rate was reported and whether any quality control measures were taken.

#### 2.5.9 Confounding bias.

2.5.9.1 Population stratification assessed whether any population stratification was done.

2.5.9.2 Confounding variables assessed whether any confounding variables were adjusted when means were compared between allele/genotype groups.

2.5.10 Multiple testing.

2.5.10.1 Assessed whether any adjustment was done for multiple testing.

2.5.11 Selective reporting.

2.5.11.1 Assessed whether the study reported the results of all polymorphisms mentioned in objectives, regardless significant or not.

2.5.12 HWE.

2.5.12.1 Assessed whether HWE was checked.

Each item was asked whether there was low risk of bias and four possible answers were:

- Yes, meant there was low risk of bias.
- No, meant there was high risk of bias
- Unclear, if the information was not clearly given and thus could not make the decision to be yes or no.
- Inapplicability meant that the item could not be assessed.

## 2.6 Studied polymorphism

The three SNPs studied were rs7041, rs4588, and rs2282679.

The SNP rs2282679 has the alleles T (major) and G (minor) and the genotypes TT, GT, and GG.

The SNP rs7041 has the alleles G (major) and T (minor) and the genotypes GG, TG, and TT.

The SNP rs4588 has the alleles C (major) and A (minor) and the genotypes CC, AC, and AA.

## 2.7 Outcome of interest

The outcome of interest was the serum vitamin D reported according to the individual studies as:

### 2.7.1 25(OH)D levels

The 25(OH)D levels were reported in the units of ng/ml. For those studies reported as nmol/l, they were converted to ng/ml by dividing with the conversion factor of 2.496<sup>40</sup>.

### 2.7.2 Vitamin D deficiency or vitamin D insufficiency

Vitamin D was classified as deficiency or insufficiency according to original studies. Briefly, the 25(OH)D < 20 ng/ml and 21 – 29 ng/ml were respectively considered as vitamin D deficiency and insufficiency, as for the American Endocrine Society clinical practice guideline<sup>41</sup>.

## 2.8 Statistical analysis

### 2.8.1 Kappa test for agreement in study selection.

Percent agreement of study selection between 2 reviewers was assessed using Kappa statistic. The kappa was calculated using the equation below<sup>42</sup>.

$$K = \frac{(O - E)}{(1 - E)}$$

Where is  $K$  the Kappa,  $O$  is the observed agreement, and  $E$  is the expected agreement.

Observed agreement is the total number of agreements (%) in both included and excluded studies by both reviewers (IA and TA), i.e., (a +d) as shown in Table 2.1.

Expected agreement was calculated using equation given below <sup>42</sup>.

$$E = \left( \left( \frac{n_1}{n} \right) \left( \frac{m_1}{n} \right) \right) + \left( \left( \frac{n_0}{n} \right) \left( \frac{m_0}{n} \right) \right)$$

Where *E* is the expected agreement, *n*<sub>1</sub> is the total number of included studies by reviewer 1, *m*<sub>1</sub> is the total number of included studies by reviewer 2, *n*<sub>0</sub> is the total number of excluded studies by reviewer 1, *m*<sub>0</sub> is the total number of excluded studies by reviewer 2, *n* is the total number of selected studies

Level of agreement can be categorized as poor, slightly poor, fair, moderate, substantial, and perfect chance of agreement if the Kappa statistic ranged from 0.0 to <0.2 , 0.2 to < 0.4, 0.4 to < 0.6, 0.6 to <0.8, 0.8 to <1, and 1.0, respectively.

**Table 2.1** Example of observed and expected values of both reviewers for calculating Kappa

Observed		Reviewer 2 (TA)		
		Included	Excluded	Total
Reviewer 1 (IA)	Included	a	b	m <sub>1</sub>
	Excluded	c	d	m <sub>0</sub>
Total		n <sub>1</sub>	n <sub>0</sub>	n
Expected		Reviewer 2 (TA)		
		Included	Excluded	Total
Reviewer 1 (IA)	Included	e <sub>11</sub>	e <sub>12</sub>	m <sub>1</sub>
	Excluded	e <sub>21</sub>	e <sub>22</sub>	m <sub>0</sub>
Total		n <sub>1</sub>	n <sub>0</sub>	n

### 2.8.2 HWE

The HWE was assessed only in the control groups if the study design was case-control whereas the whole data was used for cohort/cross-sectional studies. Suppose C and A was the major and minor alleles respectively. The HWE was assessed to see if the distribution of the genotypes CC, AC, and AA in the control group were complied with frequency of  $p^2$ ,  $2pq$ , and  $q^2$  respectively. We used the Chi square or Fisher's exact test where appropriate (i.e., if the sample size was too small in more than 20% of the cells) to see whether they deviated from the HWE. A sensitivity analysis was done for those studies which showed Hardy-Weinberg disequilibrium (HWD).

### 2.8.3 Pooling minor allele prevalence

The minor allele frequency of the studied SNP was pooled using data from control groups if the design was case-control study; otherwise the whole data was used. Where the data for type of subject (i.e., children and adults) and ethnicity were available, we pooled the prevalence of minor allele, stratified accordingly by ethnicity and type of subjects.

Overall prevalence of minor allele A was pooled using the inverse variance method. This was done according to the equation for pooling as given<sup>43</sup>.

$$\bar{p} = \frac{\sum w_i p_i}{\sum w_i}$$

Where  $\bar{p}$  was the pooled prevalence of the minor allele,  $p_i$  was the prevalence of the minor allele in each study, and  $w_i$  was the inverse variance

$\left( \frac{1}{\text{var}(p_i)} \right)$ , which was the weight of each study.

The 95% CI was calculated using the using the equation as given<sup>43</sup>.

$$95\%CI = \bar{p} \pm \frac{1.96}{\sqrt{\sum w_i}}$$

*Preparing the data for pooling prevalence.*

As for minor and major A and C alleles, a total number of A and C alleles, total number of alleles (A +C) in the control population were calculated from genotype data as follows:

$$\text{Total A allele} = 2 \times n(\text{AA}) + n(\text{AC})$$

$$\text{Total C allele} = 2 \times n(\text{CC}) + n(\text{AC})$$

$$\text{Total number of alleles} = \text{Total A allele} + \text{Total C allele.}$$

The 'pmeta' command from STATA was used to estimate the pooled prevalence of the minor allele A along with its 95% CI, and also heterogeneity parameters. If the heterogeneity was present, a random-effect model was applied for pooling.

#### 2.8.4 Assess the gene effects

We applied both per-allele and per genotype approaches for assessing gene effects as follows.

##### 2.8.4.1 Per-allele approach

Suppose that A and C were minor and major alleles, respectively.

##### 2.8.4.1.1 For continuous outcome

For 25(OH)D outcome, the MD of 25(OH)D between C and A alleles along with its variance and 95% CIs were estimated for each study. For those studies provided only coefficients and the SE of allele effect, these data were later appended with calculated MDs. Unstandardized MD was applied to pool MDs across studies, using the equations below:

$$\hat{D} = \frac{\sum_{i=1}^k w_i d_i}{\sum_{i=1}^k w_i}$$

$$d_i = (\bar{x}_{1i} - \bar{x}_{2i}), \text{ var}(d_i) = \frac{sd_{1i}^2}{n_{1i}} + \frac{sd_{2i}^2}{n_{2i}}, w_i = \frac{1}{\text{var}(d_i)}$$

$$95\%CI = d_i \pm 1.96 \sqrt{\text{var}(d_i)}$$

Where  $\hat{D}$  is the pooled unstandardized mean difference, and  $d_i$  is the unstandardized mean differences.  $\bar{x}_{1i}$  and  $\bar{x}_{2i}$  are the means of 25(OH)D levels in subjects with C allele and A allele.  $w_i$  is the weight, and  $\text{var}(d_i)$  is the variance of the unstandardized mean differences.  $n_{1i}$  and  $n_{2i}$  are the number of A and C alleles.  $sd_{1i}^2$  and  $sd_{2i}^2$  are the SDs of the MDs for subjects with A and C alleles.

The fixed-effect model was applied if there was no heterogeneity; otherwise a random-effect model was applied.

#### 2.8.4.1.2 Dichotomous outcome

For studies which reported the frequency data of A and C alleles between controls and vitamin D deficiency/insufficiency, log OR along with its variance and the 95%CI were estimated for each study. Some studies did not report the frequency of genotype data but instead provided the ORs and the 95%CIs. In that case, this data were later combined with ORs of those studies that reported frequency of genotype data. The pooled OR was then estimated using inverse variance method if the heterogeneity was not present, otherwise a random-effect model using the Der-Simonian and Laird method was applied.

For the fixed effects model, the equation was given below:

$$\ln \hat{OR}_{pooled} = \frac{\sum_j^k w_j \ln OR_j}{\sum_j^k w_j}$$

$$OR_j = \frac{a_j d_j}{b_j c_j}$$

$$\text{var} \ln(OR_j) = \ln \left( \frac{1}{a_j} + \frac{1}{b_j} + \frac{1}{c_j} + \frac{1}{d_j} \right)$$

$$w_j = \frac{1}{\text{var}_j}$$

$$95\%CI \ln(\hat{OR})_j = \ln(\hat{OR})_j \pm 1.96\sqrt{\text{var} \ln(\hat{OR})_j}$$

$\ln \hat{OR}_{pooled}$  is the pooled log (OR) of allele A versus C,  $\ln OR_j$  is the natural logarithm of OR of each study,  $\text{var} \ln(OR_j)$  is the variance of the log (OR), and  $w_j$  is the weight. The frequencies of C alleles for cases and controls are a and b respectively. The frequencies of A alleles for cases and controls are c and d respectively.

#### 2.8.4.2 The genotype approach.

##### 2.8.4.2.1 For continuous outcomes

The MD<sub>1</sub> (AA versus CC) and MD<sub>2</sub> (AC versus CC) and their respective SEs were estimated for each study. A multivariate meta-analysis with a random-effect model was applied for pooling genotype effects of MD<sub>1</sub> and MD<sub>2</sub>. A maximum likelihood function was used for estimation of the pooled MDs assuming zero variance-covariance between MD<sub>1</sub> and MD<sub>2</sub>. In addition, pooled MD<sub>3</sub> (AA versus AC) was then estimated using a linear combination of MD<sub>1</sub> – MD<sub>2</sub> from the multi-variate meta-analysis model.

##### 2.8.4.2.2 For dichotomous outcomes

For dichotomous outcome, the data were reshaped from wide format to long format with 3 × 2 observations per study. The summary long format data were then expanded to be individual patient data using ‘expand’ command. The log-scales for OR<sub>1</sub> (AA versus CC), OR<sub>2</sub> (AC versus CC), variance of  $\ln(OR_1)$ ,  $\ln(OR_2)$ , and covariance ( $\ln OR_1$  and  $\ln OR_2$ ) were then estimated using the ‘mvmeta make’ command with logit model. Summary ORs and their variances were then appended to these estimations for those studies where frequency data were not available. The multi-variate meta-analysis was then used to pool OR<sub>1</sub> and OR<sub>2</sub> across studies simultaneously. In addition, the pooled OR<sub>3</sub> (AA versus AC) was then estimated using a linear combination of  $\ln OR_1 - \ln OR_2$  from the multi-variate meta-analysis model.

### 2.8.5 Assess the heterogeneity.

To check for heterogeneity of gene effects, we used the Cochran Q test and quantified the degree of heterogeneity ( $I^2$ ). The p value  $< 0.1$  from Cochran Q test or the  $I^2 \geq 25$  was considered as presence of heterogeneity. If heterogeneity was present, we pooled the data using the random-effect model; otherwise the fixed-effect model was used. The null hypothesis for heterogeneity states that there is no heterogeneity among the effect size (e.g., ORs/MDs) across studies. Heterogeneity was assessed by the following equations:

2.8.5.1 Assessing the heterogeneity for pooling minor allele prevalence.

$$Q = \sum w_i (p_i - \bar{p})^2$$

Where  $Q$  is the heterogeneity,  $\bar{p}$  is the pooled prevalence of the allele,  $p_i$  is the prevalence of the allele in each study, and  $w_i$  is the inverse variance ( $1/\text{var}(p_i)$ ).

If heterogeneity was present, then the between-study variation ( $\tau^2$ ) was estimated with the equation given below<sup>43</sup>:

$$\tau^2 = \frac{Q - (k-1)}{\sum w_i - \frac{\sum w_i^2}{w_i}} \quad \text{if } Q > k-1 \text{ or } 0 \text{ otherwise.}$$

The weight could be calculated taking into account for  $\tau^2$  as

$$w_i^* = \frac{1}{\text{var}(p_i) + \tau^2}$$

The pooled prevalence could then be calculated with the random-effect equation as

$$\bar{p}^* = \frac{\sum w_i^* p_i}{\sum w_i^*}$$

And the 95% CI could be estimated using the equation

$$95\%CI = \bar{p}^* \pm \frac{1.96}{\sqrt{\sum w_i^*}}$$

2.8.5.2 Heterogeneity for OR.

$$Q = \sum w_i (\hat{\theta}_i - \hat{\theta}_p)^2$$

$$\hat{\theta}_p = \ln \widehat{OR}_p = \frac{\sum_{i=1}^k w_i \ln \widehat{OR}_i}{\sum_{i=1}^k w_i}$$

$$\hat{\theta}_i = \ln \widehat{OR}_i$$

In the above equations,  $Q$  is heterogeneity,  $w_i$  and  $var(\ln \widehat{OR}_i)$  are the variance and weight as defined previously<sup>44-46</sup>.

2.8.5.3 Heterogeneity for MD

$$Q = \sum_i^k w_i (d_i - \widehat{D})^2$$

$$\widehat{D} = \frac{\sum_i^k w_i d_i}{\sum_i^k w_i}$$

$$w_i = \frac{1}{var(d_i)}$$

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$Q$  is heterogeneity Chi-square test,  $w_i$  is weight which is the

inverse variance,  $var(d_i)$  is the variance of MD as defined previously,  $d_i$  is the unstandardized MD, and  $\hat{D}$  is the pooled MD. The subscript  $i$  stands for the study  $i$ .

A meta-regression analysis was done to explore for the source of heterogeneity by adding the covariables (e.g., age, sex, obesity, physical activity, kidney disease, liver disease, ethnicity, sun exposure, season, country, alcohol intake, and vitamin D supplements) into the model one by one, if the studies reported the covariables needed. If adding the variable could decrease the between-study variation, or the  $I^2$ , this would indicate that added variable may be a source of heterogeneity. A subgroup analysis by this variable was then done next<sup>44-47</sup>.

#### 2.8.6 Publication bias

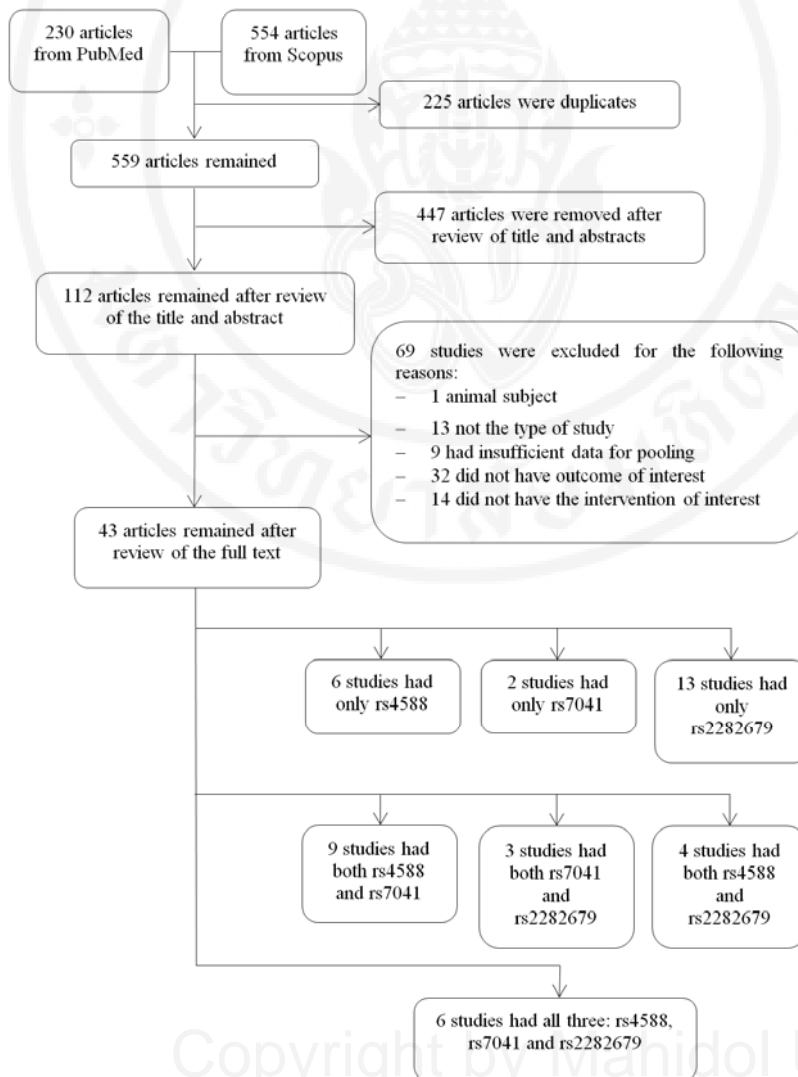
For both dichotomous and continuous data, publication bias was assessed by plotting a funnel plot, which plots the log OR (for dichotomous outcome) and the MD (for continuous outcomes) on the x-axis and the SE of the log OR or the SE of the MD on the y-axis. If there was asymmetry, this suggested that there was the possibility of small study effects, or it could be due to heterogeneity or publication bias. An Egger test was done to test for asymmetry. We also did a contour enhanced funnel plot to differentiate between publication bias and heterogeneity.

STATA version 13<sup>48</sup> was used to perform all statistical analyses. P value of  $< 0.05$  was considered as statistically significant, except in the Cochran Q test where the p value of  $< 0.1$  was use.

## CHAPTER III RESULTS

### 3.1 Study selection

As described in Figure 3.1, 559 studies were identified from the two main databases, of which 43 studies<sup>10, 12, 20-29, 33-38, 49-73</sup> were finally met inclusion criteria.



**Figure 3.1** Flow chart of the study selection

Among the 559 studies, 447 studies were excluded after review of the title and abstract. From the remaining 112 studies, 69 studies were excluded after review of the full text. Of which, 13 studies<sup>14, 74-85</sup> were not observational studies, 1 study<sup>86</sup> was not done in human, 14 studies<sup>7, 87-99</sup> did not have the gene of interest, 32 studies<sup>11, 16, 100-129</sup> did not have the outcome of interest, and 9 studies<sup>130-138</sup> did not have sufficient data for pooling. Among 43 studies selected, 6<sup>12, 23, 36, 50, 53, 58</sup>, 2<sup>54, 59</sup>, and 13<sup>10, 28, 49, 51, 52, 55, 62, 65-67, 69, 71, 72</sup> studies had data for rs4588, rs7041, and rs2282679, respectively. Nine studies<sup>20, 22, 24, 27, 37, 38, 60, 61, 64</sup> had data for both rs4588 and rs7041, 3 studies<sup>29, 63, 68</sup> had data for both rs7041 and rs2282679, 4 studies<sup>25, 26, 57, 70</sup> had data for both rs4588 and rs2282679, and 6 studies<sup>21, 33-35, 56, 73</sup> had data for all three SNPs. Agreement for selection of studies between the two reviewers was described in Table 3.1, which suggested 97.2% agreement with the Kappa statistic of 0.779. Characteristics of these included studies were described as follows:

**Table 3.1** Selected studies by reviewers

		Reviewer 1 (TA)		Total
		Included	Excluded	
Reviewer 2 (IA)	Included	29	6	35
	Excluded	9	488	497
Total		38	494	532

### 3.2 Characteristics of included selection

Characteristic of included studies were described in Table 3.2 Among 43 studies, most studies were done on Caucasian (n =22) and Asian populations (n=12), and only a few studies conducted in Africans (n=4), Hispanics (n=3), and some (n = 9) had mixed ethnicities; some studies reported for more than one ethnicity. Most common study designs were cross-sectional (n = 25), followed by case-controls (n = 10), cohort studies (n = 7), and 1 study<sup>66</sup> had reported both cross sectional and cohort study designs. Mean age and body mass index ranged from 5.9 to 83.8 and 17.4

to 48.6, respectively. Male percentage ranged from 0 to 100. Most studies (n = 19) used radioimmunoassay method for measurement of vitamin D.

### 3.3 Risk of bias

We assessed the risk of bias as described in Tables 3.3-3.4. Among 43 studies, the risk of bias was highest in selective reporting (11/43, 25.58%), followed by confounding bias (8/43, 18.60%) and HWE (7/43, 16.28%). The risk of bias was lowest in adjustment for multiple testing (1/43, 2.33%), followed by representativeness (2/43, 4.65%), and ascertainment of 25(OH)D level of controls in case control studies and for the non-exposure group in other study designs (2/43, 4.65%).

### 3.4 rs4588

Twenty five<sup>12, 20-27, 33-38, 50, 53, 56-58, 60, 61, 64, 70, 73</sup> studies were included in the pooling for the SNP rs4588, out of which, 4<sup>33, 34, 37, 38</sup> studies had data for pooling OR for vitamin D deficiency/insufficiency, 18<sup>12, 20, 22-27, 35, 36, 50, 53, 56, 58, 60, 61, 64, 73</sup> studies had data for pooling MD for vitamin D level, and 3 studies<sup>21, 57, 70</sup> had both data. From the 7 with vitamin D deficiency/insufficiency, 5<sup>21, 33, 34, 37, 38</sup> studies reported frequency data for genotypes, 2<sup>57, 70</sup> studies reported ORs and 95% CIs for additive model.

For 21 studies with vitamin D level, 10 studies<sup>12, 20, 22-25, 27, 36, 53, 61</sup> reported means by genotypes, 1 study<sup>26</sup> reported MD, 2 studies<sup>58, 64</sup> reported MD by combined genotypes (AC and AA), 6 studies<sup>21, 35, 56, 57, 70, 73</sup> reported MD for additive model, 2 studies<sup>50, 60</sup> reported MD for allele data.

**Table 3.2** Characteristics of the included studies

First Author, Year, (Reference No.)	Study design	Age group	Ethnicity	Mean Age	BMI	% Male	Method of measurement of vitamin D
Sinotte, 2009 <sup>27</sup>	Cross-sectional study	Adult	Caucasian	46.8	25.2	0	Radioimmunoassay
Ahn, 2010 <sup>49</sup>	Case-control study	Adult	Caucasian			60.9	Chemiluminescence assay
Bu, 2010 <sup>35</sup>	Cross-sectional study	Mixed <sup>a</sup>	Caucasian	53	27.0		Radioimmunoassay
Janssens, 2010 <sup>20</sup>	Case-control study	> 50 years	Caucasian		26.9	79.0	Radioimmunoassay
Wang, 2010 <sup>67</sup>	Cross-sectional study	> 50 years	Caucasian	48.7	26.9	36	Radioimmunoassay, Chemiluminescence assay, ELISA,
Gozdzik, 2011 <sup>53</sup>	Cross-sectional study	Adult	South Asian	22.3	23.7	35.6	Mass spectrometry
	Cross-sectional study	Adult	East Asian	20.5	22.4	29.1	Liquid chromatography
	Cross-sectional study	Adult	Caucasian	20.9	23.5	34.7	Liquid chromatography
Signorello, 2011 <sup>62</sup>	Cohort study	Adult	African	51.9	28.3	50.7	Radioimmunoassay
Azad, 2012 <sup>12</sup>	Cross-sectional study	Adult	Mixed <sup>b</sup>			79	Not given
Berry, 2012 <sup>50</sup>	Cross-sectional study	Adult	Caucasian				ELISA
Hibler, 2012 <sup>54</sup>	Cross-sectional study	Adult	Caucasian	66.0	27.9	66.1	Radioimmunoassay
Kitanaka, 2012 <sup>33</sup>	Case-control study	Children	Asian			86.7	Not given
Lu, 2012 <sup>56</sup>	Cross-sectional study	> 50 years	Asian	58.6	24.4	44.3	Radioimmunoassay
Muindi, 2012 <sup>57</sup>	Case-control study	Adult	Mixed			54.2	Liquid chromatography
Petta, 2012 <sup>59</sup>	Cross-sectional study	Mixed	Mixed	52.8		49.2	Liquid chromatography
Suzuki, 2012 <sup>64</sup>	Cross-sectional study	Adult	Asian	72.1	22.1	47.5	Radioimmunoassay
Theodoratou, 2012 <sup>28</sup>	Case-control study	Adult	Caucasian	60.7	26.8	56.5	Liquid chromatography

**Table 3.2** Characteristics of the included studies (continued)

First Author, Year, (Reference No.)	Study design	Age group	Ethnicity	Mean Age	BMI	% Male	Method of measurement of vitamin D
Trummer, 2012 <sup>66</sup>	Cross-sectional study	Adult	Caucasian	55.3	48.6	64	Radioimmunoassay
Buijsse, 2013 <sup>10</sup>	Cohort study	Adult	Caucasian	83.8	25.7	15	Radioimmunoassay
Carpenter, 2013 <sup>23</sup>	Cohort study	Adult	Mixed	50.6	26.0	43.1	Mass chromatography
Cheung, 2013 <sup>51</sup>	Cross-sectional study	Children	Mixed				Radioimmunoassay
Foucan, 2013 <sup>52</sup>	Cross-sectional study	Mixed	Asian	48.4	22.8		Radioimmunoassay
Ismail, 2013 <sup>55</sup>	Cross-sectional study	Adult	African	46	27	42	Chemilluminescence assay
Nimitphong, 2013 <sup>58</sup>	Case-control study	Children	African	7.8		54.9	ELISA
Perna, 2013 <sup>26</sup>	Case-control study	Adult	Mixed	36.2	22.8	18.0	Liquid chromatography
Powe, 2013 <sup>60</sup>	Cohort study	> 50 years	Caucasian	62			Chemilluminescence assay
Robien, 2013 <sup>61</sup>	Cross-sectional study	Adult	Caucasian	48.3	29.8	44	Radioimmunoassay
Santos, 2013 <sup>37</sup>	Cross-sectional study	Adult	African	48.3	29.4	44.3	Radioimmunoassay
Singh, 2013 <sup>22</sup>	Cohort study	Adult	Asian	55.7	22.8	43.7	Chemilluminescence assay
Trummer, 2013 <sup>65</sup>	Cross-sectional study	Children	Hispanic	13.2		0	Radioimmunoassay
Zhang, 2013 <sup>70</sup>	Cross-sectional study	Adult	Mixed		26.3	27.3	Liquid chromatography
Gilbert, 2014 <sup>73</sup>	Cohort study	> 50 years	Caucasian	62.6	22.6	69.7	Radioimmunoassay
Laursen, 2014 <sup>29</sup>	Cross-sectional study	Adult	Asian		23.3	20.5	Electrochemiluminescence
Leong, 2014 <sup>72</sup>	Case-control study	> 50 years	Caucasian	62.6		100	Mass spectrometry
Li, 2014 <sup>21</sup>	Cross-sectional study	Mixed	Mixed	42.4		29	Chemilluminescence assay
	Case-control study	Adult	Mixed	65.7	27.5	34.1	Chemilluminescence assay
	Cross-sectional study	Adult	Asian	48.6	23.6	62.5	Radioimmunoassay

**Table 3.2** Characteristics of the included studies (continued)

First Author, Year, (Reference No.)	Study design	Age group	Ethnicity	Mean Age	BMI	% Male	Method of measurement of vitamin D
Navas-Nazario, 2014 <sup>24</sup>	Cohort study	Children	Hispanic	5.9		47	Radioimmunoassay
Nissen, 2014 <sup>25</sup>	Cross-sectional study	Adult	Caucasian		25.5	50.5	Liquid chromatography
	Cross-sectional study	Children	Caucasian		17.4	52.0	Liquid chromatography
Pekkinen, 2014 <sup>36</sup>	Cross-sectional study	Children	Caucasian	12.9	19.7	30.7	Liquid chromatography
Strawbridge, 2014 <sup>63</sup>	Cross-sectional study	> 50 years	Caucasian		26.8	48.1	Chemilluminescence assay
Suaini, 2014 <sup>34</sup>	Cross-sectional study	Children	Caucasian				Liquid chromatography
	Cross-sectional study	Children	Asian				Liquid chromatography
Thongthai, 2014 <sup>71</sup>	Cross-sectional study	Adult	Asian	47.8	23.6	61.4	Liquid chromatography
Touvier, 2014 <sup>38</sup>	Cross-sectional study	Adult	Caucasian			46	Chemilluminescence assay
Wang, 2014 <sup>68</sup>	Case-control study	Adult	Hispanic	52.8	29.7	0	Liquid chromatography
	Case-control study	Adult	Caucasian	56.8	26.8	0	Liquid chromatography
Yoshida, 2014 <sup>69</sup>	Cohort study	Mixed	Asian	64.3	21.2	12.3	Radioimmunoassay

<sup>a</sup>The age range of these studies was not specified, <sup>b</sup>The ethnicity of these study populations was not specified  
BMI; body mass index





**Table 3.3** Risk of bias assessment (continued)

First Author, Year, (Reference No.)	Selection bias		Information bias		Confounding bias				HWE
	Rep <sup>a</sup> cont <sup>b</sup>	Diff part <sup>c</sup>	Ascertainment of vitamin D (cases/exposure group)	Ascertainment of vitamin D (controls/non- exposure group)	Genotyp <sup>d</sup>	Population stratification	Multip Test <sup>e</sup>	Selective reporting	
Singh, 2013 <sup>22</sup>	Yes		Yes	Yes	Yes	Yes	Yes	Yes	Yes
Trummer, 2013 <sup>65</sup>	Yes		Yes	Yes	Yes	Yes	Yes	Yes	Yes
Zhang, 2013 <sup>70</sup>	Yes		Yes	Yes	Yes	Yes	Yes	No	Yes
Laursen, 2014 <sup>29</sup>	Yes		Yes	Yes	Yes	No	Yes	Yes	No
Li, 2014 <sup>21</sup>	Yes		Yes	Yes	Yes	Yes	Yes	Yes	Yes
Navas-Nazario, 2014 <sup>24</sup>	Yes		Yes	Yes	Yes	No	Yes	Yes	Yes
Nissen, 2014 <sup>25</sup>	Yes		Yes	Yes	Yes	No	Yes	Yes	Yes
Pekkinen, 2014 <sup>36</sup>	Yes		Yes	Yes	Yes	Yes	Yes	No	Yes
Strawbridge, 2014 <sup>63</sup>	Yes		Yes	Yes	Yes	Yes	Yes	Yes	Yes
Suaini, 2014 <sup>34</sup>	Yes		Yes	Yes	Yes	Yes	Yes	Yes	Yes
Thongthai, 2014 <sup>71</sup>	Yes		Yes	Yes	Yes	Yes	Yes	Yes	Yes
Touvier, 2014 <sup>38</sup>	Yes		Yes	Yes	Yes	Yes	Yes	Yes	Yes
Yoshida, 2014 <sup>69</sup>	Yes		Yes	Yes	Yes	Yes	Yes	Yes	Yes

<sup>a</sup> Representativeness of cases in case control studies and subjects in other study designs, <sup>b</sup> representativeness of controls in case control studies, <sup>c</sup> differential participation of cases and controls, <sup>d</sup> method of genotyping, <sup>e</sup> adjustment for multiple testing  
HWE; Hardy-Weinberg equilibrium

**Table 3.4** Summary of risk of bias assessment

Domain	Items assessed for risk of bias	low risk of bias		Moderate to high risk of bias	
		Frequency	%	Frequency	%
Selection bias	Representativeness	41	95.35	2	4.65
	Representativeness of controls in case control studies	7	70.00	3	30.00
	Differential participation for case and control	4	40.00	6	60.00
Information bias	Ascertainment of 25hydroxyvitamin D level	40	93.02	3	6.98
	Ascertainment of 25hydroxyvitamin D level of controls in case control studies and for the non-exposure group in other study designs	41	95.35	2	4.65
	Ascertainment of genotyping	39	88.64	4	9.30
Confounding bias	Population stratification	35	81.40	8	18.60
Multiple testing	Adjusted for multiple comparisons	42	97.67	1	2.33
Selective reporting	Selective reporting	32	74.42	11	25.58
HWE	Hardy-Weinberg equilibrium	36	83.72	7	16.28

### 3.4.1 Minor allele prevalence

Data used for pooling minor allele prevalence from 21 studies were described by ethnicity and type of subjects, see Table 3.5. Data for pooling minor allele prevalence was not available in 4 studies<sup>34, 35, 50, 57</sup>. HWE was checked for each study, for some studies where genotype data were not provided, we relied on assessing HWE from what was reported in the articles. There were 4 studies<sup>23, 24, 36, 37</sup> in children Caucasians, 1 study<sup>25</sup> reported for adult and children Caucasians, 7 studies<sup>12, 20, 22, 26, 27, 38, 73</sup> in adult Caucasians, 2 studies<sup>34, 53</sup> reported in both Caucasians and Asians, 1 study<sup>33</sup> in children Asians, and 6 studies<sup>21, 56, 58, 61, 64, 70</sup> in adult Asians. The pooled minor A allele prevalence were estimated in children Caucasians, adult Caucasians, children Asians and adult Asians. The pooled prevalence across ethnicity and type of subjects were not much different with corresponding pooled prevalence of 0.232 (95%CI: 0.190, 0.274), 0.287 (95%CI: 0.278, 0.296), 0.281 (95%CI: 0.158, 0.404), and 0.293 (95%CI: 0.275, 0.312), respectively.

### 3.4.2 Vitamin D level

The overall MDs for genotype data for AA versus CC (MD<sub>1</sub>) and AC versus CC (MD<sub>2</sub>) were described by ethnicity and type of subjects, see Table 3.6. There were 11 studies conducted in Caucasians. Among 4 studies<sup>23-25, 36</sup> in children, MD<sub>1</sub> was high heterogeneity ( $I^2 = 76\%$ ) and no heterogeneity for MD<sub>2</sub> ( $I^2 = 0\%$ ). The pooled MD<sub>1</sub> and MD<sub>2</sub> were respectively -3.705 (95%CI: -6.241, -1.169) and -2.211 (95%CI: -2.930, -1.491), which could be interpreted that subjects with AA and AC genotypes had 25(OH)D about 3.7 and 2.2 ng/ml significantly lower than subjects with CC genotype, see Figure 3.2 a-b. Publication bias for pooling MDs was assessed where at least 3 studies were used for pooling using a funnel plot, which graphed SE on y-axis against MD on x-axis. For children Caucasians, the funnel plot suggested a little deviated from the funnel for MD<sub>1</sub> but not for MD<sub>2</sub>, see Figure 3.3 a-b. This corresponded with the Egger's tests which yielded non-significant asymmetry of the funnels for both MD<sub>1</sub> and MD<sub>2</sub>, with (Coefficient = -0.555, SE = 8.599, P = 0.954) and (Coefficient = -1.976, SE = 1.919, P = 0.411) respectively.

Among 7<sup>12, 20, 22, 25-27, 53</sup> studies in adults, the MDs were not much heterogeneous with the estimated  $I^2$ s of 25% and 0% for MD<sub>1</sub> and MD<sub>2</sub>, respectively.

The corresponding pooled MDs were -3.619 (95%CI: -4.978, -2.261) and -1.601 (95%CI: -2.369, -0.833); which similar gene effects in children Caucasians i.e., adult Caucasians with AA and AC genotypes had 3.6 ng/ml and 1.6 ng/ml lower levels of vitamin D compared to those who have CC genotypes, see Figure 3.4 a-b. Publication bias was assessed for adult Caucasians, the funnel plots looked symmetry for MD<sub>1</sub> but not for MD<sub>2</sub>, see Figure 3.5 a-b. This was corresponded with the Egger's tests which suggested symmetry for MD<sub>1</sub> (Coefficient = -0.836, SE = 0.788, P = 0.337) but asymmetry for MD<sub>2</sub> and (Coefficient = -1.144, SE = 0.328, P = 0.018). A contour-enhanced funnel plot was performed for MD<sub>2</sub> (see Figure 3.5 c) which indicated that some studies were missing, which could be either high quality with negative results or low quality with positive results. Thus, publication bias might be present for this pooling.

Only 2 studies<sup>53, 61</sup> in adult Asians were available for pooling MDs, which were homogenous for both MDs ( $I^2 = 0\%$ ) with MD<sub>1</sub> and MD<sub>2</sub> of -4.042 (95%CI: (-5.621, -2.462) and, -1.722 (95%CI: -2.754, -0.69), respectively. There were no studies in children Asians.

Six studies<sup>21, 35, 56, 57, 70, 73</sup> reported the genotype effects using the additive model, 3 studies in adult Caucasians, and 3 studies in adult Asians. The pooled MDs were -1.932 (95%CI: -2.719, -1.146) and -0.03 (95%CI: -0.037, 0.022) in Caucasians and Asians, respectively. The gene effects were mild heterogeneous in adult Caucasians ( $I^2 = 4\%$ ) but very heterogeneous in adult Asians ( $I^2 = 91.9\%$ ), see Figure 3.6 a-b. This could be interpreted that for every one copy of A allele increased in adult Caucasians, the 25(OH)D level significantly decreased 1.93 ng/ml; but this was not replicated in Asians. For pooling additive effects, a funnel plot suggested symmetry of the funnel for adult Caucasians but not for adult Asians, see Figure 3.7 a-b. The Egger's tests were non-significant for both MDs with (Coefficient = 1.105, SE = 2.206, P = 0.704) and (Coefficient = 0.023, SE = 3.672, P = 0.996), respectively.

### 3.4.3 Vitamin D deficiency/insufficiency

Among 5 studies<sup>21, 33, 34, 37, 38</sup> assessed A vs C allele effect on vitamin D deficiency/insufficiency, 2<sup>37, 38</sup> and 2 studies<sup>21, 33</sup> were Caucasians and Asians, and 1 study<sup>34</sup> conducted in both ethnicities. Their data have been described in Table 3.7.

Pooling these ORs stratified by ethnicity found homogeneity ( $I^2 = 0\%$ ) for both Caucasians and Asians with the pooled OR of 1.522 (95%CI: 1.336, 1.733) and 1.255 (95%CI: 1.058, 1.487), respectively. This suggested that Caucasians and Asians, who had A allele were approximately at 52% and 26% significantly higher risk of developing vitamin D deficiency/insufficiency than those Caucasians and Asians who had C allele. Publication bias for pooling ORs was assessed. For pooling allele effect (A versus C) in Caucasians and Asians, funnel plots showed symmetry for both, respectively, see Figure 3.8 a-b. The Egger's tests were non-significant for both Caucasians and Asians, with (Coefficient = 1.300, SE = 0.414, P = 0.196) and (Coefficient = -0.807, SE = 0.455, P = 0.327) respectively.

Genotype data for AA versus CC ( $OR_1$ ) and AC versus CC ( $OR_2$ ) were described by ethnicity, see Table 3.8. Three studies each were conducted in Caucasians and Asians, respectively. In Caucasians, the genotype effects were homogenous for both ORs ( $I^2 = 0\%$ ) with the pooled  $OR_1$  and pooled  $OR_2$  2.207 (95%CI: 1.629, 2.99) and 1.529 (95%CI: 1.28, 1.826), respectively. This suggested that Caucasians with AA genotype and AC genotypes were approximately 2.21 and 1.53 times higher risk of developing vitamin D deficiency/insufficiency than those Caucasians with CC genotype. For publication bias, funnel plots also showed no evidence of asymmetry for both genotypic ORs in Caucasians, see Figure 3.9 a-b. This was corresponded with the Egger's tests, which suggested non-significant of asymmetry for both  $OR_1$  and  $OR_2$  with (Coefficient = 0.693, SE = 0.059, P = 0.054) and (Coefficient = -1.397, SE = 0.687, P = 0.291), respectively.

The genotype effects were also homogenous ( $I^2 = 0\%$  for both ORs) in Asians, with the pooled  $OR_1$  and pooled  $OR_2$  of 1.578 (95%CI: 1.06, 2.348) and 1.251 (95%CI: 0.991, 1.58), respectively. This suggested that Asians with AA and AC genotypes were approximately at 58% and 25% significantly higher risk of developing vitamin D deficiency/insufficiency than those Asians with CC genotype. There was no evidence of publication bias for pooling genotypic effects in Asians, see Figure 3.10 a-b. The Egger's tests were non-significant for both  $OR_1$  and  $OR_2$ , with (Coefficient = -0.862, SE = 0.377, P = 0.263) and (Coefficient = -0.325, SE = 0.272, P = 0.444) respectively.

**Table 3.5** Pooling the prevalence of minor A allele of rs4588

Author, Year, (Reference No.)	Ethnicity	Type of subjects	Total number of alleles	Minor A allele frequency	Prevalence of the minor allele (95% CI)	HWE
Carpenter, 2013 <sup>23</sup>	Caucasian	Children	1550	274	0.177 (0.158, 0.196)	0.847
Santos, 2013, <sup>37</sup>	Caucasian	Children	396	106	0.268 (0.224, 0.311)	0.511
Navas-Nazario, 2014 <sup>24</sup>	Caucasian	Children	668	141	0.211 (0.18, 0.242)	0.772
Pekkinen, 2014 <sup>36</sup>	Caucasian	Children	466	87	0.187 (0.151, 0.222)	0.035 <sup>a</sup>
Nissen, 2014 <sup>25</sup>	Caucasian	Children	688	184	0.267 (0.234, 0.301)	0.321
Suaini, 2014 <sup>34</sup>	Caucasian	Children	968	278	0.287 (0.259, 0.316)	0.065
Pooled			4340	964	0.232 (0.190, 0.274)	
Azad 27	Caucasian	Adult	912	281	0.308 (0.278, 0.338)	0.415
Janssens, 2010 <sup>20</sup>	Caucasian	Adult	300	75	0.25 (0.201, 0.299)	0.114
Perna, 2013 <sup>26</sup>	Caucasian	Adult	7482	2092	0.28 (0.269, 0.29)	0.6
Singh, 2013 <sup>22</sup>	Caucasian	Adult	356	100	0.281 (0.234, 0.328)	0.987
Sinotte, 2009 <sup>27</sup>	Caucasian	Adult	1466	430	0.293 (0.27, 0.317)	0.483
Touvier, 2014 <sup>38</sup>	Caucasian	Adult	3548	1071	0.302 (0.287, 0.317)	0.77
Gilbert, 2014 <sup>73</sup>	Caucasian	Adult	1796	508	0.283 (0.262, 0.304)	0.604
Gozdzik, 2011 <sup>53</sup>	Caucasian	Adult	222	62	0.279 (0.22, 0.338)	0.757
Nissen, 2014 <sup>25</sup>	Caucasian	Adult	828	229	0.277 (0.246, 0.307)	0.567
Pooled			16910	4848	0.287 (0.278, 0.296)	

**Table 3.5** Pooling the prevalence of minor A allele of rs4588 (continued)

Author, Year, (Reference No.)	Ethnicity	Type of subjects	Total number of alleles	Minor A allele frequency	Prevalence of the minor allele (95% CI)	HWE
Kitanaka, 2012 <sup>33</sup>	Asian	Children	132	29	0.22 (0.149, 0.29)	0.559
Suaini, 2014 <sup>34</sup>	Asian	Children	142	49	0.345 (0.267, 0.423)	0.741
Pooled			274	78	0.281 (0.158, 0.404)	
Li, 2014 <sup>21</sup>	Asian	Adult	2398	709	0.296 (0.277, 0.314)	0.94
Lu, 2012 <sup>56</sup>	Asian	Adult	6350	2008	0.316 (0.305, 0.328)	0.345
Nimitphong, 2013 <sup>58</sup>	Asian	Adult	78	23	0.295 (0.194, 0.396)	0.045 <sup>a</sup>
Robien, 2013 <sup>61</sup>	Asian	Adult	958	251	0.262 (0.234, 0.29)	0.148
Suzuki, 2012 <sup>64</sup>	Asian	Adult	274	68	0.248 (0.197, 0.299)	0.475
Zhang, 2013 <sup>70</sup>	Asian	Adult	5794	1848	0.319 (0.307, 0.331)	0.36
Gozdzik, 2011 <sup>53</sup>	Asian	Adult	398	107	0.269 (0.225, 0.312)	0.453
Pooled			16524	5092	0.293 (0.275, 0.312)	

<sup>a</sup> These 2 studies had Hardy Weinberg disequilibrium but was included in the analysis after a sensitivity analysis

CI; confidence interval, HWE; Hardy-Weinberg Equilibrium

**Table 3.6** Describes genotype data and pooled mean difference of vitamin D levels for rs4588

First Author, Year, (Reference No.)	CC				AC				AA		MD <sub>1</sub>	MD <sub>2</sub>
	n	mean	SD	n	mean	SD	n	mean	SD			
<b>Children Caucasians</b>												
Carpenter, 2013 <sup>23</sup>	526	27.16	9.62	224	25.12	4.41	25	24.4	6.01	6.01	-2.764 (-5.259, -0.268)	-2.043 (-3.047, -1.039)
Navas-Nazario, 2014 <sup>24</sup>	207	28.41	12.38	113	24.88	6.05	14	26.08	5.25	5.25	-2.33 (-5.556, 0.896)	-3.53 (-5.552, -1.508)
Nissen, 2014 <sup>25</sup>	181	30.57	6.88	142	28.08	6.33	21	23.12	5.25	5.25	-7.45 (-9.909, -4.991)	-2.49 (-3.935, -1.045)
Pekkinen, 2014 <sup>36</sup>	159	18.31	5.84	61	16.79	4.72	13	16.26	3.5	3.5	-2.05 (-4.158, 0.058)	-1.52 (-3.012, -0.028)
<b>Pooled MD</b>											-3.705 (-6.241, -1.169)	-2.211 (-2.930, -1.491)
<b>Adult Caucasians</b>												
Azad, 2012 <sup>12</sup>	222	26.88	8.81	187	24.59	8.01	47	21.79	8.41	8.41	-5.09 (-7.759, -2.421)	-2.29 (-3.921, -0.659)
Gozdzik, 2011 <sup>53</sup>	57	26.82	9.38	46	25.22	8.49	8	22.66	7.49	7.49	-4.16 (-9.893, 1.573)	-1.6 (-5.057, 1.857)
Janssens, 2010 <sup>20</sup>	88	25.80	8.80	49	23.8	8.6	13	20.6	4.6	4.6	-5.2 (-8.304, -2.096)	-2 (-5.03, 1.03)

**Table 3.6** Describes genotype data and pooled mean difference of vitamin D levels for rs4588 (continued)

First Author, Year, (Reference No.)	CC				AC				AA		MD <sub>1</sub>	MD <sub>2</sub>
	n	mean	SD	n	mean	SD	n	mean	SD			
Nissen, 2014 <sup>25</sup>	219	29.69	9.68	161	27.92	9.08	34	25.48	9.78	-4.21	(-7.739, -0.681)	(-3.67, 0.13)
Perna, 2013 <sup>26</sup>										-2.20	(-3.298, -1.102)	(-1.469, -0.371)
Singh, 2013 <sup>22</sup>	92	26.56	11.26	72	23.84	10.3	14	29.41	21	2.85	(-8.363, 14.063)	(-6.03, 0.59)
Sinotte, 2009 <sup>27</sup>	370	26.92	7.70	296	25.32	7.58	67	23.64	7.87	-3.28	(-5.321, -1.239)	(-2.767, -0.433)
<b>Pooled MD</b>										<b>-3.619</b>	<b>(-4.978, -2.261)</b>	<b>(-2.369, -0.833)</b>
<b>Adult Asians</b>												
Gozdzik, 2011 <sup>53</sup>	106	15.62	6.55	79	13.96	5.11	14	12.37	4.04	-3.25	(-5.706, -0.794)	(-3.341, 0.021)
Robien, 2013 <sup>61</sup>	267	27.52	7.01	173	25.76	6.72	39	22.92	6	-4.6	(-6.662, -2.538)	(-3.068, -0.452)
<b>Pooled MD</b>										<b>-4.042</b>	<b>(-5.621, -2.462)</b>	<b>(-2.754, -0.69)</b>

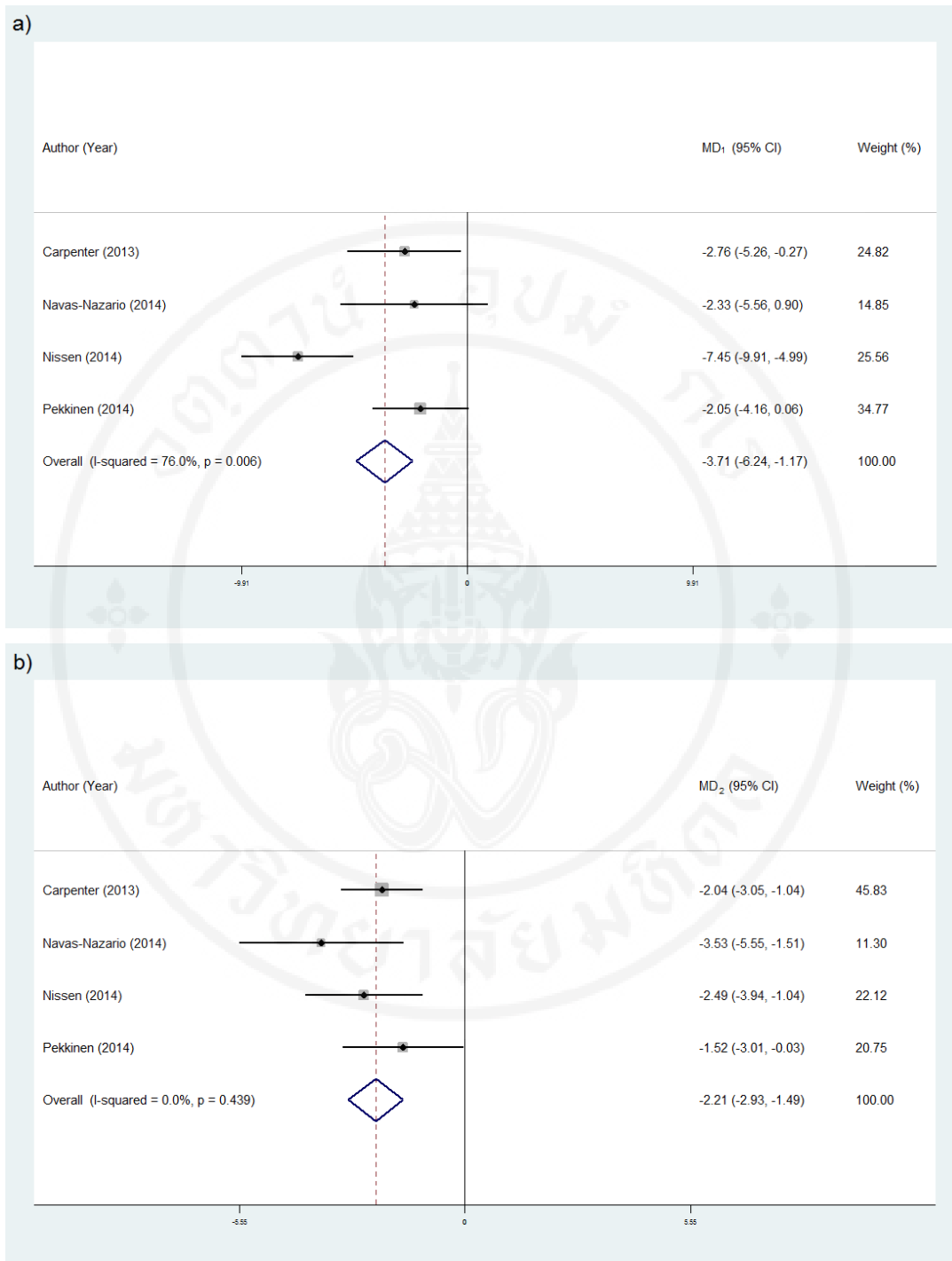
**Table 3.6** Describes genotype data and pooled mean difference of vitamin D levels for rs4588 (continued)

First Author, Year, (Reference No.)	CC			AC			AA			MD <sub>1</sub>	MD <sub>2</sub>
	n	mean	SD	n	mean	SD	n	mean	SD		
Additive model											
Adult Caucasians											
Bu, 2010 <sup>35</sup>										-0.693 (-2.575, 1.188)	
Muindi, 2012 <sup>57</sup>										-2.43 (-4.429, -0.431)	
Gilbert, 2014 <sup>73</sup>										-2.14 (-3.1, -1.18)	
Pooled MD											
Adult Asians											
Lu, 2012 <sup>56</sup>										-0.031 (-0.038, -0.023)	
Zhang, 2013 <sup>70</sup>										0.163 (0.06, 0.267)	

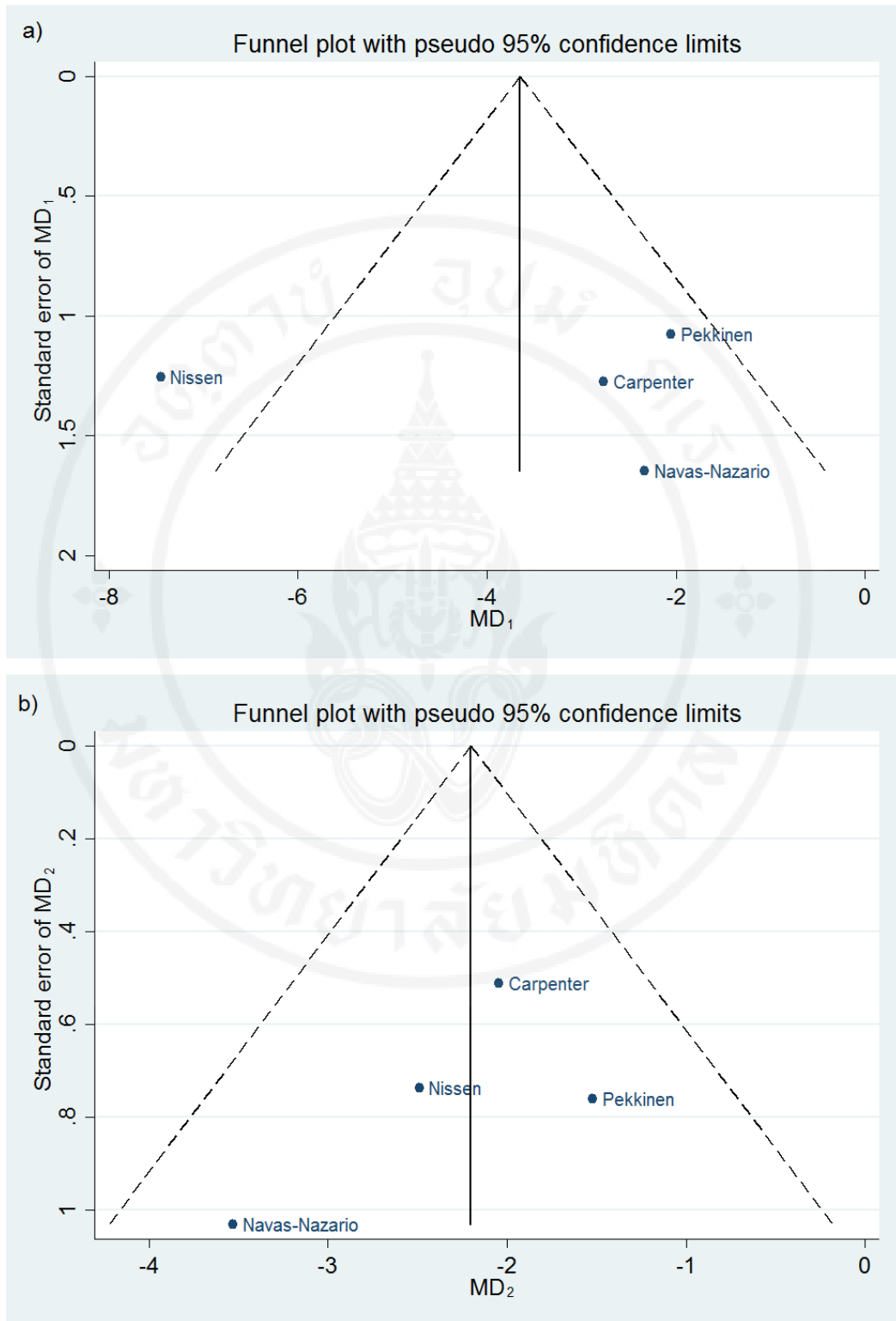
**Table 3.6** Describes genotype data and pooled mean difference of vitamin D levels for rs4588 (continued)

First Author, Year, (Reference No.)	CC			AC			AA			MD <sub>1</sub>	MD <sub>2</sub>
	n	mean	SD	n	mean	SD	n	mean	SD		
Li, 2014 <sup>21</sup>										-1.24 (-1.946, -0.534)	
Pooled MD										-0.03 (-0.037, 0.022)	

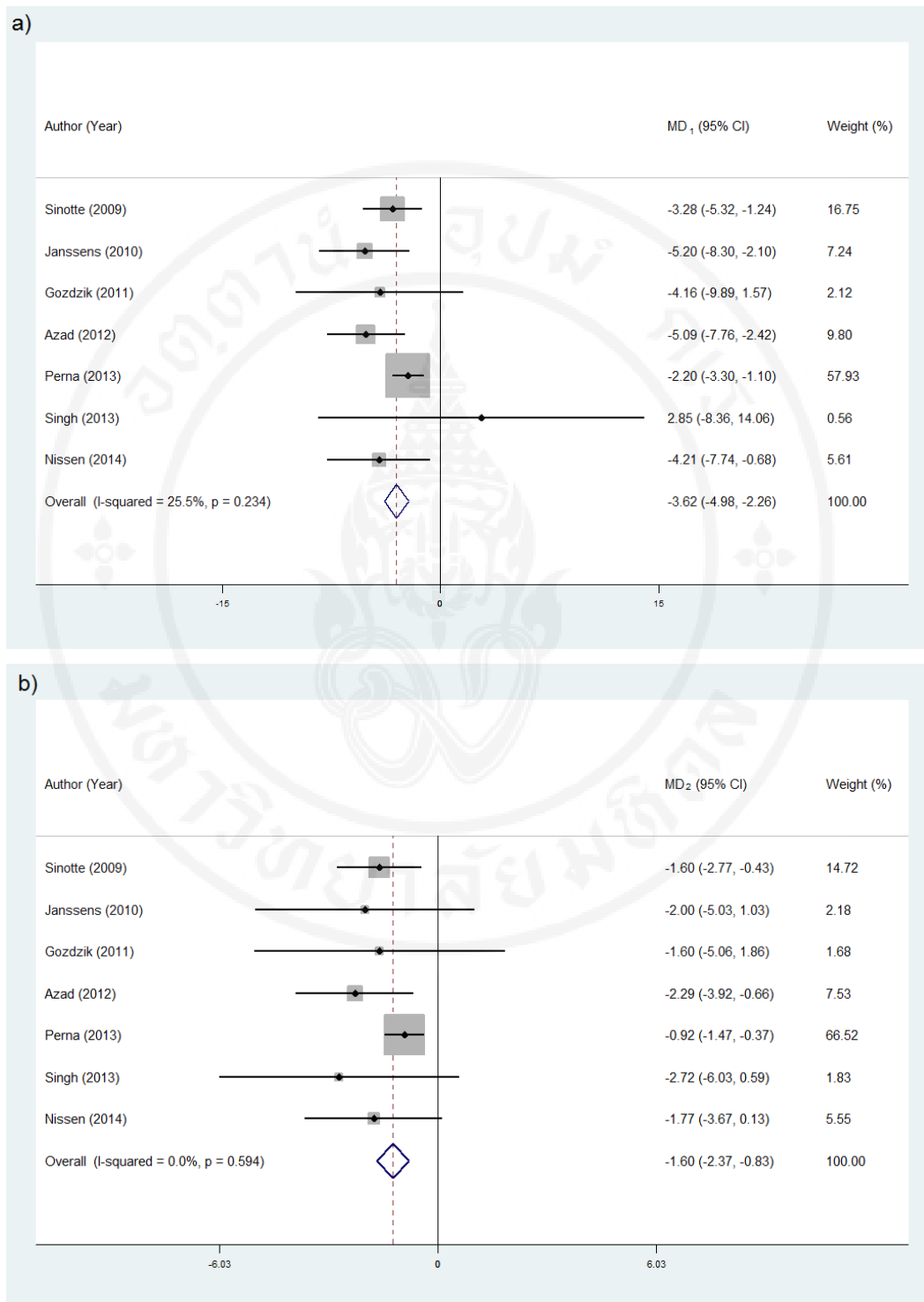
MD<sub>1</sub>; mean difference of AA versus CC, MD<sub>2</sub>; mean difference of AC versus CC, n; number of subjects, SD; standard deviation



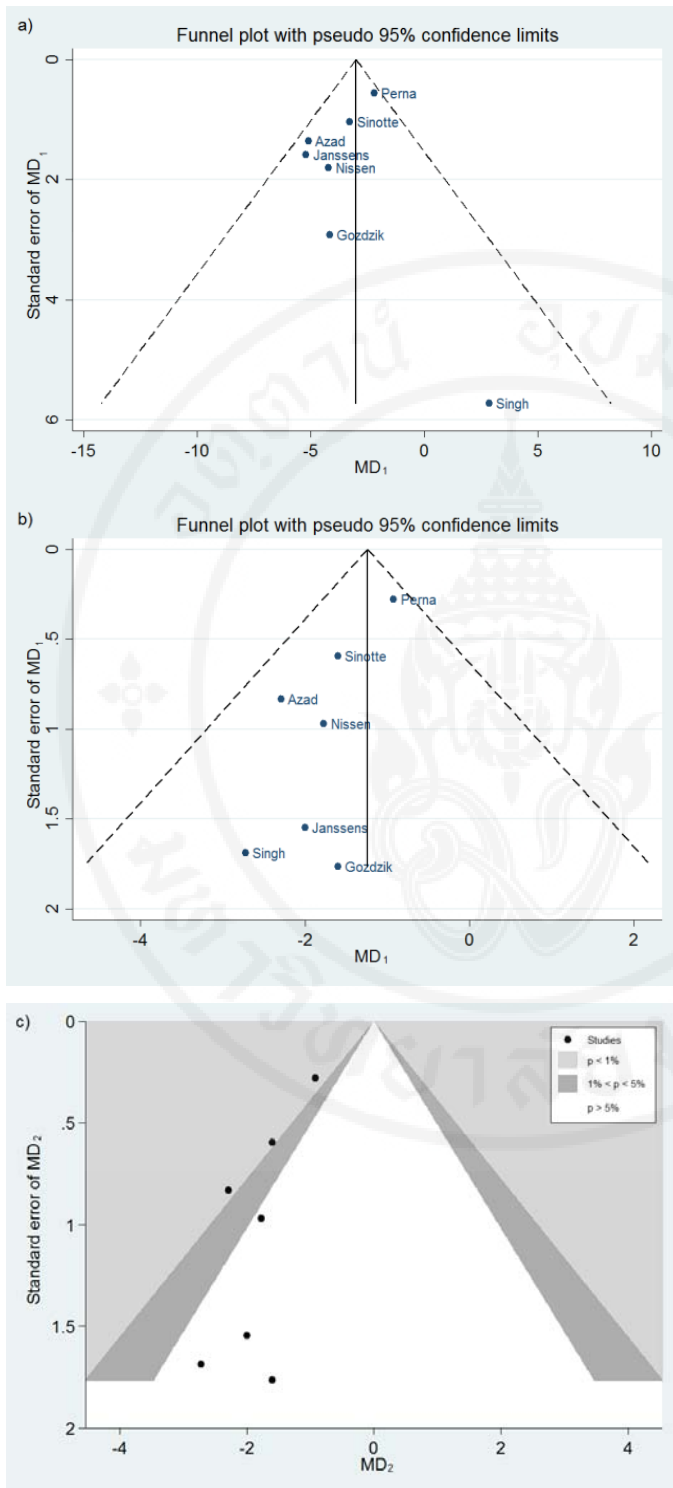
**Figure 3.2** Forest plot of pooled MD in children Caucasians for rs4588 a) MD<sub>1</sub> for AA versus CC, b) MD<sub>2</sub> for AC versus CC



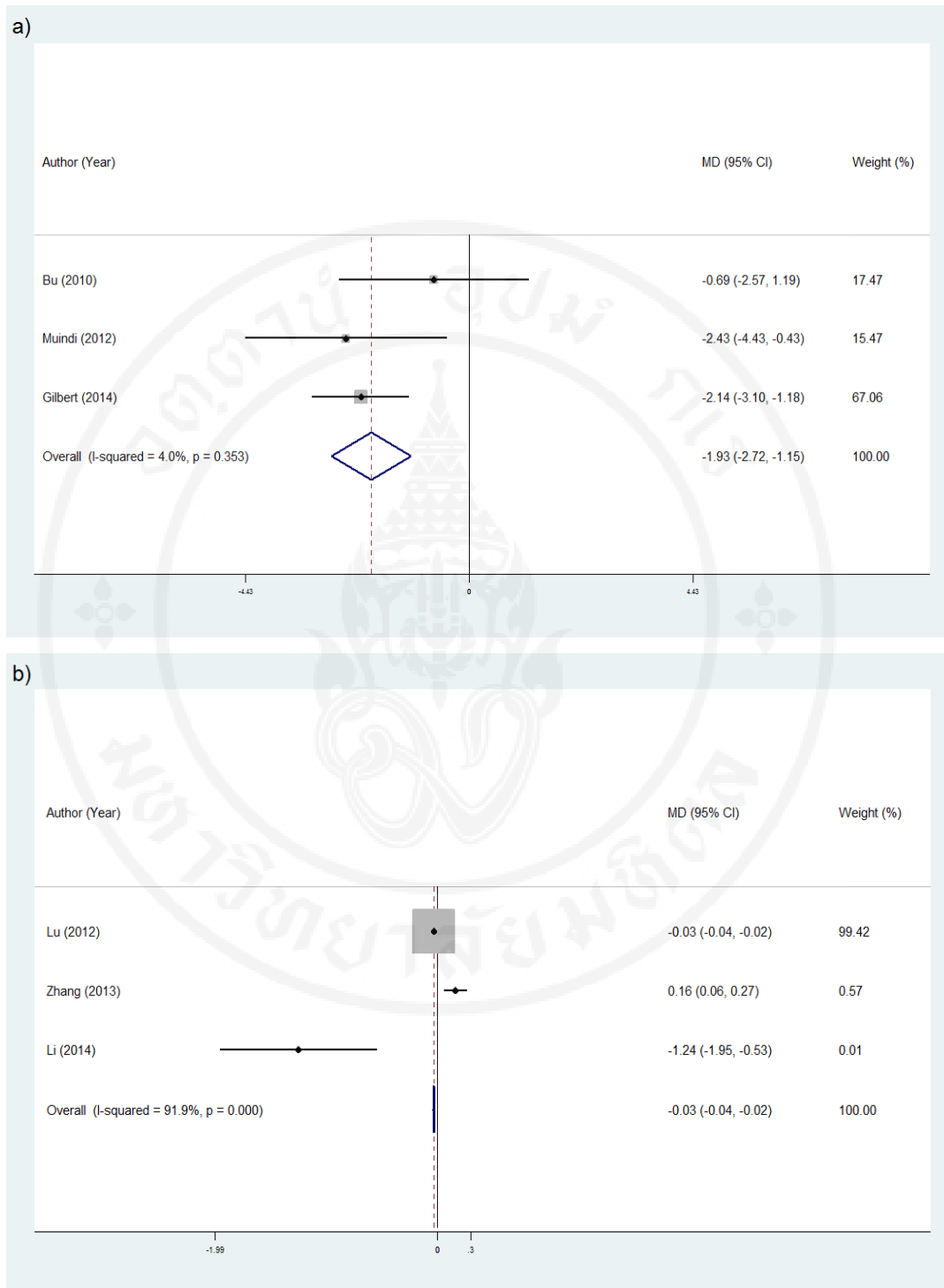
**Figure 3.3** Funnel plots for pooling MD<sub>1</sub> and MD<sub>2</sub> for rs4588 in children Caucasian studies a) MD<sub>1</sub> (AA versus CC), b) MD<sub>2</sub> (AC versus CC)



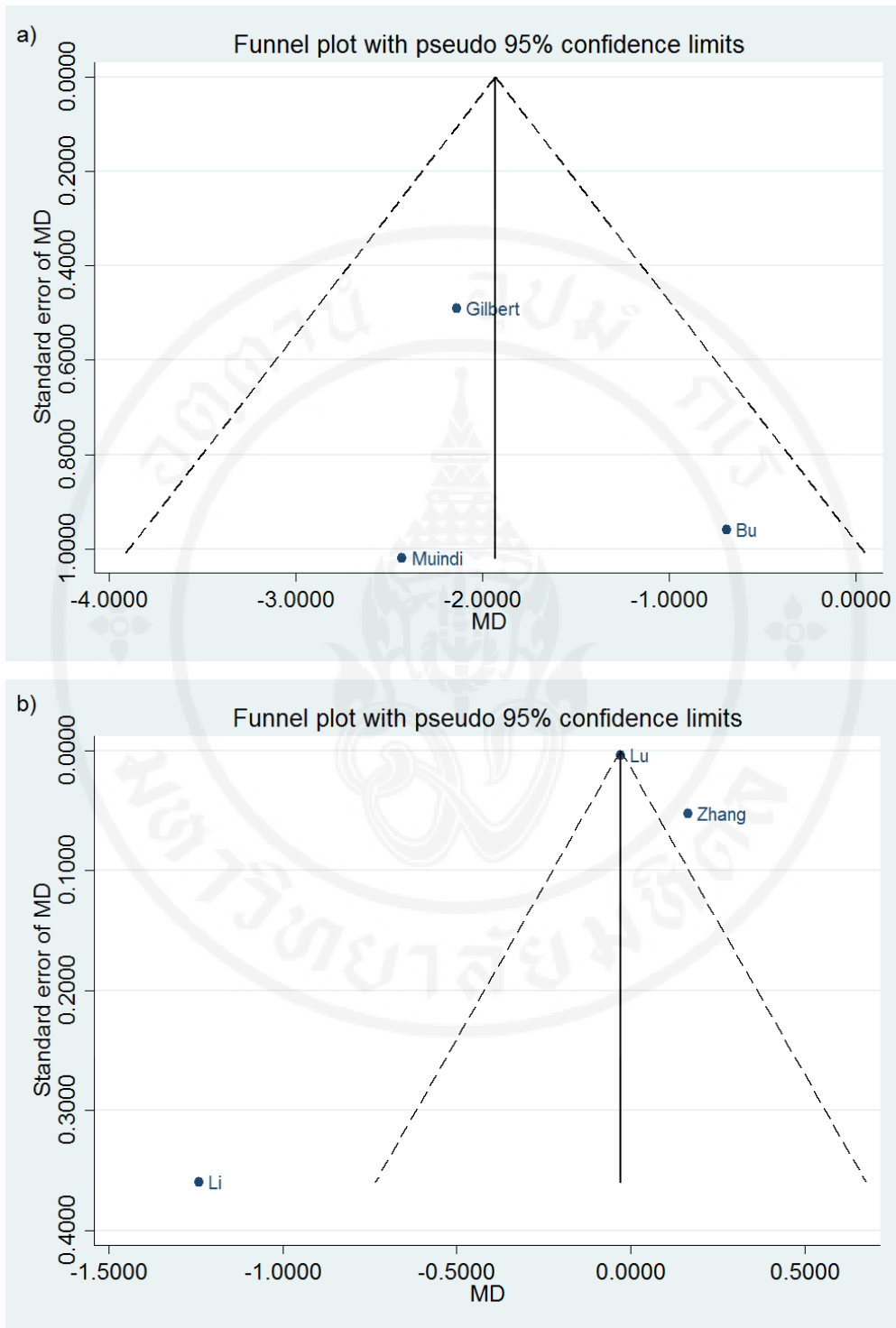
**Figure 3.4.** Forest plot of pooled MD in adult Caucasians for rs4588 a) MD<sub>1</sub> for AA versus CC, b) MD<sub>2</sub> for AC versus CC



**Figure 3.5** Funnel plots for pooling MD<sub>1</sub> and MD<sub>2</sub> for rs4588 in adult Caucasians studies a) MD<sub>1</sub> (AA versus CC), b) MD<sub>2</sub> (AC versus CC), c) Contour enhanced funnel plot for MD<sub>2</sub>



**Figure 3.6** Forest plot of pooling MDs as additive effect for rs4588 a) adult Caucasians, b) adult Asians

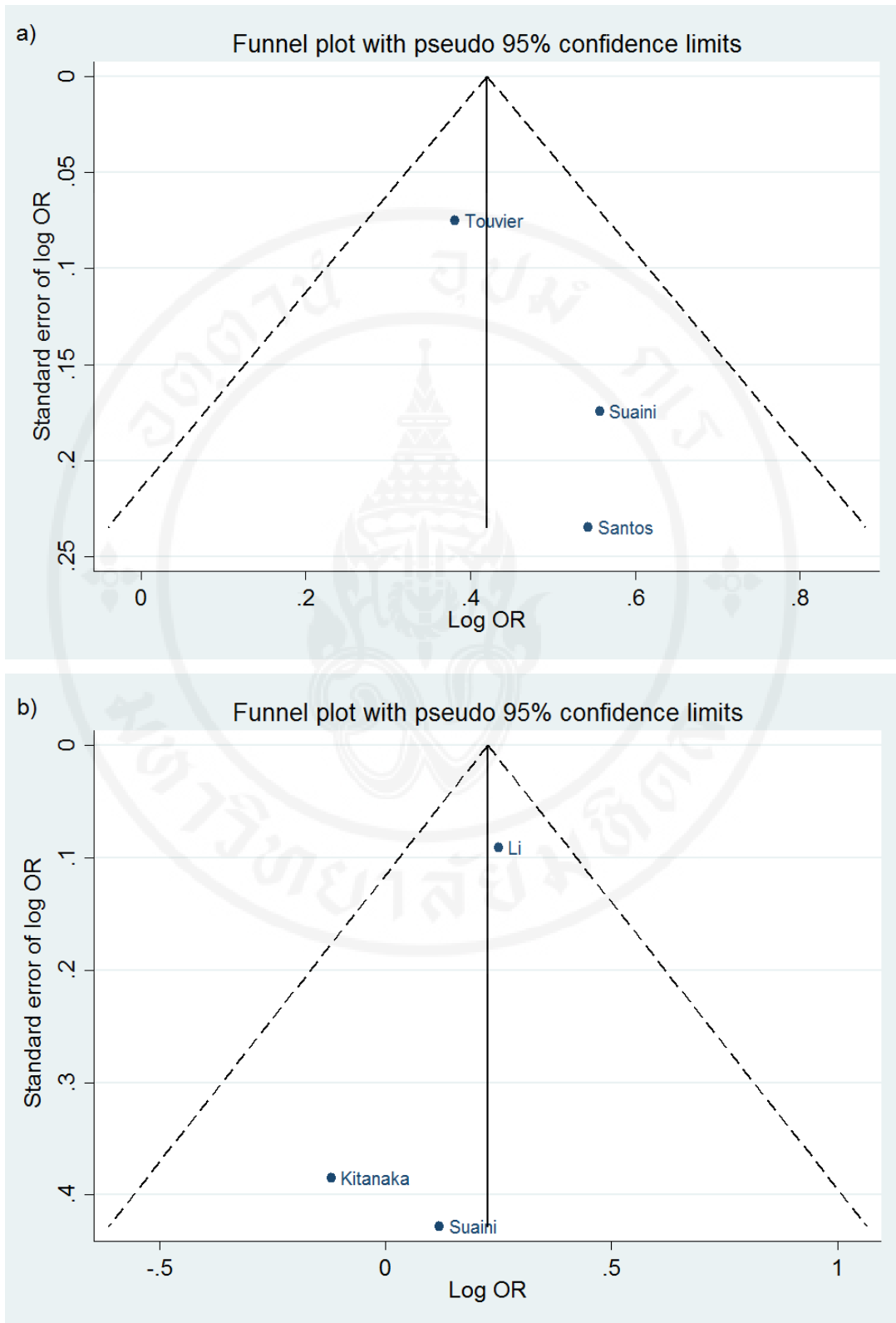


**Figure 3.7** Funnel plot of pooling MDs as additive effect for rs4588 a) adult Caucasians, b) adult Asians

**Table 3.7** Pooling allele effect of rs4588 on vitamin D deficiency/insufficiency

First Author, Year, (Reference No.)	Case		Control		OR (95% CI)
	C	A	C	A	
Caucasian					
Santos, 2013 <sup>37</sup>	87	45	203	61	1.721 (1.087, 2.726)
Touvier, 2014 <sup>38</sup>	1357	685	1120	386	1.465 (1.264, 1.698)
Suaini, 2014 <sup>34</sup>	108	68	582	210	1.745 (1.24, 2.456)
Pooled	1552	798	1905	657	1.522 (1.336, 1.733)
Asians					
Kitanaka, 2012 <sup>33</sup>	48	12	103	29	0.888 (0.417, 1.889)
Li, 2014 <sup>21</sup>	625	305	1064	404	1.285 (1.075, 1.536)
Suaini, 2014 <sup>34</sup>	88	118	488	416	1.127 (0.487, 2.61)
Pooled	761	435	1655	849	1.255 (1.058, 1.487)

CI; confidence interval, OR; odds ratio



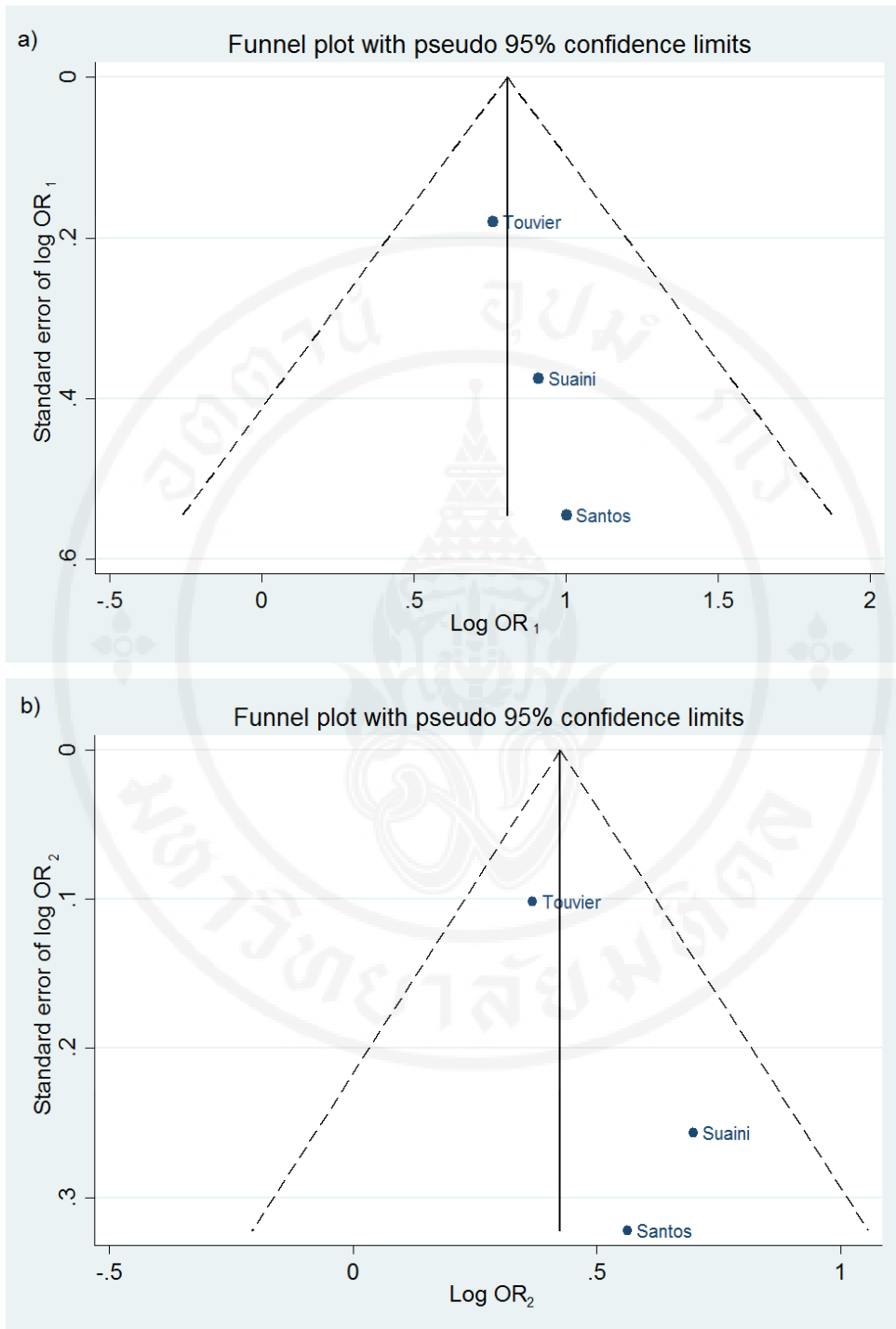
**Figure 3.8** Funnel plots of rs4588 allele effect on vitamin D deficiency/insufficiency

a) Log OR (A versus C) in Caucasians, b) Log OR (A versus C) in Asians

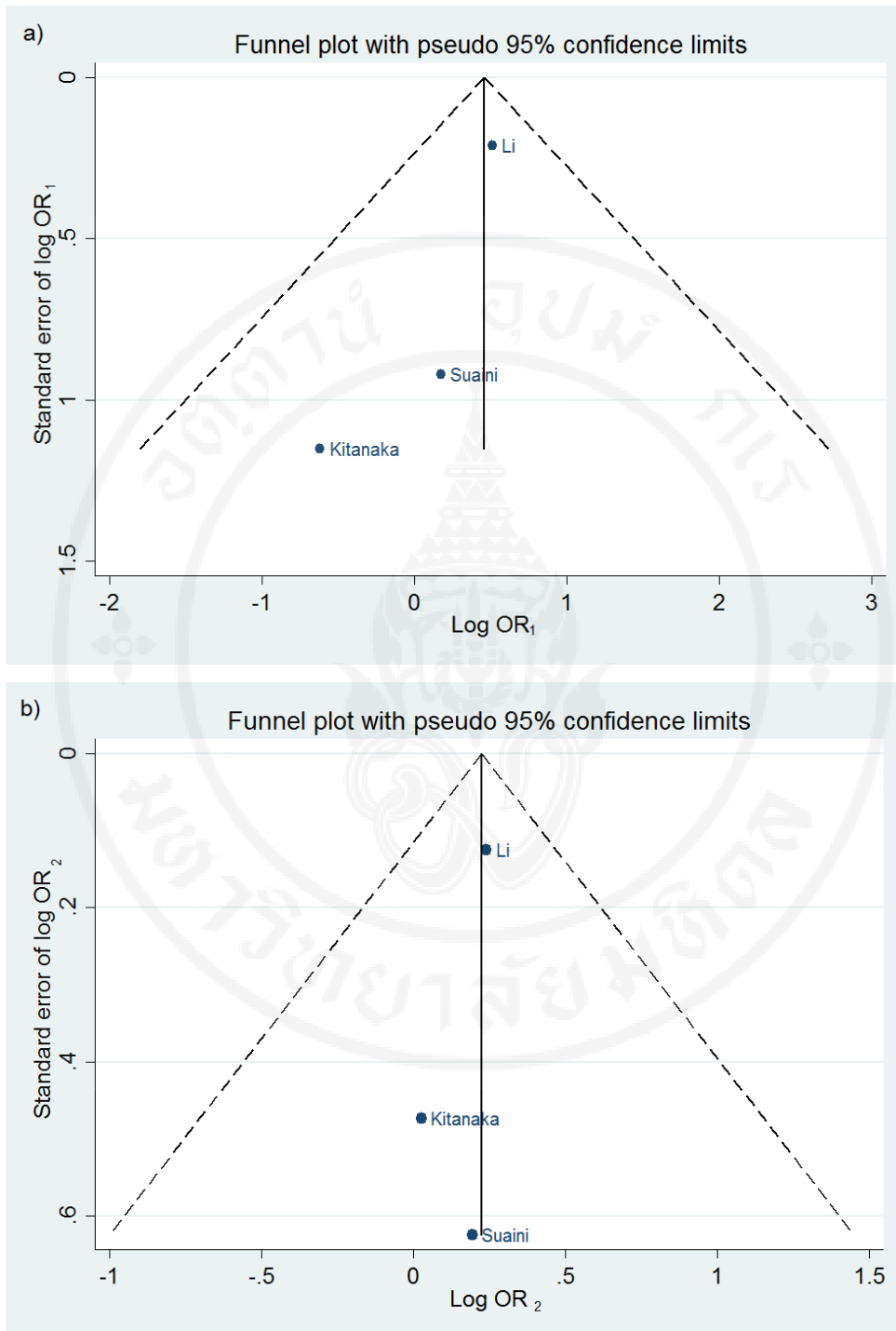
**Table 3.8** Pooling genotype effect of rs4588 on vitamin D deficiency/insufficiency

First Author, Year, (Reference No.)	Case				Control				OR <sub>1</sub> (95% CI)	OR <sub>2</sub> (95% CI)
	CC		AA		CC		AA			
	CC	AC	AA	AC	CC	AC	AA			
<b>Caucasian</b>										
Santos, 2013 <sup>37</sup>	29	29	8	45	79	45	8	2.724 (0.936, 7.929)	1.756 (0.933, 3.302)	
Touvier, 2014 <sup>38</sup>	455	447	119	284	418	284	51	2.144 (1.505, 3.054)	1.446 (1.185, 1.765)	
Suaini, 2014 <sup>34</sup>	33	42	13	140	221	140	35	2.487 (1.194, 5.183)	2.009 (1.215, 3.321)	
<b>Pooled</b>	<b>517</b>	<b>518</b>	<b>140</b>	<b>469</b>	<b>718</b>	<b>469</b>	<b>94</b>	<b>2.207 (1.629, 2.99)</b>	<b>1.529 (1.28, 1.826)</b>	
<b>Asian</b>										
Kitanaka, 2012 <sup>33</sup>	19	10	1	21	41	21	4	0.539 (0.056, 5.159)	1.028 (0.406, 2.602)	
Li, 2014 <sup>21</sup>	211	203	51	292	386	292	56	1.666 (1.1, 2.523)	1.272 (0.995, 1.626)	
Suaini, 2014 <sup>34</sup>	6	7	2	24	25	24	7	1.19 (0.196, 7.249)	1.215 (0.357, 4.141)	
<b>Pooled</b>	<b>236</b>	<b>220</b>	<b>54</b>	<b>337</b>	<b>452</b>	<b>337</b>	<b>67</b>	<b>1.578 (1.06, 2.348)</b>	<b>1.251 (0.991, 1.58)</b>	

CI; confidence interval, OR<sub>1</sub>; odds ratio of AA versus CC, OR<sub>2</sub>; odds ratio of AC versus CC



**Figure 3.9** Funnel plots of rs4588 genotype effect on vitamin D deficiency/insufficiency in Caucasians a) Log OR<sub>1</sub> (AA versus CC), b) Log OR<sub>2</sub> (AC versus CC)



**Figure 3.10** Funnel plots of rs4588 genotype effect on vitamin D deficiency/insufficiency in Asians a) Log OR<sub>1</sub> (AA versus CC), b) Log OR<sub>2</sub> (AC versus CC)

### 3.5 SNP rs7041

Twenty studies<sup>20-22, 24, 27, 29, 33-35, 37, 38, 54, 56, 59-61, 63, 64, 68, 73</sup> were included in the pooling for rs7041, 4 studies<sup>33, 34, 37, 38</sup> had the outcome of interests as vitamin D deficiency/insufficiency, 15 studies<sup>20, 22, 24, 27, 29, 35, 54, 56, 59-61, 63, 64, 68, 73</sup> had the outcome of interests as vitamin D level, and 1 study<sup>21</sup> reported for both data. Among these 16 studies, 7 studies<sup>20, 22, 24, 27, 29, 61, 68</sup> reported MDs among genotypes, 6 studies<sup>21, 35, 54, 56, 63, 73</sup> reported additive effects of minor genotype, 1 study<sup>59</sup> reported additive effect of major genotype, 1 study<sup>64</sup> had combined effect of TG and TT, and 1<sup>60</sup> study had MD for allele data.

#### 3.5.1 Minor allele prevalence

The data for minor allele prevalence from 14 studies<sup>20-22, 24, 27, 29, 33, 34, 37, 38, 56, 61, 68, 73</sup> were described and for 6 studies<sup>35, 54, 59, 60, 63, 64</sup>, the data was not available, see Table 3.9. HWE was checked for each study, and for studies where genotype data were not provided, we relied on assessing HWE from what was reported in the articles. Among 10 Caucasian studies, 3 studies<sup>24, 34, 37</sup> were conducted in children and 7 studies<sup>20, 22, 27, 29, 38, 68, 73</sup> were conducted in adults. The pooled minor allele T prevalence of these corresponding groups were respectively 0.442 (95%CI: 0.401, 0.483) and 0.436 (95%CI: 0.353, 0.519). Among 5 Asian studies, 2<sup>33, 34</sup> and 3 studies<sup>21, 56, 61</sup> were conducted in children and adults, with the pooled minor allele prevalence of 0.506 (95%CI: 0.049, 0.964) and 0.287 (95%CI: 0.258, 0.316), respectively.

#### 3.5.2 Vitamin D level.

The MDs of 25(OH)D for TT versus GG (MD<sub>1</sub>) and TG versus GG (MD<sub>2</sub>) were described by ethnicity, see Table 3.10. There was only 1 Asian study<sup>61</sup>, which was not included after a sensitivity analysis. Among 6 Caucasian studies, 5<sup>20, 22, 27, 29, 68</sup> and 1<sup>24</sup> studies were adults and children, respectively. There were not enough studies to pool by type of subject in Caucasians. Also the pooled prevalence among children and adult Caucasians were not much different, therefore, the genotype effects were combined regardless type of subjects. The MDs were high heterogeneous for MD<sub>1</sub> ( $I^2 = 71.4\%$ ) but not for MD<sub>2</sub> ( $I^2 = 0\%$ ); with the corresponding pooled MDs of -

2.707 (95%CI: -4.91, -0.504) and -1.407 (95%CI: -2.203, -0.612), see Figure 3.11 a-b. This suggested that Caucasians with TT genotype and TG genotype had 2.71 ng/ml and 1.41 ng/ml lower level of vitamin D than those who have GG genotype, respectively. Publication bias was assessed for pooling MD<sub>1</sub> and MD<sub>2</sub>, which showed little asymmetry for MD<sub>1</sub> but symmetry for MD<sub>2</sub>, see Figure 3.12. The Egger's tests were non-significant for both MD<sub>1</sub> and MD<sub>2</sub>, with (Coefficient = 0.459, SE = 2.008, P = 0.831) and (Coefficient = -0.373, SE = 0.946, P = 0.713) respectively.

Six studies<sup>21, 35, 54, 56, 63, 73</sup> reported additive effects of minor genotype, 1 study<sup>59</sup> reported additive effect of major genotype. Among 6 studies reported additive effects, only 4 studies<sup>35, 54, 63, 73</sup> and 2 studies<sup>21, 56</sup> were conducted in Caucasians and Asians, with the pooled MDs of -0.023 (95%CI: (-0.031, -0.015) and, -0.015 (95%CI: -0.023, -0.008). Both poolings were high heterogeneities of ( $I^2 = 86.4\%$ ) and ( $I^2 = 93.4\%$ ), respectively.

### 3.5.3 Vitamin D deficiency/insufficiency

A total of 5 studies that assessed T vs G allele effect on vitamin D deficiency/insufficiency, 2<sup>37, 38</sup>, 2<sup>21, 33</sup>, and 1<sup>34</sup> studies were conducted in Caucasians, Asians, and both ethnicities, respectively. Their data were described in Table 3.11. Pooling these ORs stratified by ethnicity yielded the pooled ORs for Caucasians and Asians of 0.854 (95%CI: 0.487, 1.499) and 0.79 (95%CI: 0.662, 0.943), with high heterogeneity ( $I^2 = 91.3\%$ ) and homogeneity ( $I^2 = 0\%$ ), respectively, see Figure 3.13. This suggested that Caucasians and Asians, who have T allele, were approximately at 15% and 21% lower risk of developing vitamin D deficiency/insufficiency than those Caucasians and Asians who have G allele. For publication bias, funnel plots indicated asymmetry of allele-effect in Caucasian but not for in Asian, see Figure 3.14. However, the Egger's tests were non-significant for both Caucasians (Coefficient = 1.903, SE = 5.637, P = 0.793) and Asians (Coefficient = 0.699, SE = 0.503, P = 0.397).

Genotype data for TT versus GG (OR<sub>1</sub>) and TG versus GG (OR<sub>2</sub>) were described by ethnicity and type of subjects, i.e., 3<sup>34, 37, 38</sup> and 3 studies<sup>21, 33, 34</sup> were Caucasian and Asians, see Table 3.12. The ORs of Caucasians were OR<sub>1</sub> and OR<sub>2</sub>, 0.761 (95%CI: 0.249, 2.329) and 0.963 (95%CI: 0.509, 1.822) with high heterogeneity

( $I^2 = 90.5\%$  and  $78.5\%$ , respectively), see Figure 3.15 a-b. The funnel plots showed asymmetry for both ORs in Caucasians (see Figure 3.16 a-b) but the Egger's tests were non-significant for both ORs, i.e., Coefficient = 1.871 (SE = 5.166, P = 0.779) and 1.344 (SE = 3.856, P = 0.787) respectively. The ORs of Asians were both homogenous

( $I^2 = 0\%$ ) with OR<sub>1</sub> and OR<sub>2</sub> of 0.675 (95%CI: 0.452, 1.007) and 0.746 (95%CI: 0.589, 0.946), respectively, see Figure 3.17 a-b. The funnel plots also suggested a little deviation from asymmetry for both ORs in Asians, see Figure 3.18 a-b. The Egger's tests were non-significant for both OR<sub>1</sub> (coefficient = 0.375, SE = 0.674, P = 0.677) and OR<sub>2</sub> (coefficient = 0.802, SE = 0.254, P = 0.196), respectively.

**Table 3.9** Pooling the prevalence of the minor T allele of rs7041

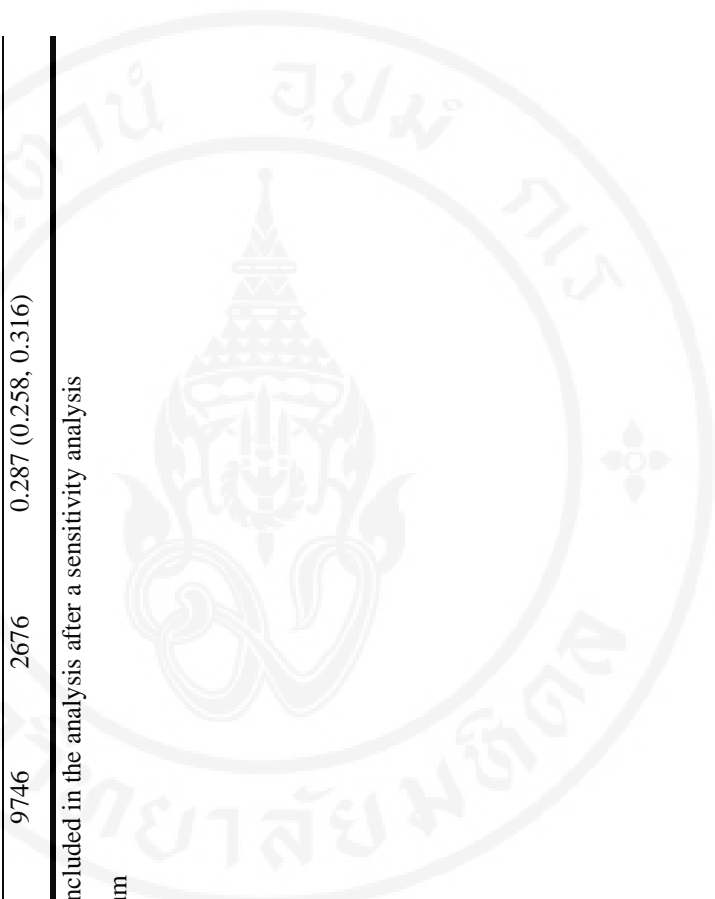
Author, Year, (Reference No.)	Ethnicity	Type of subjects	Total number of alleles	Minor T allele frequency	Prevalence of the minor allele (95% CI)	HWE
Navas-Nazario, 2014 <sup>24</sup>	Caucasian	Children	668	270	0.404 (0.367, 0.441)	0.086
Santos, 2013 <sup>37</sup>	Caucasian	Children	396	192	0.485 (0.436, 0.534)	0.326
Suaini, 2014 <sup>34</sup>	Caucasian	Children	968	429	0.443 (0.412, 0.474)	0.454
Pooled			2032	891	0.442 (0.401, 0.483)	
Gilbert, 2014 <sup>73</sup>	Caucasian	Adult	3750	1097	0.293 (0.278, 0.307)	0.287
Janssens, 2010 <sup>20</sup>	Caucasian	Adult	300	119	0.397 (0.341, 0.452)	0.837
Laurssen, 2014 <sup>29</sup>	Caucasian	Adult	2982	1259	0.422 (0.404, 0.44)	0.851
Singh, 2013 <sup>22</sup>	Caucasian	Adult	354	166	0.469 (0.417, 0.521)	0.074
Sinotte, 2009 <sup>27</sup>	Caucasian	Adult	1466	633	0.432 (0.406, 0.457)	0.192
Touvier, 2014 <sup>38</sup>	Caucasian	Adult	3568	1985	0.556 (0.54, 0.573)	0.071
Wang, 2014 <sup>68</sup>	Caucasian	Adult	1746	843	0.483 (0.459, 0.506)	0.001 <sup>a</sup>
Pooled			14166	6102	0.436 (0.353, 0.519)	
Kitanaka, 2012 <sup>33</sup>	Asian	Children	132	36	0.273 (0.197, 0.349)	0.573
Suaini, 2014 <sup>34</sup>	Asian	Children	142	105	0.739 (0.667, 0.812)	0.613
Pooled			274	141	0.506 (0.049, 0.964)	
Li, 2014 <sup>21</sup>	Asian	Adult	2398	675	0.281 (0.263, 0.299)	0.063
Lu, 2012 <sup>56</sup>	Asian	Adult	6366	1683	0.264 (0.254, 0.275)	0.218

**Table 3.9** Pooling the prevalence of the minor T allele of rs7041 (continued)

Author, Year, (Reference No.)	Ethnicity	Type of subjects	Total number of alleles	Minor T allele frequency	Prevalence of the minor allele (95% CI)	HWE
Robien, 2013 <sup>61</sup>	Asian	Adult	982	318	0.324 (0.295, 0.353)	0.755
Pooled			9746	2676	0.287 (0.258, 0.316)	

<sup>a</sup> This study had Hardy-Weinberg disequilibrium but was included in the analysis after a sensitivity analysis

CI; confidence interval, HWE; Hardy-Weinberg Equilibrium



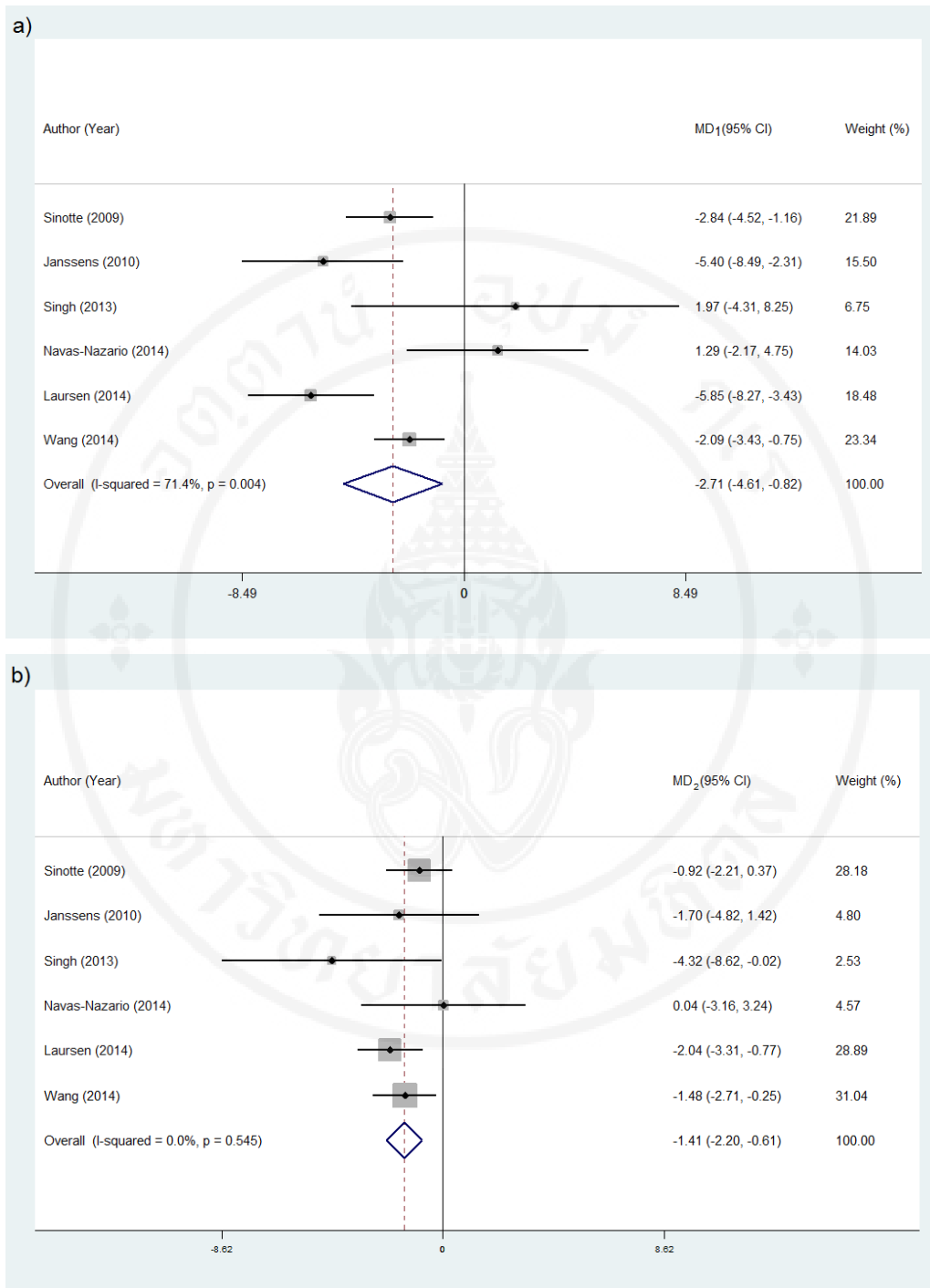
**Table 3.10** Describes genotype data and pooled mean difference of vitamin D levels for rs7041

First Author, Year, (Reference No.)	GG			TG			TT		MD <sub>1</sub>	MD <sub>2</sub>	
	n	mean	SD	n	mean	SD	n	mean			SD
Caucasians											
Sinotte, 2009 <sup>27</sup>	228	26.96	7.86	377	26.04	7.78	128	24.12	7.7	-2.84 (-4.519, -1.161)	-0.92 (-2.207, 0.367)
Janssens, 2010 <sup>20</sup>	54	26.3	8.7	73	24.6	9.1	23	20.9	5	-5.4 (-8.492, -2.308)	-1.7 (-4.821, 1.421)
Singh, 2013 <sup>22</sup>	44	27.4	12.94	100	23.08	10.02	33	29.37	14.62	1.97 (-4.315, 8.255)	-4.32 (-8.618, -0.022)
Navas-Nazario, 2014 <sup>24</sup>	111	26.92	16.59	176	26.96	5.69	47	28.21	5.49	1.29 (-2.172, 4.752)	0.04 (-3.159, 3.239)
Laursen, 2014 <sup>29</sup>	496	33.57	9.96	731	31.53	12.71	264	27.72	18.66	-5.85 (-8.266, -3.434)	-2.04 (-3.312, -0.768)
Wang, 2014 <sup>68</sup>	258	24.12	7.78	387	22.64	7.8	228	22.03	7.32	-2.09 (-3.433, -0.747)	-1.48 (-2.707, -0.253)
<b>Pooled MD</b>										<b>-2.707</b>	<b>-1.407</b>
Additive model											
Caucasians											
Bu, 2010 <sup>35</sup>										1.18	(-0.643, 3.003)

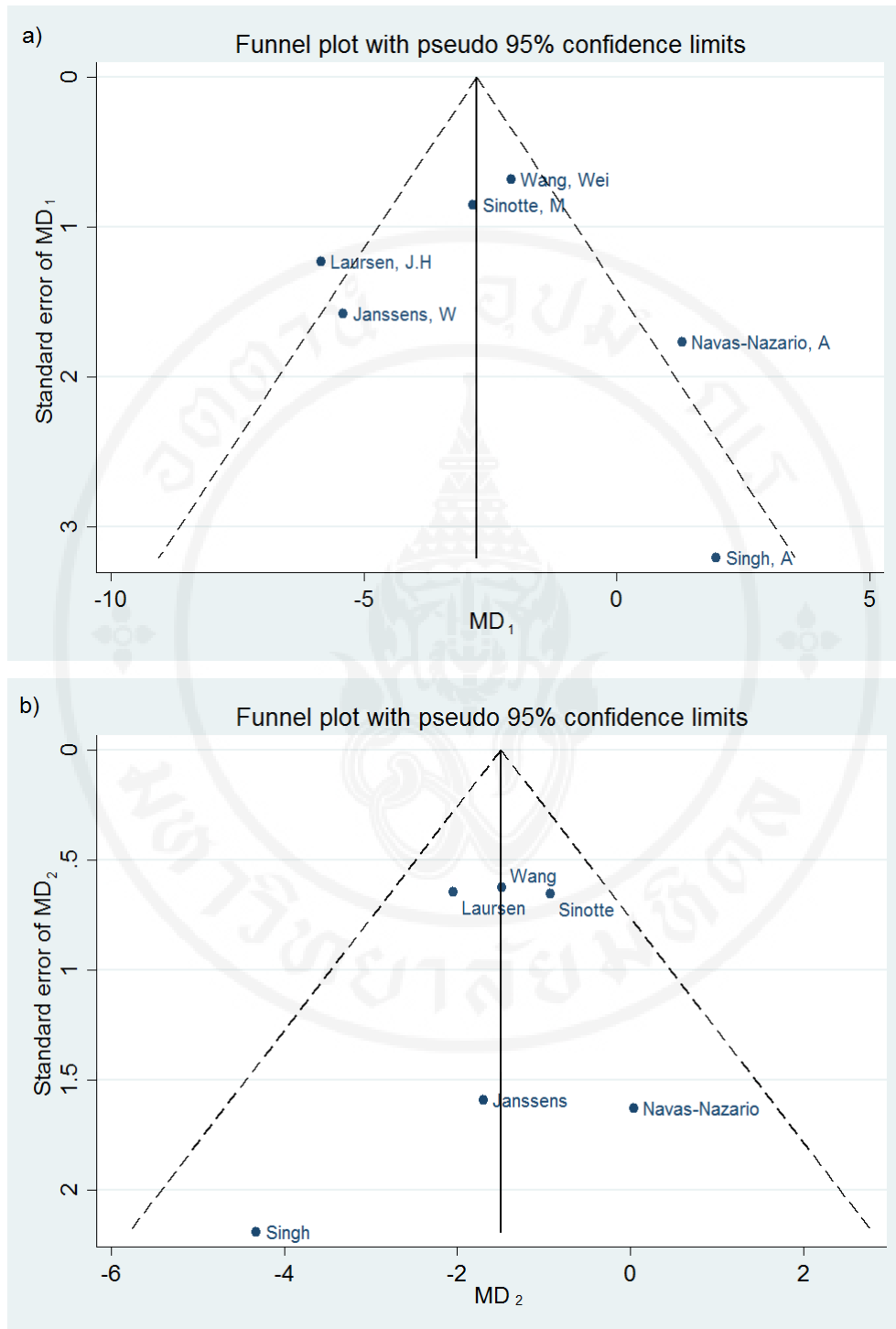
**Table 3.10** Describes genotype data and pooled mean difference of vitamin D levels for rs7041 (continued)

First Author, Year, (Reference No.)	GG			TG			TT			MD <sub>1</sub>	MD <sub>2</sub>
	n	mean	SD	n	mean	SD	n	mean	SD		
Hibler, 2012 <sup>54</sup>										-2.22 (-3.533, -0.907)	
Strawbridge, 2014 <sup>63</sup>										-0.023 (-0.031, -0.015)	
Gilbert, 2014 <sup>73</sup>										-1.65 (-2.532, -0.768)	
Pooled MD										-0.023 (-0.031, -0.015)	
Asians											
Lu, 2012 <sup>56</sup>										-0.015 (-0.023, -0.007)	
Li, 2014 <sup>21</sup>										-1.42 (-2.126, -0.714)	
Pooled MD										-0.015 (-0.023, -0.008)	

MD<sub>1</sub>; mean difference of TT versus GG, MD<sub>2</sub>; mean difference of TG versus GG, n; number of subjects, SD; standard deviation



**Figure 3.11** Forest plot of pooled MD in adult Caucasians for rs7041 a) MD<sub>1</sub> for TT versus GG, b) MD<sub>2</sub> for TG versus GG

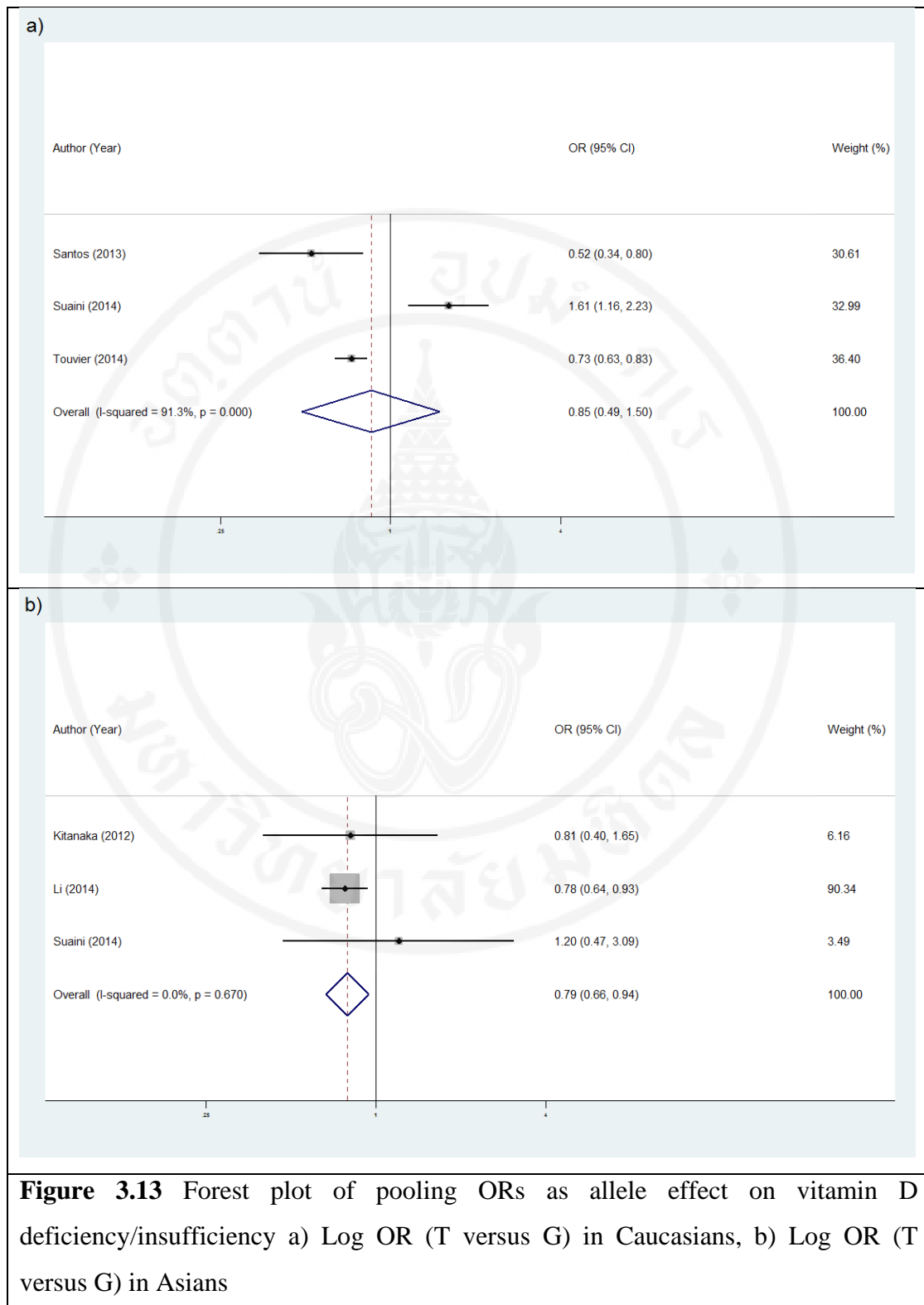


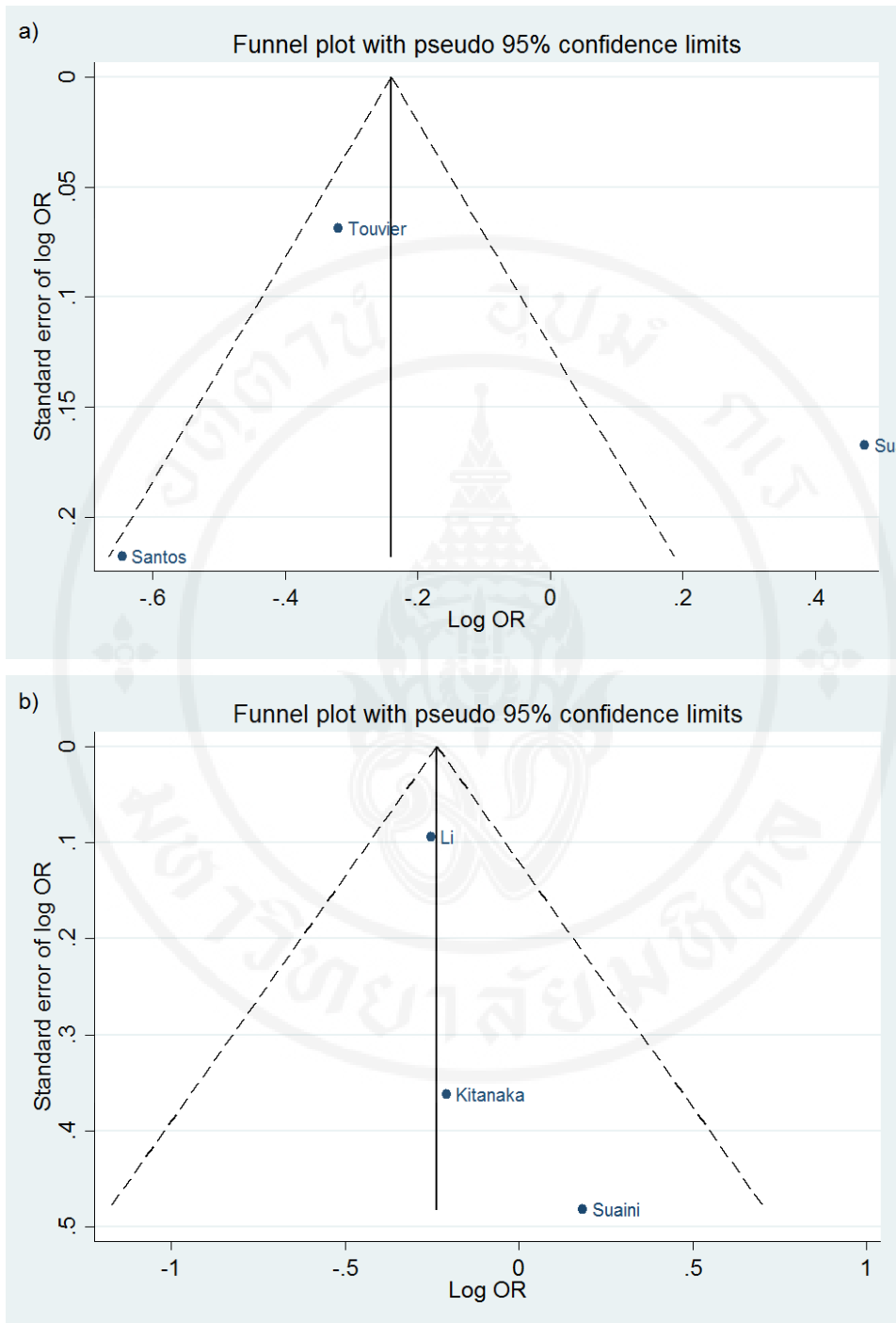
**Figure 3.12** Funnel plots for pooling MD<sub>1</sub> and MD<sub>2</sub> for rs7041 in Caucasian studies a) MD<sub>1</sub> (TT versus GG), b) MD<sub>2</sub> (TG versus GG)

**Table 3.11** Pooling allele effect of rs7041on vitamin D deficiency/insufficiency

First Author, Year, (Reference No.)	Case		Control		OR
	G	T	G	T	
Caucasians					
Santos, 2013 <sup>37</sup>	82	50	122	142	0.524 (0.342, 0.803)
Suaini, 2014 <sup>34</sup>	81	95	458	334	1.608 (1.158, 2.233)
Touvier, 2014 <sup>38</sup>	984	1080	599	905	0.726 (0.635, 0.831)
Pooled	1147	1225	1179	1381	0.854 (0.487, 1.499)
Asians					
Kitanaka, 2012 <sup>33</sup>	46	14	96	36	0.812 (0.399, 1.651)
Li, 2014 <sup>21</sup>	697	233	1026	442	0.776 (0.645, 0.934)
Suaini, 2014 <sup>34</sup>	7	23	30	82	1.202 (0.468, 3.089)
Pooled	750	270	1152	560	0.79 (0.662, 0.943)

CI; confidence interval, OR; odds ratio



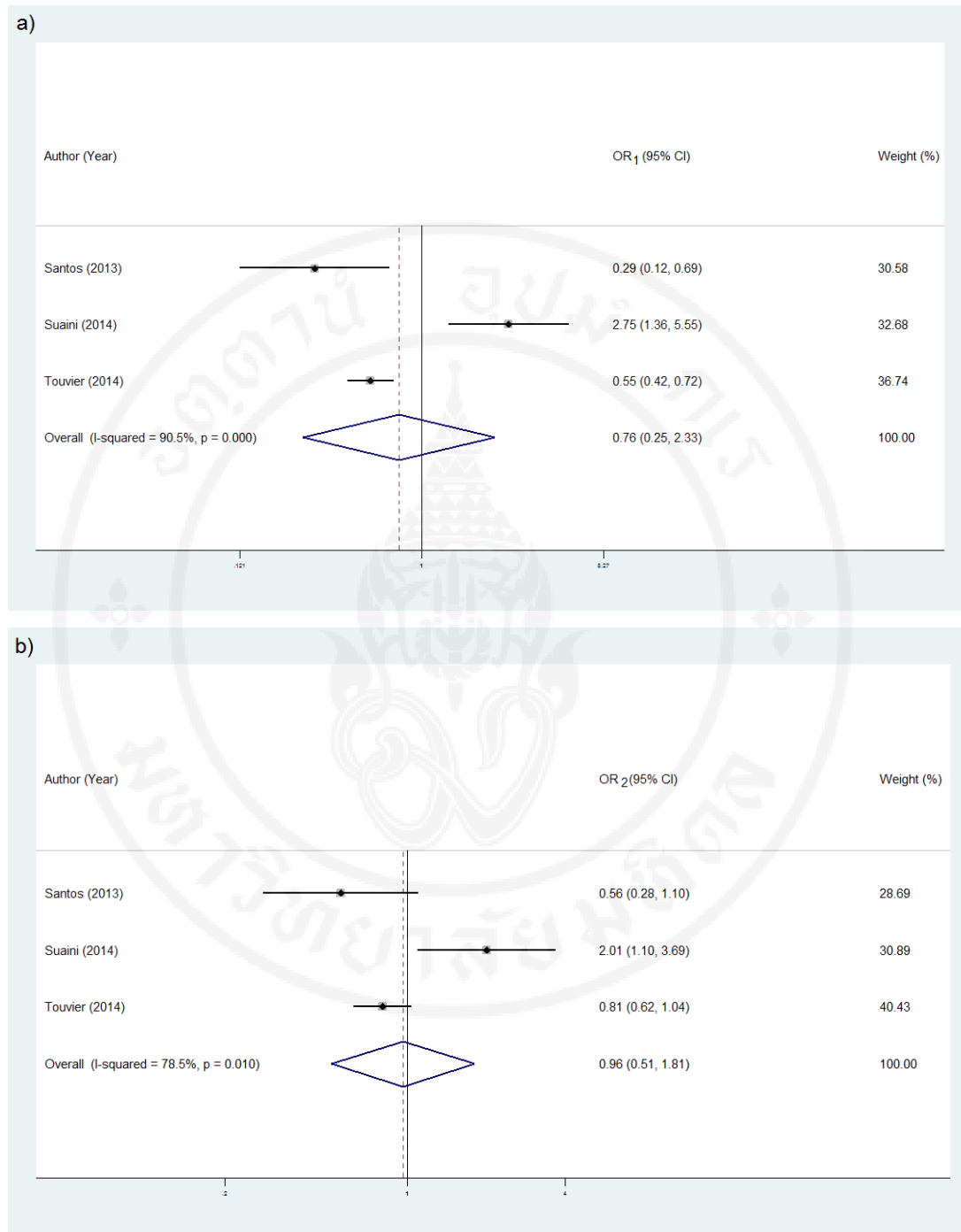


**Figure 3.14** Funnel plots of rs7041 allele effect on vitamin D deficiency/insufficiency a) Log OR (T versus G) in Caucasians, b) Log OR (T versus G) in Asians

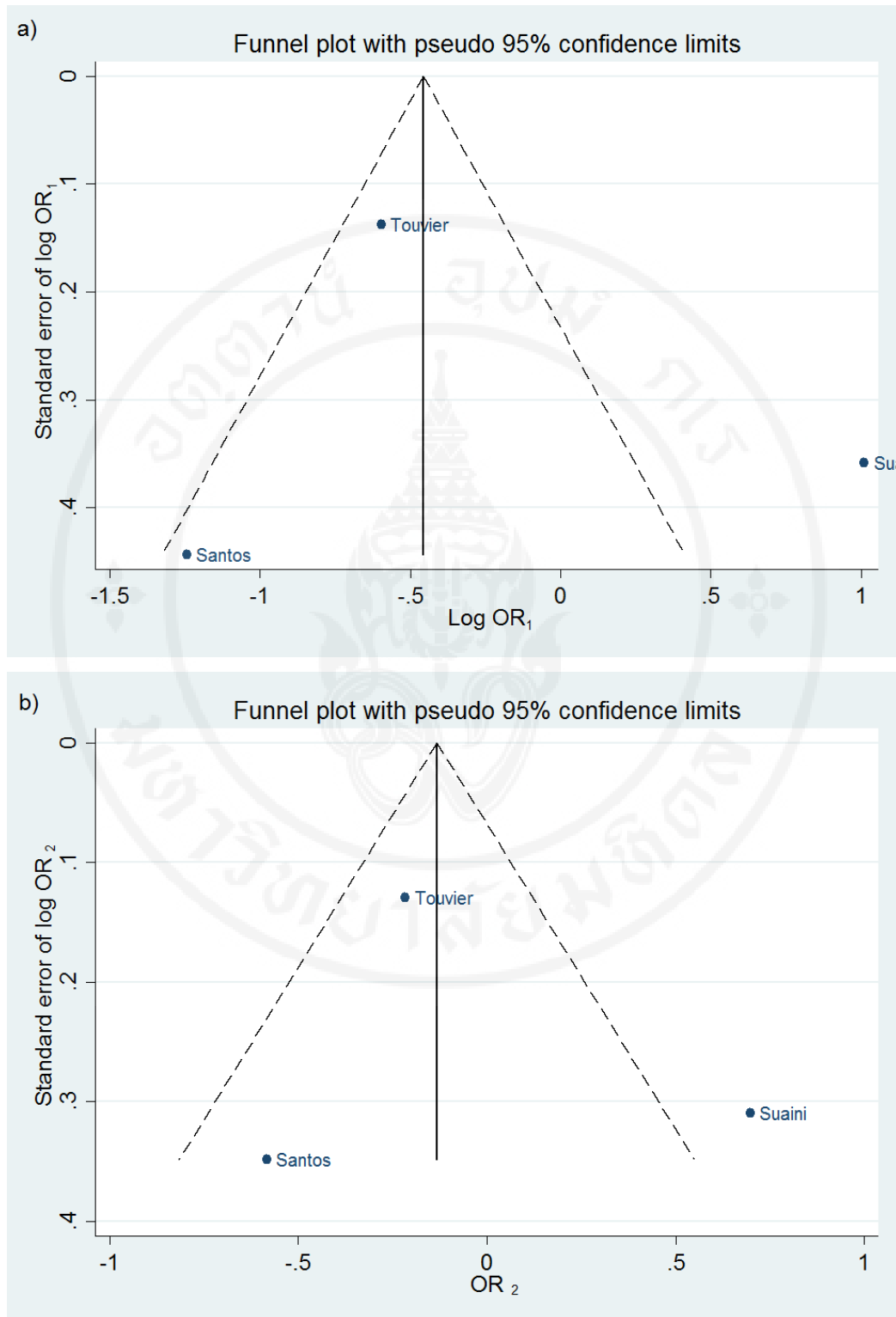
**Table 3.12** Pooling genotype effect of rs7041on vitamin D deficiency/insufficiency

First Author, Year, (Reference No.)	Case		Control		OR <sub>1</sub> (95% CI)	OR <sub>2</sub> (95% CI)
	GG	TG	TT	Control		
<b>Caucasians</b>						
Santos, 2013 <sup>37</sup>	26	30	10	30	0.288 (0.121, 0.688)	0.558 (0.282, 1.105)
Suaini, 2014 <sup>34</sup>	16	49	23	130	2.748 (1.362, 5.547)	2.011 (1.097, 3.686)
Touvier, 2014 <sup>38</sup>	240	504	288	130	0.551 (0.421, 0.722)	0.805 (0.625, 1.038)
<b>Pooled OR</b>	<b>282</b>	<b>583</b>	<b>321</b>	<b>290</b>	<b>0.761 (0.249, 2.329)</b>	<b>0.963 (0.509, 1.822)</b>
<b>Asians</b>						
Kitanaka, 2012 <sup>33</sup>	17	12	1	34	0.5 (0.052, 4.827)	0.857 (0.351, 2.092)
Li, 2014 <sup>21</sup>	268	161	36	364	0.679 (0.442, 1.044)	0.734 (0.572, 0.941)
Suaini, 2014 <sup>34</sup>	0	7	8	4	2.508 (0.123, 51.346)	3 (0.144, 62.488)
<b>Pooled OR</b>	<b>285</b>	<b>180</b>	<b>45</b>	<b>402</b>	<b>0.675 (0.452, 1.007)</b>	<b>0.746 (0.589, 0.946)</b>

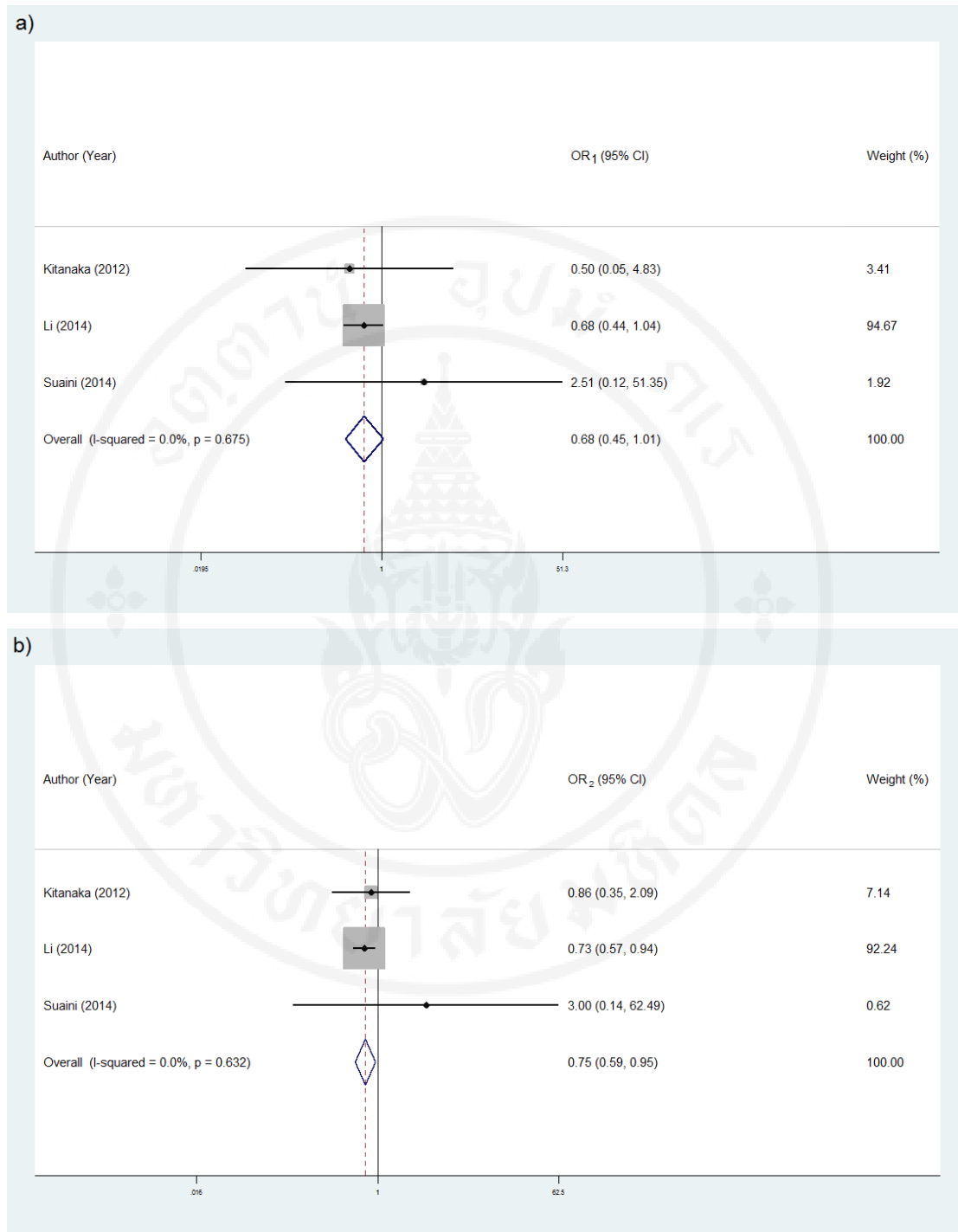
CI; confidence interval, OR<sub>1</sub>; odds ratio of TT versus GG, OR<sub>2</sub>; odds ratio of TG versus GG



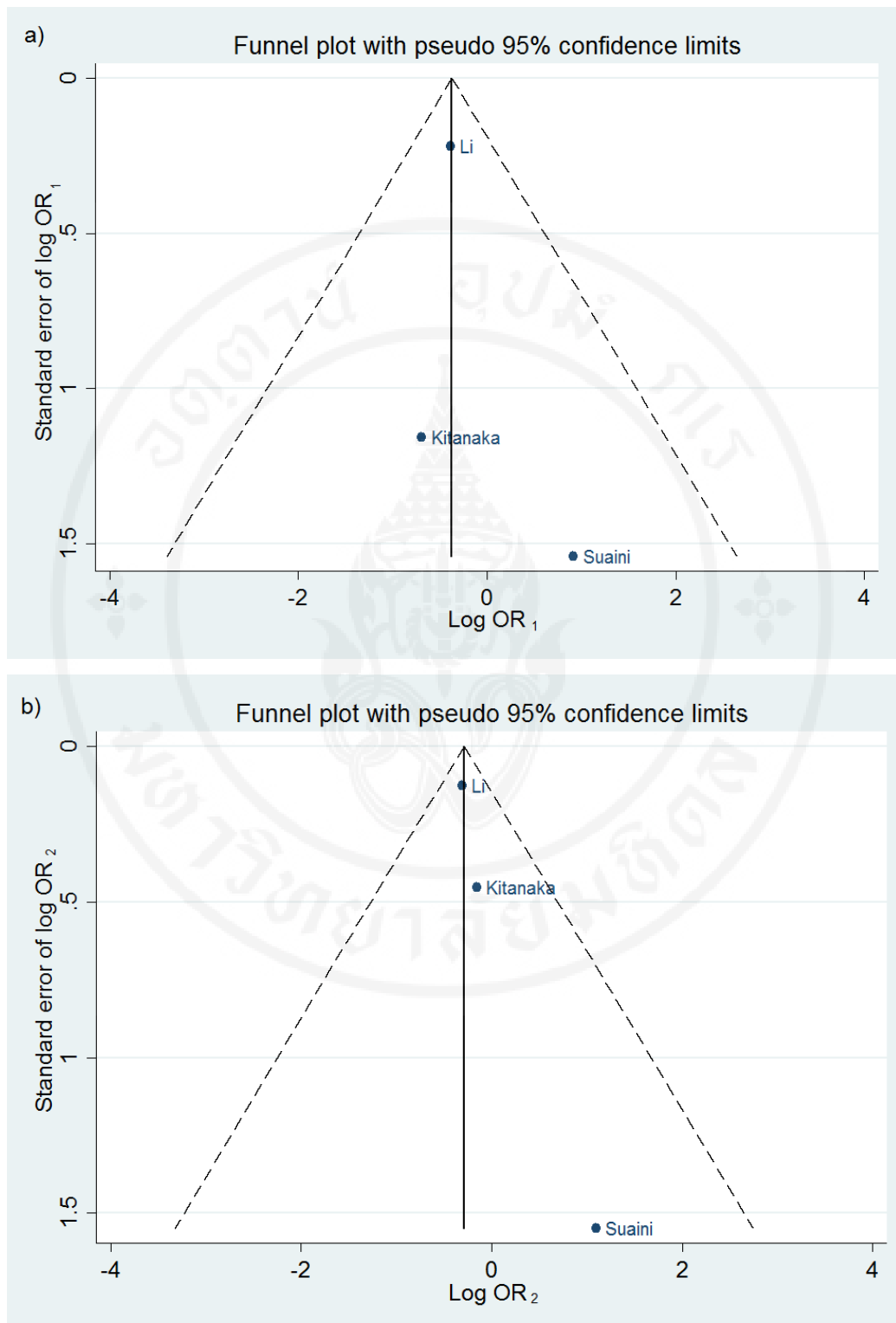
**Figure 3.15** Forest plots of rs7041 genotype effect on vitamin D deficiency/insufficiency in Caucasians a) Log OR<sub>1</sub> (TT versus GG), b) Log OR<sub>2</sub> (TG versus GG)



**Figure 3.16** Funnel plots of rs7041 genotype effect on vitamin D deficiency/insufficiency in Caucasians a) Log OR<sub>1</sub> (TT versus GG), b) Log OR<sub>2</sub> (TG versus GG).



**Figure 3.17** Forest plots of rs7041 genotype effect on vitamin D deficiency/insufficiency in Asians a) Log OR<sub>1</sub> (TT versus GG), b) Log OR<sub>2</sub> (TG versus GG)



**Figure 3.18** Funnel plots of rs7041 genotype effect on vitamin D deficiency/insufficiency in Asians a) Log OR<sub>1</sub> (TT versus GG), b) Log OR<sub>2</sub> (TG versus GG)

### 3.6 SNP rs2282679

Among 26 studies<sup>10, 21, 25, 26, 28, 29, 33-35, 49, 51, 52, 55-57, 62, 63, 65-73</sup> for rs2282679, 3<sup>33, 34, 67</sup>, 18<sup>10, 25, 26, 29, 35, 49, 55, 56, 62, 63, 65, 66, 68-73</sup>, and 5 studies<sup>21, 28, 51, 52, 57</sup> had vitamin D deficiency/insufficiency, vitamin D level, and both as the outcomes, respectively. From the 8 studies for pooling ORs, 5<sup>21, 28, 33, 34, 52</sup> and 2 studies<sup>51, 57</sup> had genotype frequencies and additive genotype effects, 1 study<sup>67</sup> had assessed allele effect.

From the 23 studies, 7 studies<sup>25, 28, 29, 65, 66, 68, 71</sup> reported means among genotypes, 1 study<sup>26</sup> reported the MDs, 10 studies<sup>10, 21, 35, 49, 56, 57, 63, 69, 70, 73</sup> reported additive genotype effects, 3 studies<sup>52, 55, 62</sup> had combined genotype effects, and 2 studies<sup>51, 72</sup> had MD for allele effect.

#### 3.6.1 Minor allele prevalence

Among 26 studies the data from 16 studies<sup>21, 25, 26, 28, 29, 33, 34, 52, 56, 62, 65, 66, 68, 70, 71, 73</sup> were used for pooling minor allele prevalence (see Table 3.13), because 1 study<sup>55</sup> observe HWE, and 9 studies<sup>10, 35, 49, 51, 57, 63, 67, 69, 72</sup> did not have data for pooling minor allele prevalence. There were 9 studies in Caucasians, 6 studies in Asians and 2 studies in Africans. Among 9 Caucasian studies, 2<sup>25, 34</sup> and 8 studies<sup>25, 26, 28, 29, 65, 66, 68, 73</sup> were respectively children and adults with the pooled minor allele G prevalence of 0.279 (95%CI: 0.257, 0.300) and 0.292 (95%CI: 0.259, 0.326). Among 6 Asian studies, 2<sup>33, 34</sup> and 4 studies<sup>21, 56, 70, 71</sup> were respectively children and adults with the minor allele G prevalence of 0.285 (95%CI: 0.170, 0.400) and 0.311 (95%CI: 0.226, 0.396). Among 2 African studies, with the pooled minor allele G prevalence of 0.068 (95%CI: 0.047, 0.089).

#### 3.6.2 Vitamin D level

The overall MDs for GG versus TT (MD<sub>1</sub>) and GT versus TT (MD<sub>2</sub>) from 7/8 studies<sup>25, 26, 28, 29, 65, 66, 68</sup> were described by ethnicity, see Table 3.14. Among them, 7 studies<sup>25, 26, 28, 29, 65, 66, 68</sup> were Caucasians, and 1 study<sup>71</sup> was Asian. Pooling in adult Caucasians yielded MD<sub>1</sub> and MD<sub>2</sub> of -3.407 (95%CI: -4.686, -2.127) and -1.673 (95%CI: -2.471, -0.875) with high heterogeneity for both ( $I^2 = 73.3\%$  and  $79.8\%$ , respectively), see Figure 3.19 a-b. This suggested that Caucasians with GG and GT genotypes had 3.4 ng/ml and 1.67 ng/ml lower levels of 25(OH)D than those with TT

genotype. Publication bias was assessed, funnel plots for pooling MD<sub>1</sub> and MD<sub>2</sub>, which showed asymmetry for both, see Figure 3.20 a-b. The Egger's tests were non-significant for both MD<sub>1</sub> and MD<sub>2</sub>, with (Coefficient = -1.041, SE = 1.973, P = 0.620) and (Coefficient = -3.591, SE = 2.064, P = 0.142) respectively.

There were 9/10 studies that reported additive genotype effects and their data were available for pooling. There were 5 studies<sup>35, 49, 57, 63, 73</sup> with Caucasians and 4 studies<sup>21, 56, 69, 70</sup> with Asians, with the MDs of -0.037 (95%CI: -0.046, -0.027) and -0.031 (95%CI: -0.039, -0.023) and both had high heterogeneity of ( $I^2 = 92.9%$ ) and ( $I^2 = 88.1%$ ), respectively. Publication bias for studies which reported the MD for the additive model, funnel plots of SE of MD against MD for Caucasians and Asians showed asymmetry for both, respectively, see Figure 3.21 a-b. The Egger's tests were non-significant for Caucasians with (Coefficient = -3.379, SE = 1.248, P = 0.073) and significant for Asians with (Coefficient = -3.081, SE = 0.349, P = 0.013). A contour enhanced funnel plot showed that there was the possibility of publication bias, see Figure 3.21 c.

### 3.6.3 Vitamin D deficiency/insufficiency

Among 4/5 studies that assessed G vs T allele effect on vitamin D deficiency/insufficiency and their data were available for pooling, 2<sup>28, 34</sup> and 3<sup>21, 33, 34</sup> were respectively studies in Caucasians and Asians, 1 study<sup>52</sup> was African, see Table 3.15. The pooled ORs (G vs T) for Caucasians and Asians were 1.397 (95%CI: 0.874, 2.234) and 1.272 (95%CI: 1.073, 1.507), with the heterogeneity  $I^2$  of 85.9% and  $I^2$  0%, respectively, see Figure 3.22. This suggested that Caucasians and Asians carrying G allele were approximately at 40% and 27% higher risk of developing vitamin D deficiency/insufficiency than those Caucasians and Asians carrying T allele. Publication bias was assessed for the Asians studies for the allele effect, funnel plot which showed symmetry, see Figure 3.23. The Egger's tests was non-significant with (coefficient = 0.063, SE = 0.466, P = 0.915).

Genotype data from studies for GG versus TT (OR<sub>1</sub>) and GT versus TT (OR<sub>2</sub>) were described, i.e., 2<sup>28, 34</sup> and 3 studies<sup>21, 33, 34</sup> were respectively Caucasian and Asians, see Table 3.16, 1 study<sup>52</sup> was African which was not included in the pooling after a sensitivity analysis. There were not enough studies to stratify them by type of subject

(children and adults). The  $OR_1$  and  $OR_2$  of Caucasians were, 1.733 (95%CI: 0.788, 3.811) and 1.507 (95%CI: 0.846, 2.686), with the heterogeneities  $I^2$  of 77.2% and 78.2%, respectively. The ORs of Asians,  $OR_1$  and  $OR_2$  were, 1.569 (95%CI: .055, 2.332) and 1.297 (95%CI: 1.028, 1.636), both homogenous with ( $I^2 = 0\%$ ) with respectively, see Figure 3.24 a-b. This suggested that Caucasians and Asians, who have GG genotype were approximately at 1.73 and 1.57 times higher risk of developing vitamin D deficiency/insufficiency than those Caucasians and Asians who have TT genotype. For publication bias for genotype effect of Asian studies, the funnel plots for both  $OR_1$  and  $OR_2$  showed symmetry, see Figure 3.25 a-b. The Egger's tests were non-significant for both  $OR_1$  and  $OR_2$ , with coefficients of -0.010 (SE = 0.418, P = 0.984) and 0.051 (SE = 0.149, P = 0.790), respectively.

**Table 3.13** Pooling the prevalence of the minor G allele of rs2282679

Author, Year, (Reference No.)	Ethnicity	Type of subjects	Total number of alleles	Minor G allele frequency	Prevalence of the minor allele (95% Confidence Interval)	HWE
Nissen, 2014 <sup>25</sup>	Caucasian	Children	680	180	0.265 (0.232, 0.298)	0.923
Suaini, 2014 <sup>34</sup>	Caucasian	Children	968	280	0.289 (0.261, 0.318)	0.060
Pooled			1648	460	0.279 (0.257, 0.300)	
Theodoratou, (2012) <sup>28</sup>	Caucasian	Adult	5556	1596	0.287 (0.275, 0.299)	0.298
Trummer (2012) <sup>66</sup>	Caucasian	Adult	2540	721	0.284 (0.266, 0.301)	0.250
Perna (2013) <sup>26</sup>	Caucasian	Adult	7482	2083	0.278 (0.289, 0.268)	0.868
Trummer (2013) <sup>65</sup>	Caucasian	Adult	6260	1769	0.283 (0.271, 0.294)	0.330
Gilbert (2014) <sup>73</sup>	Caucasian	Adult	3750	1097	0.293 (0.278, 0.307)	0.287
Laursen (2014) <sup>29</sup>	Caucasian	Adult	2962	818	0.276(0.26, 0.292)	0.439
Nissen, 2014 <sup>25</sup>	Caucasian	Adult	818	224	0.274 (0.243, 0.304)	0.923
Wang (2014) <sup>68</sup>	Caucasian	Adult	1746	843	0.483 (0.459, 0.506)	0.001 <sup>a</sup>
Pooled			23632	7068	0.291 (0.263, 0.320)	
Kitanaka, 2012 <sup>33</sup>	Asian	Children	132	30	0.227 (0.156, 0.299)	0.679
Suaini, 2014 <sup>34</sup>	Asian	Children	142	49	0.345 (0.267, 0.423)	0.774
Pooled			274	79	0.285 (0.170, 0.400)	
Lu (2012) <sup>56</sup>	Asian	Adult	6334	2613	0.413(0.4, 0.425)	0.124
Zhang (2013) <sup>70</sup>	Asian	Adult	5796	1797	0.31(0.298, 0.322)	0.127
Li (2014) <sup>21</sup>	Asian	Adult	2398	701	0.292 (0.274, 0.311)	0.830

**Table 3.13** Pooling the prevalence of the minor G allele of rs2282679 (continued)

Author, Year, (Reference No.)	Ethnicity	Type of subjects	Total number of alleles	Minor G allele frequency	Prevalence of the minor allele (95% Confidence Interval)	HWE
Thongthai, 2014 <sup>71</sup>	Asian	Adult	8952	2035	0.227 (0.219, 0.236)	0.363
Pooled			23480	7146	0.311 (0.226, 0.396)	
Signorello, 2011 <sup>62</sup>	African	Adult	750	59	0.079 (0.059, 0.098)	0.347
Foucan, 2013 <sup>52</sup>	African	Adult	594	34	0.057 (0.039, 0.076)	0.610
Pooled			1344	93	0.068 (0.047, 0.089)	

<sup>a</sup>This study had Hardy-Weinberg disequilibrium

CI; confidence interval, HWE; Hardy-Weinberg Equilibrium

**Table 3.14** Describes genotype data and pooled mean difference of vitamin D levels for rs2282679

First Author, Year, (Reference No.)	TT			GT			GG			MD <sub>1</sub>	MD <sub>2</sub>
	n	mean	SD	n	mean	SD	n	mean	SD		
<b>Adult Caucasians</b>											
Theodoratou (2012) <sup>28</sup>	1,400	14.01	8.65	1,160	13.29	8.24	218	12.59	8.45	-1.42 (-2.63, -0.21)	-0.72 (-1.376, -0.064)
Trummer (2012) <sup>66</sup>	643	16.9	9.56	533	13.77	7.75	94	11.66	5.75	-5.24 (-6.617, -3.863)	-3.13 (-4.119, -2.141)
Perna <sup>a</sup> (2013) <sup>26</sup>										-2.320 (-3.418, -1.222)	-0.880 (-1.429, -0.331)
Trummer (2013) <sup>65</sup>	1,622	18.05	10.19	1,247	16.86	9.34	261	14.33	7.76	-3.72 (-4.784, -2.656)	-1.19 (-1.907, -0.473)
Laursen (2014) <sup>29</sup>	770	33.21	5.29	604	29.89	14.07	107	28.08	36.3	-5.13 (-12.018, 1.758)	-3.32 (-4.503, -2.137)
Nissen (2014) <sup>25</sup>	219	29.57	9.68	156	28.08	9.19	34	25.48	8.58	-4.09 (-7.246, -0.934)	-1.49 (-3.42, 0.44)
Wang (2014) <sup>68</sup>	258	24.12	7.78	387	22.64	7.8	228	22.03	7.32	-2.09 (-3.433, -0.747)	-1.48 (-2.707, -0.253)
<b>Pooled MD</b>										<b>-3.407</b>	<b>-1.673</b> (-4.686, -2.127) (-2.471, -0.875)

**Table 3.14** Describes genotype data and pooled mean difference of vitamin D levels for rs2282679 (continued)

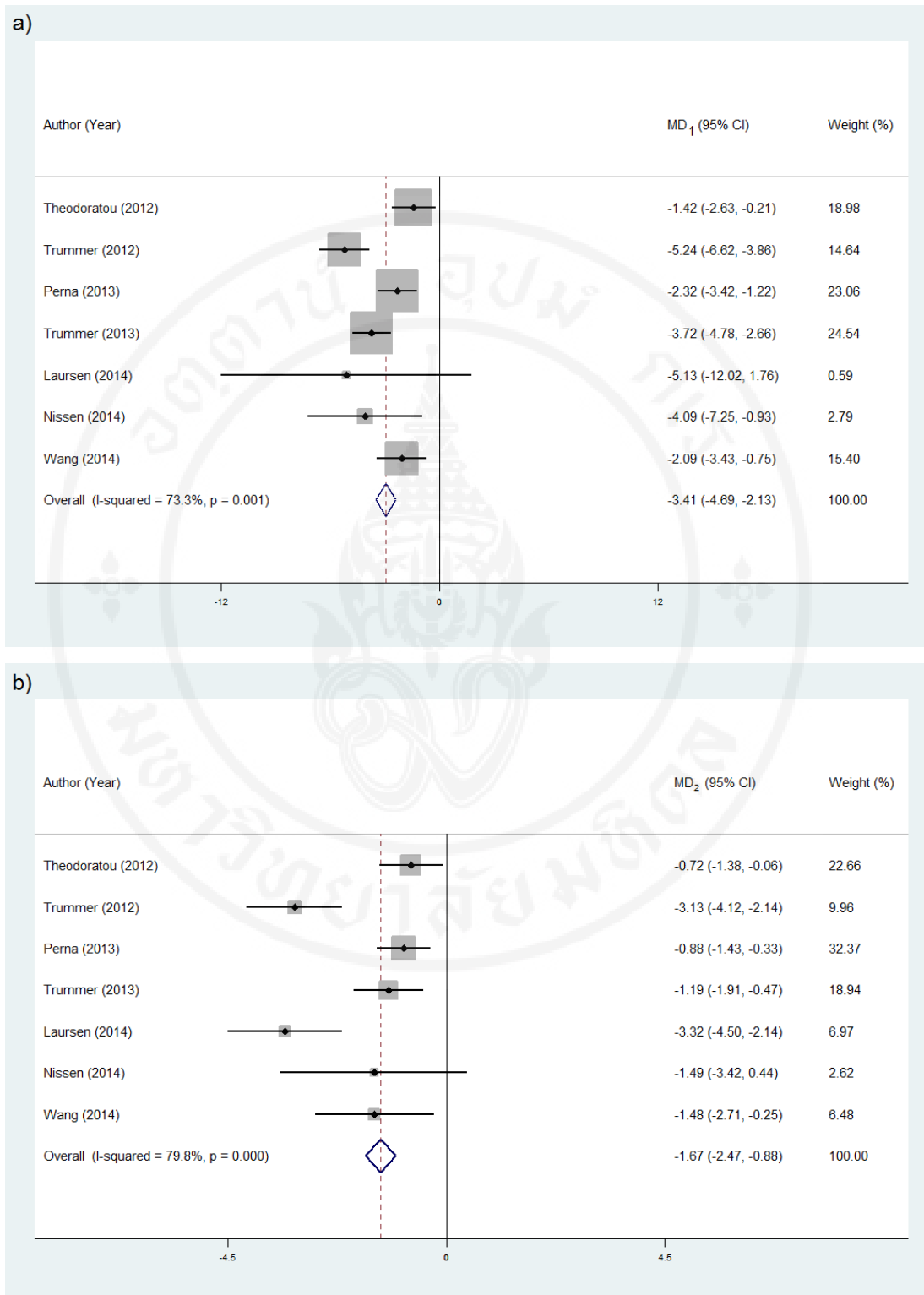
First Author, Year, (Reference No.)	TT			GT			GG			MD <sub>1</sub>	MD <sub>2</sub>
	n	mean	SD	n	mean	SD	n	mean	SD		
Additive model											
Caucasians											
Ahn (2010) <sup>49</sup>										-0.14 (-0.179, -0.101)	
Bu (2010) <sup>35</sup>										-0.99 (-2.872, 0.892)	
Muindi (2012) <sup>57</sup>										-2.49 (-4.489, -0.491)	
Strawbridge (2014) <sup>63</sup>										-0.03 (-0.04, -0.02)	
Gilbert (2014) <sup>73</sup>										-2.27 (-3.23, -1.31)	
Pooled MD										-0.037 (-0.046, -0.027)	
Asians											
Lu (2012) <sup>56</sup>										-0.029 (-0.037, -0.021)	
Zhang (2013) <sup>70</sup>										-0.153 (-0.263, -0.043)	

**Table 3.14** Describes genotype data and pooled mean difference of vitamin D levels for rs2282679 (continued)

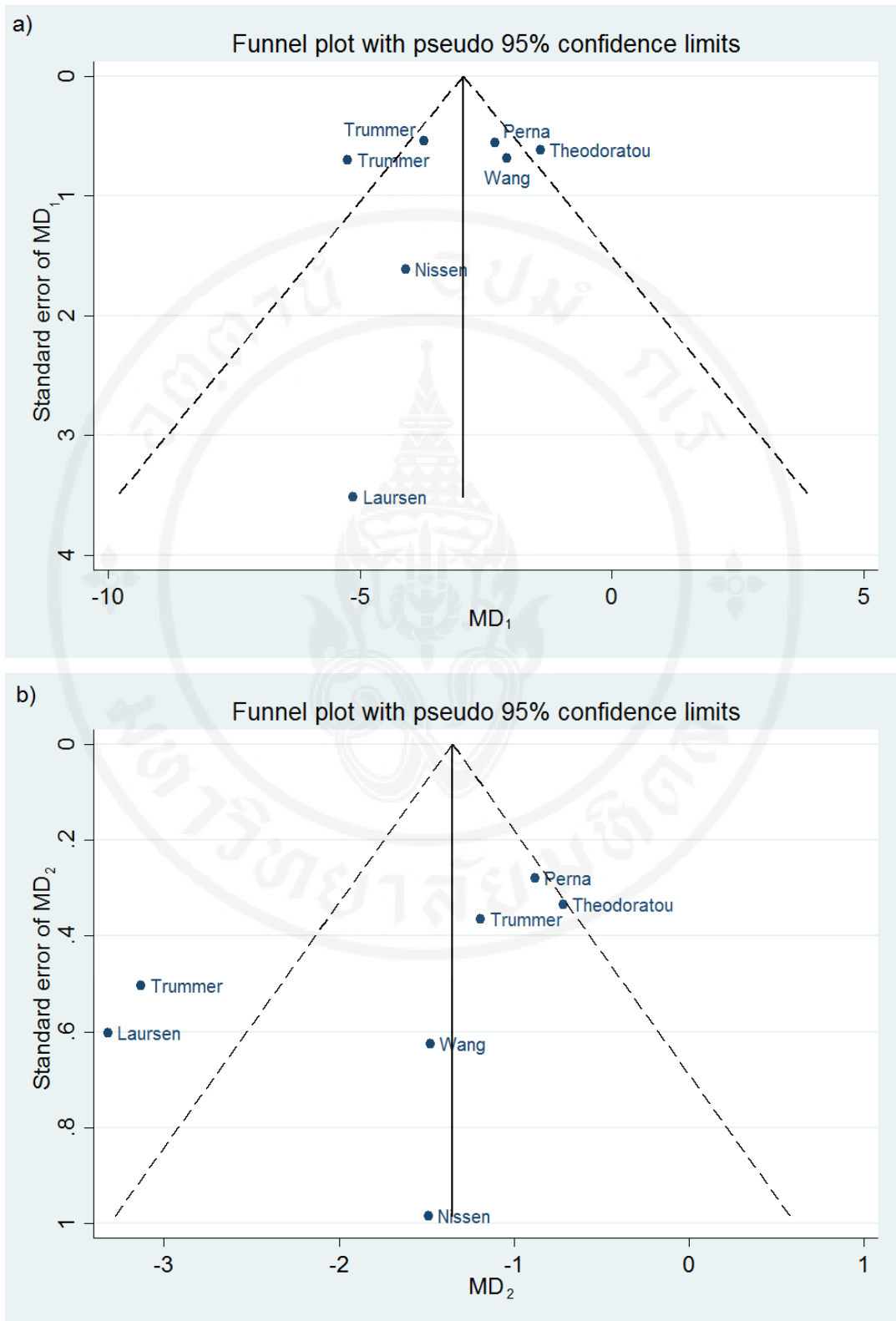
First Author, Year, (Reference No.)	TT			GT			GG			MD <sub>1</sub>	MD <sub>2</sub>
	n	mean	SD	n	mean	SD	n	mean	SD		
Li (2014) <sup>21</sup>										-1.23 (-1.936, -0.524)	
Yoshida (2014) <sup>69</sup>										-0.13 (-0.195, -0.065)	
Pooled MD										-0.031 (-0.039, -0.023)	

<sup>a</sup>Perna et al reported MD<sub>1</sub> and MD<sub>2</sub>

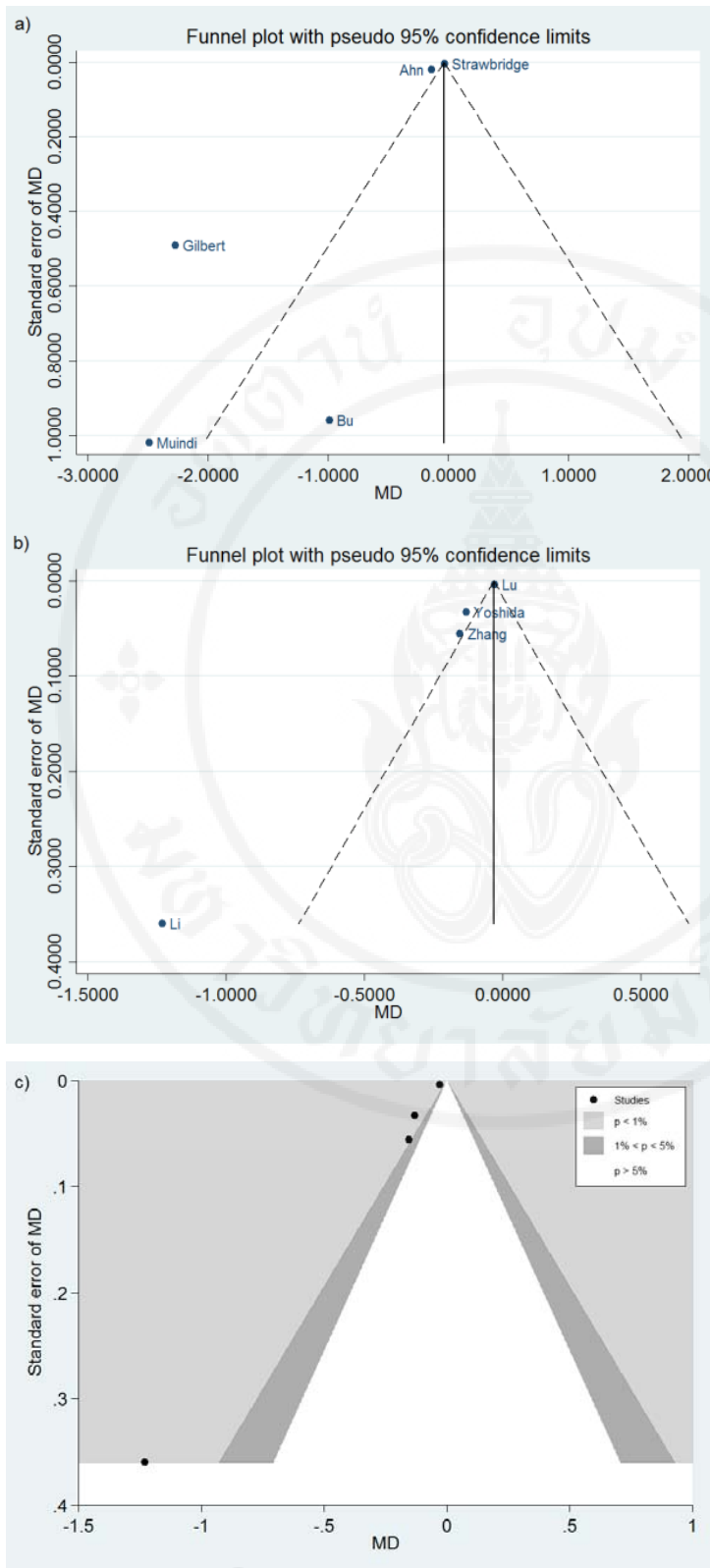
MD<sub>1</sub>; mean difference of GG versus TT, MD<sub>2</sub>; mean difference of GT versus TT, n; number of subjects, SD; standard deviation



**Figure 3.19** Forest plot of pooled MD in adult Caucasians for rs2282679 a) MD<sub>1</sub> for GG versus TT, b) MD<sub>2</sub> for GT versus TT



**Figure 3.20** Funnel plots for pooling MD<sub>1</sub> and MD<sub>2</sub> for rs2282679 in adult Caucasian studies a) MD<sub>1</sub> (GG versus TT), b) MD<sub>2</sub> (TG versus TT)

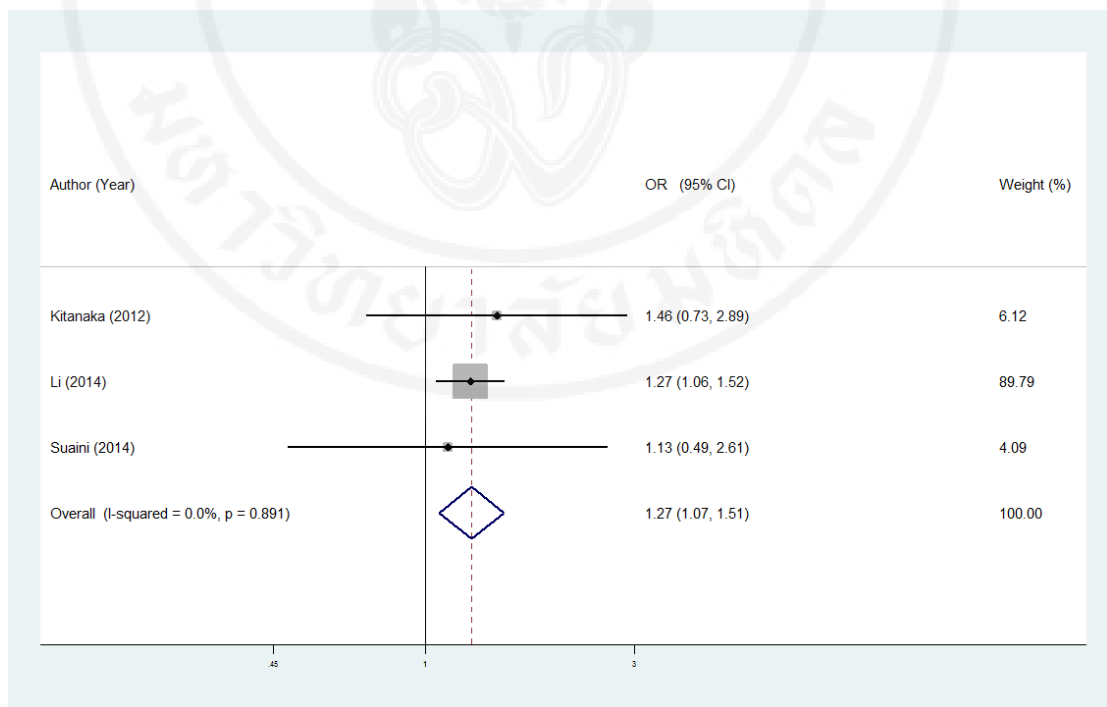


**Figure 3.21** Funnel plot of pooling MDs as additive effect for rs2282679 a) Caucasians, b) Asians, c) Contour enhanced funnel plot for Asians

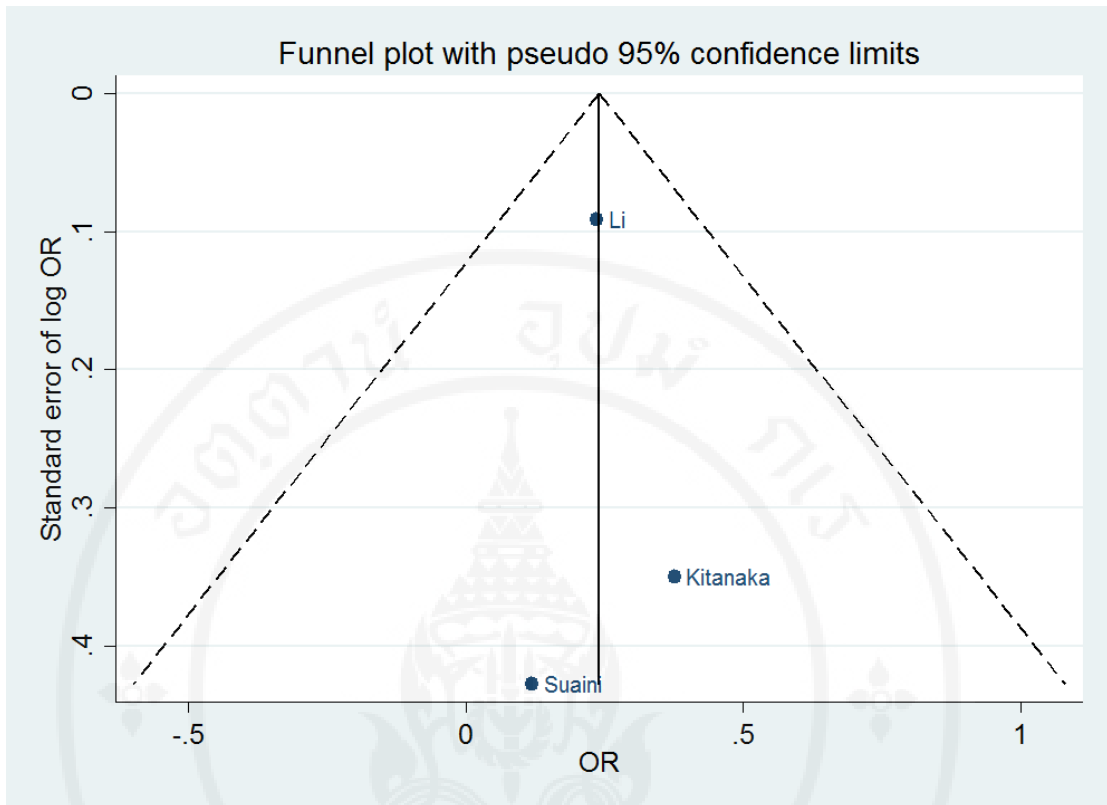
**Table 3.15** Pooling allele effect of rs2282679 on vitamin D deficiency/insufficiency

First Author, Year, (Reference No.)	Case		Control		OR (95% CI)
	T	G	T	G	
<b>Caucasians</b>					
Theodoratou, 2012 <sup>28</sup>	964	2232	1235	3231	1.13 (1.022, 1.249)
Suaini, 2014 <sup>34</sup>	70	106	210	582	1.83 (1.302, 2.573)
<b>Pooled</b>	<b>1034</b>	<b>2338</b>	<b>1445</b>	<b>3813</b>	<b>1.397 (0.874, 2.234)</b>
<b>Asians</b>					
Kitanaka, 2012 <sup>33</sup>	18	42	30	102	1.457 (0.734, 2.894)
Li, 2014 <sup>21</sup>	300	630	401	1067	1.267 (1.059, 1.516)
Suaini, 2014 <sup>34</sup>	11	19	38	74	1.127 (0.487, 2.61)
<b>Pooled</b>	<b>329</b>	<b>691</b>	<b>469</b>	<b>1243</b>	<b>1.272 (1.073, 1.507)</b>

CI; confidence interval, OR; odds ratio.



**Figure 3.22** Forest plot of rs2282679 allele effect on vitamin D deficiency/insufficiency in Asians, Log OR (G versus T)

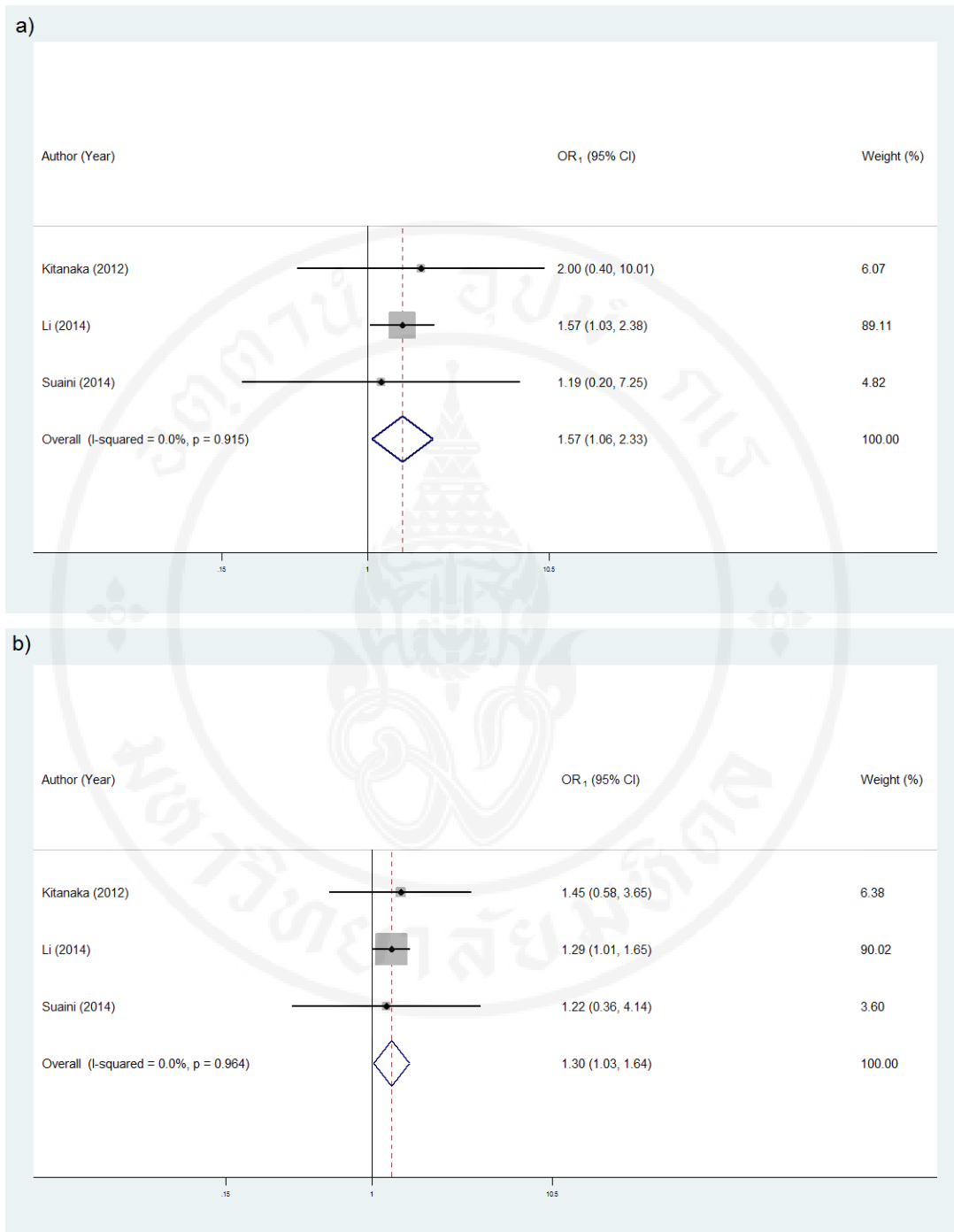


**Figure 3.23** Funnel plot of rs2282679 allele effect on vitamin D deficiency/insufficiency in Asians, Log OR (G versus T)

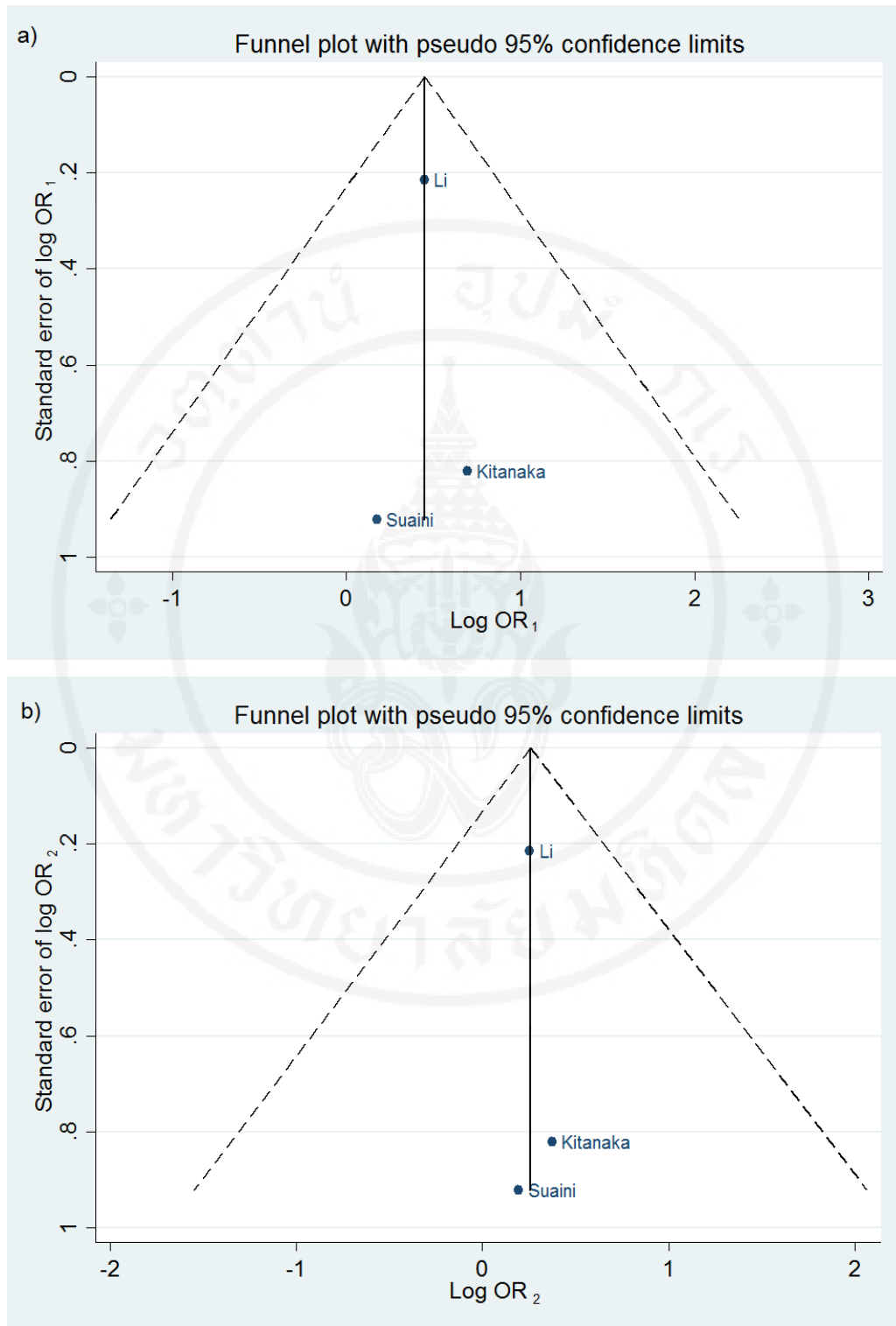
**Table 3.16** Pooling genotype effect of rs2282679 on vitamin D deficiency/insufficiency

First Author, Year, (Reference No.)	Case		Control		OR <sub>1</sub> (95% CI)	OR <sub>2</sub> (95% CI)
	TT	GG	TT	GG		
<b>Caucasians</b>						
Theodoratou, 2012 <sup>28</sup>	767	698	1163	905	1.222 (0.956, 1.563)	1.169 (1.022, 1.338)
Suaini, 2014 <sup>34</sup>	32	42	221	140	2.763 (1.342, 5.688)	2.072 (1.249, 3.438)
<b>Pooled OR</b>	<b>799</b>	<b>740</b>	<b>1384</b>	<b>1045</b>	<b>1.733 (0.788, 3.811)</b>	<b>1.507 (0.846, 2.686)</b>
<b>Asians</b>						
Kitanaka, 2012 <sup>33</sup>	15	12	40	22	2 (0.4, 10.008)	1.455 (0.58, 3.651)
Li, 2014 <sup>21</sup>	213	204	389	289	1.565 (1.028, 2.383)	1.289 (1.009, 1.647)
Suaini, 2014 <sup>34</sup>	6	7	25	24	1.19 (0.196, 7.249)	1.215 (0.357, 4.141)
<b>Pooled OR</b>	<b>234</b>	<b>223</b>	<b>454</b>	<b>335</b>	<b>1.569 (1.055, 2.332)</b>	<b>1.297 (1.028, 1.636)</b>

CI; confidence, OR<sub>1</sub>; odds ratio of GG versus TT, OR<sub>2</sub>; odds ratio of GT versus TT



**Figure 3.24** Forest plots of rs2282679 genotype effect on vitamin D deficiency/insufficiency in Asians a) Log OR<sub>1</sub> (GG versus TT), b) Log OR<sub>2</sub> (TG versus TT)



**Figure 3.25** Funnel plots of rs2282679 genotype effect on vitamin D deficiency/insufficiency in Asians a) Log OR<sub>1</sub> (GG versus TT), b) Log OR<sub>2</sub> (TG versus TT)

## CHAPTER IV

### DISCUSSION

The pooled minor allele prevalence of the rs4588, was similar for Caucasians and Asians and did not differ much among adults and children; 0.232 (95% CI: 0.190, 0.274), 0.287 (95% CI: 0.278, 0.296), 0.281 (95% CI: 0.158, 0.404) and 0.293 (95% CI: 0.275, 0.312), for children Caucasian, adults Caucasian, children Asians and adults Asians respectively. The pooled minor allele prevalence of the rs7041, was similar for children Caucasian and children Asian, but it was twice as large among adult Caucasian compared to the adult Asians; 0.442 (95% CI: 0.401, 0.483), 0.436 (95% CI: 0.353, 0.519), 0.506 (95% CI: 0.049, 0.964) and 0.287 (95% CI: 0.258, 0.316), for children Caucasian, adults Caucasian, children Asians and adults Asians respectively. The pooled minor allele prevalence of the rs2282679 was similar for Caucasians and Asians and did not differ much among adults and children but it was remarkably small among the adult Africans; 0.279 (95% CI: 0.257, 0.300), 0.291 (95% CI: 0.263, 0.320), 0.285 (95% CI: 0.170, 0.400), 0.311 (95% CI: 0.226, 0.396) and 0.068 (95% CI: 0.047, 0.089), for children Caucasian, adults Caucasian, children Asians, adults Asians and adult Africans respectively, see Table 18. There were no studies that explored the prevalence of the SNPs rs4588 and rs7041 among Africans.

The GC gene polymorphism rs4588 produces a biochemically different form of vitamin D binding protein (DBP 2)<sup>27</sup>. This different form of the DBP may have lower affinities for binding with vitamin D<sup>23</sup>. Studies have also suggested that this different form of DBP may be metabolized faster and this causes faster metabolism of 25(OH)D and so results in a lower level of vitamin D in the blood<sup>27</sup>. This meta-analysis showed that there was a negative association between those who have the SNP rs4588 and the serum vitamin D level. This negative association was seen across all ages and found in both Caucasians and Asians, see Table 19. Those who have this SNP have 4.042 ng/ml lower levels of vitamin D (for adults Asian) and since vitamin D deficiency is defined as having vitamin D levels below 20ng/ml, a

**Table 4.1** Summary of pooled prevalence

SNP	Ethnicity	Type of subjects	Pooled prevalence of the minor allele (95% CI)
rs4588	Caucasian	Children	0.232 (0.190, 0.274)
	Caucasian	Adult	0.287 (0.278, 0.296)
	Asian	Children	0.281 (0.158, 0.404)
	Asian	Adult	0.293 (0.275, 0.312)
rs7041	Caucasian	Children	0.442 (0.401, 0.483)
	Caucasian	Adult	0.436 (0.353, 0.519)
	Asian	Children	0.506 (0.049, 0.964)
	Asian	Adult	0.287 (0.258, 0.316)
rs2282679	Caucasian	Children	0.279 (0.257, 0.300)
	Caucasian	Adult	0.291 (0.263, 0.320)
	Asian	Children	0.285 (0.170, 0.400)
	Asian	Adult	0.311 (0.226, 0.396)
	African	Adult	0.068 (0.047, 0.089)

CI; confidence interval

**Table 4.2** Summary of pooled mean difference

SNP	Ethnicity/Type of subject	Pooled MD <sub>1</sub> (95% CI)	Pooled MD <sub>2</sub> (95% CI)	
rs4588	Caucasian children	-3.705 (-6.241, -1.169)	-2.211 (-2.930, -1.491)	
	Caucasian adults	-3.619 (-4.978, -2.261)	-1.601 (-2.369, -0.833)	
	Asian adults	-4.042 (-5.621, -2.462)	-1.722 (-2.754, -0.69)	
	(Additive model)	Caucasian adults	-1.932 (-2.719, -1.146)	
	Asian adults	-0.03 (-0.037, 0.022)		
rs7041	Caucasians	-2.707 (-4.91, -0.504)	-1.407 (-2.203, -0.612)	
	(Additive model)	Caucasians	-0.023 (-0.031, -0.015)	
	Asians	-0.015 (-0.023, -0.008)		
rs2282679	Caucasian adults	-3.407 (-4.686, -2.127)	-1.673 (-2.471, -0.875)	
	(Additive model)	Caucasians	-0.037 (-0.046, -0.027)	
	Asians	-0.031 (-0.039, -0.023)		

CI; confidence interval, MD<sub>1</sub>; mean difference between the homozygous risk genotype and the homozygous common genotype, MD<sub>2</sub>; mean difference between the heterozygous genotype and the homozygous common genotype

lowering of vitamin D level by 4ng/ml could be clinically significant<sup>41</sup>. Fewer studies reported the additive model, but this too showed that there is a mild negative association between rs4588 and the serum vitamin D level. There was heterogeneity and the sources of this need to be explored to assess this. In this review sources of heterogeneity could not be assessed because there was no consistency in reporting the sources of heterogeneity across studies. The odds of developing vitamin D deficiency or insufficiency were greater in those who carried the SNP rs4588, although the odds were only increased slightly, see Tables 20-21. The assessment of publication bias among studies that reported vitamin D deficiency/insufficiency and vitamin D level showed that there was no publication bias.

The rs7041 SNP produces a different isoform of the DBP (DBP 1) which may have a lower affinity for binding with vitamin D and affect the delivery of Vitamin D to the tissues and DBP 1 may also be metabolized faster which in turn causes faster metabolism of 25(OH)D and subsequent lower levels of vitamin D in the blood<sup>27 54</sup>. This meta-analysis showed that those who had this SNP had 2.707 ng/ml

**Table 4.3** Summary of Pooled allele effect of the SNP on vitamin D deficiency/insufficiency

SNP	Ethnicity	Pooled OR (95% CI)
rs4588	Caucasian	1.522 (1.336, 1.733)
	Asians	1.255 (1.058, 1.487)
rs7041	Caucasians	0.854 (0.487, 1.499)
	Asians	0.79 (0.662, 0.943)
rs2282679	Caucasians	1.397 (0.874, 2.234)
	Asians	1.272 (1.073, 1.507)

CI; confidence interval, OR; odds ratio between the risk allele and the common allele

lower levels of vitamin D. This negative association was seen in adults. There was only one study that reported the association in children. Fewer studies reported the additive model, but this too showed that there was a mild negative association between rs7041 and the serum vitamin D level. The assessment of the publication bias suggested none. The analysis of the association between the vitamin D deficiency/insufficiency and the SNP rs7041 showed a protective effect, that is, having

this SNP reduces the odds of developing vitamin D deficiency/insufficiency. Since having this SNP showed a negative association with the level of vitamin D, while at the same time the risk of having vitamin D deficiency/insufficiency is lowered, the association is giving a conflicting picture. Inability to explore the sources of heterogeneity, and the fewer number of studies could be the cause of this mixed picture of protective and harmful association. Publication bias assessment showed none.

**Table 4.4** Summary of Pooled genotype effect of the SNP on vitamin D deficiency/insufficiency

SNP	Ethnicity	Pooled OR <sub>1</sub> (95% CI)	Pooled OR <sub>2</sub> (95% CI)
rs4588	Caucasian	2.207 (1.629, 2.99)	1.529 (1.28, 1.826)
	Asians	1.578 (1.06, 2.348)	1.251 (0.991, 1.58)
rs7041	Caucasians	0.761 (0.249, 2.329)	0.963 (0.509, 1.822)
	Asians	0.675 (0.452, 1.007)	0.746 (0.589, 0.946)
rs2282679	Caucasians	1.733 (0.788, 3.811)	1.507 (0.846, 2.686)
	Asians	1.569 (1.055, 2.332)	1.297 (1.028, 1.636)

CI; confidence interval, OR<sub>1</sub>; odds ratio between the homozygous risk genotype and the homozygous common genotype, OR<sub>2</sub>; odds ratio between the heterozygous genotype and the homozygous common genotype

This meta-analysis showed that those who had the SNP rs2282679 had 3.407 ng/ml lower levels of vitamin D (Caucasians), which suggests a negative association. There were not enough studies to assess this association among adults and children or among Asians. Studies which reported the additive model suggested a negative association as well but this association was very small and the publication bias for these studies suggested that there could be missing studies among the Asian studies.

The strength of this study was in the greater sample size (overall sample size (N) = 67652, rs4588 only (n = 7840), rs7041 only (n = 879), rs2282679 only (n = 34940), rs4588 and rs7041 (n = 6077), rs7041 and rs2282679 (n = 5844), rs4588 and rs2282679 (n = 5016), and (n = 7056) had all three SNPs. which gave a more reliable indication of the effect of these three SNPs on the levels of vitamin D. In addition, two

databases were searched for relevant studies and two reviewers selected the studies. Therefore selection bias was less likely occurred.

In this review, Hispanics were pooled along in with the Caucasians, but the some studies had treated these two ethnicities as different, similarly Middle Eastern populations were pooled along in with Africans and South Asians were pooled with South East Asians. These ethnicities might not have been similar enough to be pooled together, but since there were only few of these studies, the choice was to either not include them in the pooling or pool them together with other ethnicities.

Several potential confounders (e.g., mean age, mean BMI, gender, physical activity, alcohol consumption, vitamin D supplementation, kidney disease, liver disease, serum vitamin D level, sun exposure, and smoking) were collected from the studies, but only the mean age, mean BMI and gender were reported with any consistency. This made it difficult to assess for sources of heterogeneity. Some studies<sup>27</sup> showed that there may be seasonal variations to the level of vitamin D in the blood. This study did not take seasonal variations into account because this information was not available for all the studies. Future research needs to be more consistent in the collection of confounding factors.

In conclusion, the SNPs rs4588 and rs2282679 showed a negative association with the level of serum vitamin D and the development of vitamin D deficiency or insufficiency. The SNP rs7041 gave a mixed picture and its association was inconclusive. However more studies need to be done with a consistent method for collecting the potential confounding factors, especially environmental factors. More studies need to be done on other ethnicities (such as Hispanic, African, Mediterranean and Indian) to assess their association with vitamin D and these SNPs.

## REFERENCES

- 1.Holick MF. Resurrection of vitamin D deficiency and rickets. *J Clin Invest.* 2006;116(8):2062-72.
- 2.Zhen D, Liu L, Guan C, Zhao N, Tang X. High prevalence of vitamin D deficiency among middle-aged and elderly individuals in northwestern China: its relationship to osteoporosis and lifestyle factors. *Bone.* 2015;71:1-6.
- 3.about vitamin D: vitamin D council; (<https://www.vitamindcouncil.org/about-vitamin-d/what-is-vitamin-d/#>).
- 4.Yoshida S, Ikari K, Furuya T, Toyama Y, Taniguchi A, Yamanaka H, et al. A GC polymorphism associated with serum 25-hydroxyvitamin D level is a risk factor for hip fracture in Japanese patients with rheumatoid arthritis: 10-year follow-up of the Institute of Rheumatology, Rheumatoid Arthritis cohort study. *Arthritis research & therapy.* 2014;16(2):R75.
- 5.Wang W, Ingles SA, Torres-Mejia G, Stern MC, Stanczyk FZ, Schwartz GG, et al. Genetic variants and non-genetic factors predict circulating vitamin D levels in Hispanic and non-Hispanic White women: the Breast Cancer Health Disparities Study. *Int J Mol Epidemiol Genet.* 2014;5(1):31-46.
- 6.Li LH, Yin XY, Wu XH, Zhang L, Pan SY, Zheng ZJ, et al. Serum 25(OH)D and vitamin D status in relation to VDR, GC and CYP2R1 variants in Chinese. *Endocrine journal.* 2014;61(2):133-41.
- 7.Abrahamsen BH, N. C. The role of vitamin D supplementation in patients with rheumatic diseases. *Nature Reviews Rheumatology.* 2013;9(7):411-22.
- 8.Gouni-Berthold IK, W.;Berthold, H. K. Vitamin D and cardiovascular disease. *Curr Vasc Pharmacol.* 2009;7(3):414-22.
- 9.Iyidir ÖTA, A. E. Vitamin D and diabetes mellitus. *Turkish Journal of Endocrinology and Metabolism.* 2012;16(4):89-94.
- 10.Buijsse BB, H.;Hirche, F.;Weikert, C.;Schulze, M. B.;Gottschald, M.;Kühn, T.;Katzke, V. A.;Teucher, B.;Dierkes, J.;Stangl, G. I.;Kaaks, R. Plasma

- 25-hydroxyvitamin D and its genetic determinants in relation to incident type 2 diabetes: A prospective case-cohort study. *Eur J Epidemiol.* 2013;28(9):743-52.
11. Blanton DH, Z.; Bierschenk, L.; Linga-Reddy, M. V. P.; Wang, H.; Clare-Salzler, M.; Haller, M.; Schatz, D.; Myhr, C.; She, J. X.; Wasserfall, C.; Atkinson, M. Reduced serum vitamin D-binding protein levels are associated with type 1 diabetes. *Diabetes.* 2011;60(10):2566-70.
12. Azad AKB, I.; Qiu, X.; Huang, H.; Cheng, D.; Liu, G.; Meyer, F.; Adjei, A.; Xu, W. Genetic sequence variants in vitamin D metabolism pathway genes, serum vitamin D level and outcome in head and neck cancer patients. *Int J Cancer.* 2013;132(11):2520-7.
13. Kidd LCRP, D. N.; Wang, S.; Chen, W.; Akereyeni, F.; Isaacs, W.; Ahaghotu, C.; Kittles, R. Sequence variation within the 5' regulatory regions of the vitamin D binding protein and receptor genes and prostate cancer risk. *Prostate.* 2005;64(3):272-82.
14. Davies JRC, Y. M.; Snowden, H.; Chan, M.; Leake, S.; Karpavicius, B.; Haynes, S.; Kukulizch, K.; Randerson-Moor, J.; Elliott, F.; Barth, J.; Kanetsky, P. A.; Harland, M.; Timothy Bishop, D.; Barrett, J. H.; Newton-Bishop, J. A. The determinants of serum vitamin D levels in participants in a melanoma case-control study living in a temperate climate. *Cancer Causes Control.* 2011;22(10):1471-82.
15. Grunhage F, Hochrath K, Krawczyk M, Hoblinger A, Obermayer-Pietsch B, Geisel J, et al. Common genetic variation in vitamin D metabolism is associated with liver stiffness. *Hepatology (Baltimore, Md).* 2012;56(5):1883-91.
16. Ashraf APH, C.; Alvarez, J. A.; Wang, X.; Gower, B. A. Insulin resistance indices are inversely associated with vitamin D binding protein concentrations. *J Clin Endocrinol Metab.* 2014;99(1):178-83.
17. Making SNPs Make Sense University of Utah Health Sciences: Genetic Science Learning Center; 2014 (<http://learn.genetics.utah.edu/content/pharma/snips/>).
18. Signorello LB, Shi J, Cai Q, Zheng W, Williams SM, Long J, et al. Common variation in vitamin D pathway genes predicts circulating 25-

- hydroxyvitamin D Levels among African Americans. *PloS one*. 2011;6(12):e28623.
19. Ryan LM, Chamberlain JM, Singer SA, Wood R, Tosi LL, Freishtat RJ, et al. Genetic influences on vitamin D status and forearm fracture risk in African American children. *Journal of investigative medicine : the official publication of the American Federation for Clinical Research*. 2012;60(6):902-6.
20. Janssens WB, R.;Claes, B.;Carremans, C.;Lehouck, A.;Buysschaert, I.;Coolen, J.;Mathieu, C.;Decramer, M.;Lambrechts, D. Vitamin D deficiency is highly prevalent in COPD and correlates with variants in the vitamin D-binding gene. *Thorax*. 2010;65(3):215-20.
21. Li LHY, X. Y.;Wu, X. H.;Zhang, L.;Pan, S. Y.;Zheng, Z. J.;Wang, J. G. Serum 25(OH)D and vitamin D status in relation to VDR, GC and CYP2R1 variants in Chinese. *Endocr J*. 2014;61(2):133-41.
22. Singh AF, M. K.;Subhi, Y.;Sørensen, T. L. The Association between Plasma 25-Hydroxyvitamin D and Subgroups in Age-Related Macular Degeneration: A Cross-Sectional Study. *PLoS One*. 2013;8(7).
23. Carpenter TOZ, J. H.;Parra, E.;Ellis, B. K.;Simpson, C.;Lee, W. M.;Balko, J.;Fu, L.;Wong, B. Y. L.;Cole, D. E. C. Vitamin D binding protein is a key determinant of 25-hydroxyvitamin D levels in infants and toddlers. *J Bone Miner Res*. 2013;28(1):213-21.
24. Navas-Nazario AL, F. Y.;Shabanova, V.;Weiss, P.;Cole, D. E. C.;Carpenter, T. O.;Bazzy-Asaad, A. Effect of vitamin D-binding protein genotype on the development of asthma in children. *Annals of Allergy, Asthma and Immunology*. 2014;112(6):519-24.
25. Nissen JR, L. B.;Ravn-Haren, G.;Wreford Andersen, E.;Hansen, B.;Andersen, R.;Mejborn, H.;Madsen, K. H.;Vogel, U. Common variants in CYP2R1 and GC genes predict vitamin D concentrations in healthy Danish children. *PLoS One*. 2014;9(2).
26. Perna LF, J. F.;Breitling, L. P.;Haug, U.;Raum, E.;Burwinkel, B.;Schöttker, B.;Brenner, H. Genetic variations in the vitamin d binding protein and

- season-specific levels of vitamin D among older adults. *Epidemiology*. 2013;24(1):104-9.
27. Sinotte MD, C.;Bérubé, S.;Pollak, M.;Brisson, J. Genetic polymorphisms of the vitamin D binding protein and plasma concentrations of 25-hydroxyvitamin D in premenopausal women. *Am J Clin Nutr*. 2009;89(2):634-40.
28. Theodoratou EP, T.;Zgaga, L.;Farrington, S. M.;McKeigue, P.;Din, F. V. N.;Tenesa, A.;Davey-Smith, G.;Dunlop, M. G.;Campbell, H. Instrumental variable estimation of the causal effect of plasma 25-hydroxy-vitamin D on colorectal cancer risk: A Mendelian randomization analysis. *PLoS One*. 2012;7(6).
29. Laursen JH, Sondergaard HB, Albrechtsen A, Frikke-Schmidt R, Koch-Henriksen N, Soelberg Sorensen P, et al. Genetic and environmental determinants of 25-hydroxyvitamin D levels in multiple sclerosis. *Mult Scler*. 2014.
30. Lu L, Sheng H, Li H, Gan W, Liu C, Zhu J, et al. Associations between common variants in GC and DHCR7/NADSYN1 and vitamin D concentration in Chinese Hans. *Human genetics*. 2012;131(3):505-12.
31. Porter TR, Li X, Stephensen CB, Mulligan K, Rutledge B, Flynn PM, et al. Genetic associations with 25-hydroxyvitamin D deficiency in HIV-1-infected youth: fine-mapping for the GC/DBP gene that encodes the vitamin D-binding protein. *Frontiers in genetics*. 2013;4:234.
32. Nissen J, Rasmussen LB, Ravn-Haren G, Andersen EW, Hansen B, Andersen R, et al. Common variants in CYP2R1 and GC genes predict vitamin D concentrations in healthy Danish children and adults. *PLoS One*. 2014;9(2):e89907.
33. Kitanaka SI, T.;Takaki, M.;Numakura, C.;Hayasaka, K.;Igarashi, T. Association of vitamin D-related gene polymorphisms with manifestation of vitamin D deficiency in children. *Endocr J*. 2012;59(11):1007-14.
34. Suaini NH, Koplín JJ, Ellis JA, Peters RL, Ponsonby AL, Dharmage SC, et al. Environmental and genetic determinants of vitamin D insufficiency in 12-month-old infants. *J Steroid Biochem Mol Biol*. 2014;144 Pt B:445-54.

35. Bu FXA, L.;Lappe, J.;Zhou, Y.;Gao, G.;Wang, H. W.;Recker, R.;Zhao, L. J. Comprehensive association analysis of nine candidate genes with serum 25-hydroxy vitamin D levels among healthy Caucasian subjects. *Hum Genet.* 2010;128(5):549-56.
36. Pekkinen MS, E.;Viljakainen, H. T.;Kokkonen, E.;Jakobsen, J.;Cashman, K.;Mäkitie, O.;Lamberg-Allardt, C. Vitamin D binding protein genotype is associated with serum 25-hydroxyvitamin D and PTH concentrations, as well as bone health in children and adolescents in Finland. *PLoS One.* 2014;9(1).
37. Santos BRM, L. P. G.;Boguszewski, M. C. S.;Spritzer, P. M. Variations in the Vitamin D-binding protein (DBP) gene are related to lower 25-hydroxyvitamin D levels in healthy girls: A cross-sectional study. *Horm Res Paediatr.* 2013;79(3):162-8.
38. Touvier M, Deschasaux M, Montourcy M, Sutton A, Charnaux N, Kesse-Guyot E, et al. Determinants of Vitamin D Status in Caucasian Adults: Influence of Sun Exposure, Dietary Intake, Sociodemographic, Lifestyle, Anthropometric, and Genetic Factors. *J Invest Dermatol.* 2014.
39. Thakkinstian A, McEvoy M, Chakravarthy U, Chakrabarti S, McKay GJ, Ryu E, et al. The association between complement component 2/complement factor B polymorphisms and age-related macular degeneration: a HuGE review and meta-analysis. *American journal of epidemiology.* 2012;176(5):361-72.
40. SI Unit Conversion Calculator: SBDR - SOCIETY FOR BIOMEDICAL DIABETES RESEARCH ([http://www.soc-bdr.org/rds/authors/unit\\_tables\\_conversions\\_and\\_genetic\\_dictionaries/conversion\\_in\\_si\\_units/index\\_en.html](http://www.soc-bdr.org/rds/authors/unit_tables_conversions_and_genetic_dictionaries/conversion_in_si_units/index_en.html)).
41. Holick MF, Binkley NC, Bischoff-Ferrari HA, Gordon CM, Hanley DA, Heaney RP, et al. Evaluation, treatment, and prevention of vitamin D deficiency: an Endocrine Society clinical practice guideline. *The Journal of clinical endocrinology and metabolism.* 2011;96(7):1911-30.
42. Viera AJ, Garrett JM. Understanding interobserver agreement: the kappa statistic. *Family medicine.* 2005;37(5):360-3.

43. Thakkinstian A, McEvoy M, Minelli C, Gibson P, Hancox B, Duffy D, et al. Systematic review and meta-analysis of the association between  $\beta$ 2-adrenoceptor polymorphisms and asthma: a HuGE review. *American journal of epidemiology*. 2005;162(3):201-11.
44. Bradburn MJ, J.J. Deeks, and D.G. Altman. metan-an alternative meta-analysis command. *Stata Technical Bulletin*. 1998;44:p. 4-15.
45. Cochran WG. Problems arising in the analysis of a series of experiments. *J Roy Stat Soc*. 1937; 4 (Suppl):p. 102-18.
46. Egger M, G.D. Smith, and D.G. Altman. *Systematic reviews in health care: Meta-analysis in context*. 2 nd ed ed. London: BMJ Books; 2001.
47. Petitti DB. *Meta-analysis, decision analysis, and cost-effectiveness analysis: Methods for quantitative synthesis in medicine* New York: Oxford University Press Inc 2000,.
48. StataCorp. *Stata: Release 13 . Statistical Software*. College Station, TX: StataCorp LP. 13 ed: College Station, TX: StataCorp LP.; 2013. p. Statistical Software.
49. Ahn JY, K.; Stolzenberg-Solomon, R.; Simon, K. C.; McCullough, M. L.; Gallicchio, L.; Jacobs, E. J.; Ascherio, A.; Helzlsouer, K.; Jacobs, K. B.; Li, Q.; Weinstein, S. J.; Purdue, M.; Virtamo, J.; Horst, R.; Wheeler, W.; Chanock, S.; Hunter, D. J.; Hayes, R. B.; Kraft, P.; Albanes, D. Genome-wide association study of circulating vitamin D levels. *Hum Mol Genet*. 2010;19(13):2739-45.
50. Berry DJV, K. S.; Whittaker, J. C.; Hingorani, A. D.; Hyppönen, E. Evaluation of genetic markers as instruments for Mendelian randomization studies on vitamin D. *PLoS One*. 2012;7(5).
51. Cheung CLL, K. S.; Sham, P. C.; Tan, K. C.; Kung, A. W. Genetic variant in vitamin D binding protein is associated with serum 25-hydroxyvitamin D and vitamin D insufficiency in southern Chinese. *J Hum Genet*. 2013;58(11):749-51.
52. Foucan LV-C, F. L.; Larifla, L.; Armand, C.; Deloumeaux, J.; Fagour, C.; Plumasseau, J.; Portlis, M. L.; Liu, L.; Bonnet, F.; Ducros, J. Polymorphisms in GC and

- NADSYN1 Genes are associated with vitamin D status and metabolic profile in Non-diabetic adults. *BMC Endocr Disord.* 2013;13.
53. Gozdzik AZ, J.; Wong, B. Y. L.; Fu, L.; Cole, D. E. C.; Parra, E. J. Association of vitamin D binding protein (VDBP) polymorphisms and serum 25(OH)D concentrations in a sample of young Canadian adults of different ancestry. *J Steroid Biochem Mol Biol.* 2011;127(3-5):405-12.
54. Hibler EAH, C.; Jurutka, P. W.; Martinez, M. E.; Jacobs, E. T. Polymorphic variation in the GC and CASR genes and associations with vitamin D metabolite concentration and metachronous colorectal neoplasia. *Cancer Epidemiology Biomarkers and Prevention.* 2012;21(2):368-75.
55. Ismail MFE, H. G.; Fouda, E. M. Genetic variants in vitamin D pathway in Egyptian asthmatic children: A pilot study. *Hum Immunol.* 2013;74(12):1659-64.
56. Lu LS, H.; Li, H.; Gan, W.; Liu, C.; Zhu, J.; Loos, R. J. F.; Lin, X. Associations between common variants in GC and DHCR7/NADSYN1 and vitamin D concentration in Chinese Hans. *Hum Genet.* 2012;131(3):505-12.
57. Muindi JRA, A. A.; Wu, Z. R.; Olson, I.; Huang, H.; Groman, A.; Tian, L.; Singh, P. K.; Sucheston, L. E.; Johnson, C. S.; Trump, D. L.; Fakih, M. G. Serum Vitamin D Metabolites in Colorectal Cancer Patients Receiving Cholecalciferol Supplementation: Correlation with Polymorphisms in the Vitamin D Genes. *Hormones and Cancer.* 2013;4(4):242-50.
58. Nimitphong HS, S.; Chanprasertyotin, S.; Chailurkit, L. O.; Ongphiphadhanakul, B. Changes in circulating 25-hydroxyvitamin D according to vitamin D binding protein genotypes after vitamin D3 or D2 supplementation. *Nutrition Journal.* 2013;12(1).
59. Petta SG, S.; Marco, V. D.; Scazzone, C.; MacAluso, F. S.; Cammà, C.; Cabibi, D.; Pipitone, R.; Craxì, A. Association of vitamin D serum levels and its common genetic determinants, with severity of liver fibrosis in genotype 1 chronic hepatitis C patients. *J Viral Hepat.* 2013;20(7):486-93.
60. Powe CEE, M. K.; Wenger, J.; Zonderman, A. B.; Berg, A. H.; Nalls, M.; Tamez, H.; Zhang, D.; Bhan, I.; Karumanchi, S. A.; Powe, N. R.; Thadhani, R.

- Vitamin D-binding protein and vitamin D status of black Americans and white Americans. *N Engl J Med.* 2013;369(21):1991-2000.
61. Robien KB, L. M.; Wang, R.; Beckman, K. B.; Walek, D.; Koh, W. P.; Yuan, J. M. Genetic and environmental predictors of serum 25-hydroxyvitamin D concentrations among middle-aged and elderly Chinese in Singapore. *Br J Nutr.* 2013;109(3):493-502.
62. Signorello LBS, J.; Cai, Q.; Zheng, W.; Williams, S. M.; Long, J.; Cohen, S. S.; Li, G.; Hollis, B. W.; Smith, J. R.; Blot, W. J. Common variation in vitamin D pathway genes predicts circulating 25-hydroxyvitamin D levels among African Americans. *PLoS One.* 2011;6(12).
63. Strawbridge RJD, A.; McLeod, O.; Folkersen, L.; Kavousi, M.; Gertow, K.; Baldassarre, D.; Veglia, F.; Leander, K.; Gigante, B.; Kauhanen, J.; Rauramaa, R.; Smit, A. J.; Mannarino, E.; Giral, P.; Dehghan, A.; Hofman, A.; Franco, O. H.; Humphries, S. E.; Tremoli, E.; De Faire, U.; Gustafsson, S.; Östenson, C. G.; Eriksson, P.; Öhrvik, J.; Hamsten, A. A serum 25-hydroxyvitamin D concentration-associated genetic variant in DHCR7 interacts with type 2 diabetes status to influence subclinical atherosclerosis (measured by carotid intima-media thickness). *Diabetologia.* 2014;57(6):1159-72.
64. Suzuki MY, M.; Hashimoto, M.; Murakami, M.; Kawasaki, K.; Noya, M.; Takahashi, D.; Urashima, M. 25-hydroxyvitamin D, vitamin D receptor gene polymorphisms, and severity of Parkinson's disease. *Mov Disord.* 2012;27(2):264-71.
65. Trummer OP, S.; Hoffmann, M. M.; Winkelmann, B. R.; Boehm, B. O.; März, W.; Pieber, T. R.; Obermayer-Pietsch, B.; Renner, W. Vitamin D and mortality: A Mendelian randomization study. *Clin Chem.* 2013;59(5):793-7.
66. Trummer OS, V.; Walter-Finell, D.; Lerchbaum, E.; Renner, W.; Gugatschka, M.; Dobnig, H.; Pieber, T. R.; Obermayer-Pietsch, B. Allelic determinants of vitamin D insufficiency, bone mineral density, and bone fractures. *J Clin Endocrinol Metab.* 2012;97(7):E1234-E40.

67. Wang TJZ, F.;Richards, J. B.;Kestenbaum, B.;Van Meurs, J. B.;Berry, D.;Kiel, D. P.;Streeten, E. A.;Ohlsson, C.;Koller, D. L.;Peltonen, L.;Cooper, J. D.;O'Reilly, P. F.;Houston, D. K.;Glazer, N. L.;Vandenput, L.;Peacock, M.;Shi, J.;Rivadeneira, F.;McCarthy, M. I.;Anneli, P.;De Boer, I. H.;Mangino, M.;Kato, B.;Smyth, D. J.;Booth, S. L.;Jacques, P. F.;Burke, G. L.;Goodarzi, M.;Cheung, C. L.;Wolf, M.;Rice, K.;Goltzman, D.;Hidiroglou, N.;Ladouceur, M.;Wareham, N. J.;Hocking, L. J.;Hart, D.;Arden, N. K.;Cooper, C.;Malik, S.;Fraser, W. D.;Hartikainen, A. L.;Zhai, G.;Macdonald, H. M.;Forouhi, N. G.;Loos, R. J. F.;Reid, D. M.;Hakim, A.;Dennison, E.;Liu, Y.;Power, C.;Stevens, H. E.;Jaana, L.;Vasan, R. S.;Soranzo, N.;Bojunga, J.;Psaty, B. M.;Lorentzon, M.;Foroud, T.;Harris, T. B.;Hofman, A.;Jansson, J. O.;Cauley, J. A.;Uitterlinden, A. G.;Gibson, Q.;Järvelin, M. R.;Karasik, D.;Siscovick, D. S.;Econs, M. J.;Kritchevsky, S. B.;Florez, J. C.;Todd, J. A.;Dupuis, J.;Hyppönen, E.;Spector, T. D. Common genetic determinants of vitamin D insufficiency: A genome-wide association study. *The Lancet*. 2010;376(9736):180-8.
68. Wang WI, S. A.;Torres-Mejía, G.;Stern, M. C.;Stanczyk, F. Z.;Schwartz, G. G.;Nelson, D. O.;Fejerman, L.;Wolff, R. K.;Slattery, M. L.;John, E. M. Genetic variants and non-genetic factors predict circulating vitamin D levels in Hispanic and non-Hispanic White women: The breast cancer health disparities study. *International Journal of Molecular Epidemiology and Genetics*. 2014;5(1):31-46.
69. Yoshida SI, K.;Furuya, T.;Toyama, Y.;Taniguchi, A.;Yamanaka, H.;Momohara, S. A GC polymorphism associated with serum 25-hydroxyvitamin D level is a risk factor for hip fracture in Japanese patients with rheumatoid arthritis: 10-year follow-up of the Institute of Rheumatology, Rheumatoid Arthritis cohort study. *Arthritis Res Ther*. 2014;16(2):R75.
70. Zhang ZH, J. W.;Fu, W. Z.;Zhang, C. Q.;Zhang, Z. L. An analysis of the association between the vitamin D pathway and serum 25-hydroxyvitamin D levels in a healthy Chinese population. *J Bone Miner Res*. 2013;28(8):1784-92.

71. Thongthai P, Chailurkit LO, Chanprasertyothin S, Nimitphong H, Sritara P, Aekplakorn W, et al. Vitamin D Binding Protein Gene Polymorphism as a Risk Factor for Vitamin D Deficiency in Thais. *Endocr Pract.* 2014;1-18.
72. Leong A, Rehman W, Dastani Z, Greenwood C, Timpson N, Langsetmo L, et al. The Causal Effect of Vitamin D Binding Protein (DBP) Levels on Calcemic and Cardiometabolic Diseases: A Mendelian Randomization Study. *PLoS Med.* 2014;11(10):e1001751.
73. Gilbert R, Bonilla C, Metcalfe C, Lewis S, Evans DM, Fraser WD, et al. Associations of vitamin D pathway genes with circulating 25-hydroxyvitamin-D, 1,25-dihydroxyvitamin-D, and prostate cancer: a nested case-control study. *Cancer Causes Control.* 2014.
74. Boucher BJ. Inadequate vitamin D status: Does it contribute to the disorders comprising syndrome 'X'? *Br J Nutr.* 1998;79(4):315-27.
75. Braun AK, A.; Bichlmaier, R.; Kammerer, S.; Cleve, H. A novel sequence polymorphism in exon 1 of the human vitamin D-binding protein (GC) gene. *Hum Mol Genet.* 1993;2(12):2214.
76. Engelman CDF, T. E.; Langefeld, C. D.; Hicks, P. J.; Rich, S. S.; Wagenknecht, L. E.; Bowden, D. W.; Norris, J. M. Genetic and environmental determinants of 25-hydroxyvitamin D and 1,25-dihydroxyvitamin D levels in hispanic and African Americans. *J Clin Endocrinol Metab.* 2008;93(9):3381-8.
77. Hunter DDL, M.; Snieder, H.; MacGregor, A. J.; Swaminathan, R.; Thakker, R. V.; Spector, T. D. Genetic contribution to bone metabolism, calcium excretion, and vitamin D and parathyroid hormone regulation. *J Bone Miner Res.* 2001;16(2):371-8.
78. Jiang HX, D. H.; Guo, Y. F.; Shen, H.; Xiao, P.; Yang, F.; Chen, Y.; Zhang, F.; Recker, R. R.; Deng, H. W. Association analysis of vitamin D-binding protein gene polymorphisms with variations of obesity-related traits in Caucasian nuclear families. *Int J Obes.* 2007;31(8):1319-24.
79. Konotey-Ahulu FID. Group specific component and HIV-infection. *Lancet.* 1987;1(8544):1267-9.
80. Thorsen SUM, H. B.; Carstensen, B.; Fenger, M.; Thuesen, B. H.; Husemoen, L.; Bergholdt, R.; Brorsson, C.; Pociot, F.; Linneberg, A.; Svensson, J. No

- association between type 1 diabetes and genetic variation in vitamin D metabolism genes: A Danish study. *Pediatr Diabetes*. 2013.
81. Viatte S, Yarwood A, McAllister K, Al-Mudhaffer S, Fu B, Flynn E, et al. The role of genetic polymorphisms regulating vitamin D levels in rheumatoid arthritis outcome: A Mendelian randomisation approach. *Ann Rheum Dis*. 2014;73(7):1430-3.
82. Yarwood A, Viatte S, Plant D, Morgan AW, Isaacs J, Wilson AG, et al. Testing the role of vitamin D in response to antitumour necrosis factor  $\alpha$  therapy in a UK cohort: A Mendelian randomisation approach. *Ann Rheum Dis*. 2014;73(5):938-40.
83. Iruzubieta P, Teran A, Crespo J, Fabrega E. Vitamin D deficiency in chronic liver disease. *World J Hepatol*. 2014;6(12):901-15.
84. Wang G, Li Y, Li L, Yu F, Cui L, Ba Y, et al. Association of the vitamin D binding protein polymorphisms with the risk of type 2 diabetes mellitus: a meta-analysis. *BMJ Open*. 2014;4(11):e005617.
85. Horita N, Miyazawa N, Tomaru K, Inoue M, Ishigatsumo Y, Kaneko T. Vitamin D binding protein genotype variants and risk of chronic obstructive pulmonary disease: A meta-analysis. *Respirology*. 2015;20(2):219-25.
86. Sahana G, Gulbrandsen B, Thomsen B, Holm LE, Panitz F, Brøndum RF, et al. Genome-wide association study using high-density single nucleotide polymorphism arrays and whole-genome sequences for clinical mastitis traits in dairy cattle. *J Dairy Sci*. 2014;97(11):7258-75.
87. Abecia EM-J, B.;Casalod, Y.;Bell, B.;Pinilla, I.;Honrubia, F. M. Genetic markers in primary open-angle glaucoma. *Int Ophthalmol*. 1996;20(1-3):79-82.
88. Berg IH, C.;Sayles, H.;Romberger, D.;Nelson, A.;Meza, J.;Miller, B.;Wouters, E. F. M.;Macnee, W.;Rutten, E. P. A.;Romme, E. A. P. M.;Vestbo, Jø;Edwards, L.;Rennard, S. Vitamin D, vitamin D binding protein, lung function and structure in COPD. *Respir Med*. 2013;107(10):1578-88.
89. Correale JY, M. C.;Gaitan, M. I. Gender differences in 1,25 dihydroxyvitamin D3 immunomodulatory effects in multiple sclerosis patients and healthy subjects. *J Immunol*. 2010;185(8):4948-58.

90. Daiger SPM, M.; Chakraborty, R. Heritability of quantitative variation at the group-specific component (Gc) locus. *Am J Hum Genet.* 1984;36(3):663-76.
91. Martineau ARL, A. C. C. S.; Anderson, S. T.; Newton, S. M.; Wilkinson, K. A.; Nicol, M. P.; Pienaar, S. M.; Skolimowska, K. H.; Rocha, M. A.; Rolla, V. C.; Levin, M.; Davidson, R. N.; Bremner, S. A.; Griffiths, C. J.; Eley, B. S.; Bonecini-Almeida, M. G.; Wilkinson, R. J. Association between Gc genotype and susceptibility to TB is dependent on vitamin D status. *Eur Respir J.* 2010;35(5):1106-12.
92. Ramos-Lopez EK, H.; Weber, S.; Kukic, A.; Penna-Martinez, M.; Badenhoop, K.; Louwen, F. Gestational diabetes mellitus and vitamin D deficiency: Genetic contribution of CYP27B1 and CYP2R1 polymorphisms. *Diabetes, Obesity and Metabolism.* 2008;10(8):683-5.
93. Speeckaert MMG, G. L.; Vanholder, R.; Van Biesen, W.; Taes, Y. E.; Clement, F.; Wehlou, C.; Delanghe, J. R. Vitamin D Binding Protein and the Need for Vitamin D in Hemodialysis Patients. *J Ren Nutr.* 2008;18(5):400-7.
94. Taes YECG, S.; Huang, G.; Van Pottelbergh, I.; De Bacquer, D.; Verhasselt, B.; Van den Broeke, C.; Delanghe, J. R.; Kaufman, J. M. Vitamin D binding protein, bone status and body composition in community-dwelling elderly men. *Bone.* 2006;38(5):701-7.
95. Zhang YY, S.; Liu, Y.; Ren, L. Relationship between polymorphisms in vitamin D metabolism-related genes and the risk of rickets in Han Chinese children. *BMC Med Genet.* 2013;14(1).
96. Maisnam I, Dutta D, Mukhopadhyay S, Chowdhury S. Lean mass is the strongest predictor of bone mineral content in type-2 diabetes and normal individuals: An eastern India perspective. *Journal of Diabetes and Metabolic Disorders.* 2014;13(1).
97. Badawi A, Sayegh S, Sadoun E, Al-Thani M, Arora P, Haddad PS. Relationship between insulin resistance and plasma vitamin D in adults. *Diabetes, Metabolic Syndrome and Obesity: Targets and Therapy.* 2014;7:297-303.
98. Smolders J, Peelen E, Thewissen M, Menheere P, Damoiseaux J, Hupperts R. Circulating vitamin D binding protein levels are not associated with

- relapses or with vitamin D status in multiple sclerosis. *Mult Scler.* 2014;20(4):433-7.
99. Wilson RT, Bortner JD, Jr., Roff A, Das A, Battaglioli EJ, Richie JP, Jr., et al. Genetic and environmental influences on plasma vitamin D-binding protein concentrations. *Transl Res.* 2014.
100. Adams LAW, S. W.; Marsh, J. A.; Lye, S. J.; Connor, K. L.; Maganga, R.; Ayonrinde, O. T.; Olynyk, J. K.; Mori, T. A.; Beilin, L. J.; Palmer, L. J.; Hamdorf, J. M.; Pennell, C. E. Association between liver-specific gene polymorphisms and their expression levels with nonalcoholic fatty liver disease. *Hepatology.* 2013;57(2):590-600.
101. Alfred TB-S, Y.; Cooper, R.; Hardy, R.; Deary, I. J.; Elliott, J.; Harris, S. E.; Hyppönen, E.; Kivimaki, M.; Kumari, M.; Maddock, J.; Power, C.; Starr, J. M.; Kuh, D.; Day, I. N. M. Genetic variants influencing biomarkers of nutrition are not associated with cognitive capability in middle-aged and older adults. *J Nutr.* 2013;143(5):606-12.
102. Anderson LNC, M.; Cole, D. E. C.; Knight, J. A. Vitamin D-related genetic variants, interactions with vitamin D exposure, and breast cancer risk among caucasian women in Ontario. *Cancer Epidemiology Biomarkers and Prevention.* 2011;20(8):1708-17.
103. Anic GMT, R. C.; Nabors, L. B.; Olson, J. J.; Browning, J. E.; Madden, M. H.; Murtagh, F. R.; Forsyth, P. A.; Egan, K. M. An exploratory analysis of common genetic variants in the vitamin D pathway including genome-wide associated variants in relation to glioma risk and outcome. *Cancer Causes Control.* 2012;23(9):1443-9.
104. Daiger SPS, M. S.; Cavalli-Sforza, L. L. Group-specific component (Gc) proteins bind vitamin D and 25-hydroxyvitamin D. *Proc Natl Acad Sci U S A.* 1975;72(6):2076-80.
105. Davies JRF, S.; Randerson-Moor, J.; Harland, M.; Kumar, R.; Anic, G. M.; Nagore, E.; Hansson, J.; Höiom, V.; Jönsson, G.; Gruis, N. A.; Park, J. Y.; Guan, J.; Sivaramakrishna Rachakonda, P.; Wendt, J.; Pjanova, D.; Puig, S.; Schadendorf, D.; Okamoto, I.; Olsson, H.; Affleck, P.; García-Casado, Z.; Puig-Butille, J. A.; Stratigos, A. J.; Kodela, E.; Donina, S.; Sucker,

- A.;Hosen, I.;Egan, K. M.;Barrett, J. H.;van Doorn, R.;Bishop, D. T.;Newton-Bishop, J. An inherited variant in the gene coding for vitamin D-binding protein and survival from cutaneous melanoma: A BioGenoMEL study. *Pigment Cell and Melanoma Research*. 2014;27(2):234-43.
- 106.Dorjgochoo TS, J.;Gao, Y. T.;Long, J.;Delahanty, R.;Xiang, Y. B.;Cai, Q.;Shu, X. O. Genetic variants in vitamin D metabolism-related genes and body mass index: Analysis of genome-wide scan data of approximately 7000 Chinese women. *Int J Obes*. 2012;36(9):1252-5.
- 107.Epstein MMA, O.;Kasperzyk, J. L.;Shui, I. M.;Penney, K. L.;Fall, K.;Rider, J. R.;Stampfer, M. J.;Andersson, S. O.;Giovannucci, E.;Mucci, L. A. Seasonal variation in expression of markers in the vitamin D pathway in prostate tissue. *Cancer Causes Control*. 2012;23(8):1359-66.
- 108.Falletti EB, D.;Fabris, C.;Fattovich, G.;Cussigh, A.;Cmet, S.;Ceriani, E.;Fornasiere, E.;Pasino, M.;Ieluzzi, D.;Pirisi, M.;Toniutto, P. Vitamin D binding protein gene polymorphisms and baseline vitamin D levels as predictors of antiviral response in chronic hepatitis C. *Hepatology*. 2012;56(5):1641-50.
- 109.Falletti EC, S.;Fabris, C.;Fattovich, G.;Cussigh, A.;Bitetto, D.;Ceriani, E.;Lenisa, I.;Dissegna, D.;Ieluzzi, D.;Rostello, A.;Pirisi, M.;Toniutto, P. Genetic polymorphisms of vitamin D pathway predict antiviral treatment outcome in slow responder naive patients with chronic hepatitis C. *PLoS One*. 2013;8(11):e80764.
- 110.Fang YVM, J. B. J.;Arp, P.;Van Leeuwen, J. P. T.;Hofman, A.;Pols, H. A. P.;Uitterlinden, A. G. Vitamin D binding protein genotype and osteoporosis. *Calcif Tissue Int*. 2009;85(2):85-93.
- 111.Grünhage FH, K.;Krawczyk, M.;Höblinger, A.;Obermayer-Pietsch, B.;Geisel, J.;Trauner, M.;Sauerbruch, T.;Lammert, F. Common genetic variation in vitamin D metabolism is associated with liver stiffness. *Hepatology*. 2012;56(5):1883-91.
- 112.Karami SA, G.;Koutros, S.;Barry, K. H.;Moore, L. E.;Han, S.;Hoppin, J. A.;Sandler, D. P.;Lubin, J. H.;Burdette, L. A.;Yuenger, J.;Yeager,

- M.;Freeman, L. E. B.;Blair, A.;Alavanja, M. C. R. Pesticide exposure and inherited variants in vitamin d pathway genes in relation to prostate cancer. *Cancer Epidemiology Biomarkers and Prevention*. 2013;22(9):1557-66.
- 113.Lange CMM, D.;Ochi, H.;Nischalke, H. D.;Bojunga, J.;Bibert, S.;Morikawa, K.;Gouttenoire, J.;Cerny, A.;Dufour, J. F.;Gorgievski-Hrisoho, M.;Heim, M. H.;Malinverni, R.;Müllhaupt, B.;Negro, F.;Semela, D.;Kutalik, Z.;Müller, T.;Spengler, U.;Berg, T.;Chayama, K.;Moradpour, D.;Bochud, P. Y.;Negro, F.;Hadengue, A.;Kaiser, L.;Rubbia-Brandt, L.;Moradpour, D.;Cellerai, C.;Rickenbach, M.;Cerny, A.;Martinetti, G.;Dufour, J. F.;Gorgievski, M.;Spicher, V. M.;Heim, M.;Hirsch, H.;Müllhaupt, B.;Helbling, B.;Regenass, S.;Malinverni, R.;Semela, D.;Dollenmaier, G.;Cathomas, G. Genetic Analyses Reveal a Role for Vitamin D Insufficiency in HCV-Associated Hepatocellular Carcinoma Development. *PLoS One*. 2013;8(5).
- 114.Laplana MS-d-l-T, M.;Puig, T.;Caruz, A.;Fibla, J. Vitamin-D pathway genes and HIV-1 disease progression in injection drug users. *Gene*. 2014;545(1):163-9.
- 115.Lasky-Su JL, N.;Brehm, J. M.;Damask, A.;Soto-Quiros, M.;Avila, L.;Celedón, J. C.;Canino, G.;Cloutier, M. M.;Hollis, B. W.;Weiss, S. T.;Litonjua, A. A. Genome-wide association analysis of circulating vitamin D levels in children with asthma. *Hum Genet*. 2012;131(9):1495-505.
- 116.Mahmoudi TK, K.;Arkani, M.;Farahani, H.;Nobakht, H.;Dabiri, R.;Asadi, A.;Vahedi, M.;Zali, M. R. Lack of associations between vitamin D metabolism-related gene variants and risk of colorectal cancer. *Asian Pac J Cancer Prev*. 2014;15(2):957-61.
- 117.Moy KAM, A. M.;Zhang, H.;Weinstein, S. J.;Wheeler, W.;Chung, C. C.;Männistö, S.;Yu, K.;Chanock, S. J.;Albanes, D. Genome-wide association study of circulating vitamin D-binding protein. *Am J Clin Nutr*. 2014;99(6):1424-31.
- 118.Pani MAR, K.;Segni, M.;Hofmann, S.;Hüfner, M.;Pasquino, A. M.;Usadel, K. H.;Badenhoop, K. A polymorphism within the vitamin D-binding protein

- gene is associated with Graves' disease but not with Hashimoto's thyroiditis. *J Clin Endocrinol Metab.* 2002;87(6):2564-7.
119. Pibiri FK, R. A.; Sandler, R. S.; Keku, T. O.; Kupfer, S. S.; Xicola, R. M.; Llor, X.; Ellis, N. A. Genetic variation in vitamin D-related genes and risk of colorectal cancer in African Americans. *Cancer Causes Control.* 2014;25(5):561-70.
120. Poynter JNJ, E. T.; Figueiredo, J. C.; Lee, W. H.; Conti, D. V.; Campbell, P. T.; Levine, A. J.; Limburg, P.; Le Marchand, L.; Cotterchio, M.; Newcomb, P. A.; Potter, J. D.; Jenkins, M. A.; Hopper, J. L.; Duggan, D. J.; Baron, J. A.; Haile, R. W. Genetic variation in the Vitamin D Receptor (VDR) and the vitamin D-binding protein (GC) and risk for colorectal cancer: Results from the colon cancer family registry. *Cancer Epidemiology Biomarkers and Prevention.* 2010;19(2):525-36.
121. Randolph AGY, W. K.; Falkenstein-Hagander, K.; Weiss, S. T.; Janssen, R.; Keisling, S.; Bont, L. Vitamin D-binding protein haplotype is associated with hospitalization for RSV bronchiolitis. *Clin Exp Allergy.* 2014;44(2):231-7.
122. Schäfer AE, S.; Kruppa, J.; Schubert, S.; Tzvetkov, M.; Mössner, R.; Reich, K.; Berking, C.; Volkenandt, M.; Pfohler, C.; Schön, M. P.; Vogt, T.; König, I. R.; Reichrath, J. No association of vitamin D metabolism-related polymorphisms and melanoma risk as well as melanoma prognosis: A case-control study. *Archives of Dermatological Research.* 2012;304(5):353-61.
123. Simon KCM, K. L.; Kraft, P.; Hunter, D. J.; De Jager, P. L.; Ascherio, A. Genetic predictors of 25-hydroxyvitamin D levels and risk of multiple sclerosis. *J Neurol.* 2011;258(9):1676-82.
124. Szkandera JA, G.; Pichler, M.; Stotz, M.; Langsenlehner, T.; Samonigg, H.; Renner, W.; Gerger, A. Association of common gene variants in vitamin D modulating genes and colon cancer recurrence. *J Cancer Res Clin Oncol.* 2013;139(9):1457-64.

125. Yan XZ, Y.;Pan, J.;Fang, K.;Wang, Y.;Li, Z.;Chang, X. Vitamin D-binding protein (group-specific component) has decreased expression in rheumatoid arthritis. *Clin Exp Rheumatol.* 2012;30(4):525-33.
126. Zhou LZ, X.;Chen, X.;Liu, L.;Lu, C.;Tang, X.;Shi, J.;Li, M.;Zhou, M.;Zhang, Z.;Xiao, L.;Yang, M. GC Glu416Asp and Thr420Lys polymorphisms contribute to gastrointestinal cancer susceptibility in a Chinese population. *Int J Clin Exp Med.* 2011;5(1):72-9.
127. Khusainova RI, Seleznyova LI, Mal'Tsev AV, Shakirova RY, Nurlygayanov RZ, Nadyrshina DD, et al. Associations between vitamin D-binding protein (DBP) gene polymorphism (TAAA) and development of osteoporosis in the Volga-Ural Region of Russia. *Bull Exp Biol Med.* 2014;157(2):253-7.
128. Kong J, Xu F, Qu J, Wang Y, Gao M, Yu H, et al. Genetic polymorphisms in the vitamin D pathway in relation to lung cancer risk and survival. *Oncotarget.* 2014.
129. Peng Q, Yang S, Lao X, Li R, Chen Z, Wang J, et al. Association of Single Nucleotide Polymorphisms in VDR and DBP Genes with HBV-Related Hepatocellular Carcinoma Risk in a Chinese Population. *PLoS One.* 2014;9(12):e116026.
130. Abbas SL, J.;Slanger, T.;Kropp, S.;Mutschelknauss, E. J.;Flesch-Janys, D.;Chang-Claude, J. The Gc2 allele of the vitamin D binding protein is associated with a decreased postmenopausal breast cancer risk, independent of the vitamin D status. *Cancer Epidemiology Biomarkers and Prevention.* 2008;17(6):1339-43.
131. Fu LY, F.;Oczak, M.;Wong, B. Y. L.;Vieth, R.;Cole, D. E. C. Common genetic variants of the vitamin D binding protein (DBP) predict differences in response of serum 25-hydroxyvitamin D [25(OH)D] to vitamin D supplementation. *Clin Biochem.* 2009;42(10-11):1174-7.
132. Johnsen MSG, G.;Figenschau, Y.;Torjesen, P. A.;Almås, B.;Jorde, R. Serum free and bio-available 25-hydroxyvitamin D correlate better with bone density than serum total 25-hydroxyvitamin D. *Scand J Clin Lab Invest.* 2014;74(3):177-83.

133. Kurylowicz AR-L, E.; Bednarczuk, T.; Badenhop, K. Vitamin D-binding protein (DBP) gene polymorphism is associated with Graves' disease and the vitamin D status in a Polish population study. *Exp Clin Endocrinol Diabetes*. 2006;114(6):329-35.
134. Ryan LMC, J. M.; Singer, S. A.; Wood, R.; Tosi, L. L.; Freishtat, R. J.; Gordish-Dressman, H.; Teach, S. J.; Devaney, J. M. Genetic influences on vitamin D status and forearm fracture risk in African American children. *J Investig Med*. 2012;60(6):902-6.
135. Wehr ET, O.; Giuliani, A.; Gruber, H. J.; Pieber, T. R.; Obermayer-Pietsch, B. Vitamin D-associated polymorphisms are related to insulin resistance and vitamin D deficiency in polycystic ovary syndrome. *European Journal of Endocrinology*. 2011;164(5):741-9.
136. Wood AMB, C.; Webster, D.; Newby, P.; Rajesh, P.; Stockley, R. A.; Thickett, D. R. Vitamin D-binding protein contributes to COPD by activation of alveolar macrophages. *Thorax*. 2011;66(3):205-10.
137. Prescott J, Bertrand KA, Reid BM, Permeth-Wey J, de Vivo I, Cramer DW, et al. Evidence of differential effects of vitamin D receptor variants on epithelial ovarian cancer risk by predicted vitamin D status. *Front Oncol*. 2014;4(OCT).
138. Elkum N, Alkayal F, Noronha F, Ali MM, Melhem M, Al-Arouj M, et al. Vitamin D Insufficiency in Arabs and South Asians Positively Associates with Polymorphisms in GC and CYP2R1 Genes. *PLoS One*. 2014;9(11):e113102.



**APPENDICES**

## APPENDIX A

### Search domains and search terms

Domain	Search terms
General domain	"allele"
	"genotype"
	"polymorphism"
	"polymorphisms"
	"SNPs"
	"single nucleotide polymorphisms"
Exposure	"rs4588"
	"rs7041"
	"rs2282679"
	"Thr420Lys"
	"Glu416Asp"
	"group specific component"
	"GC gene"
	"GC loci"
	"GC locus"
Outcome	"1, 25(OH)2D"
	"1, 25-dihydroxyvitamin D"
	"25(OH)D"
	"25 hydroxyvitamin D"
	"vitamin D"

## APPENDIX B

### Coding for the ineligibility criteria

	code	Definition
General	1	Does not have the full text of the articles available.
	2	Was not in English language.
	3	Was a narrative review or was not an observational studies (i.e. cohort, cross-sectional, or case-controls studies).
Population	4	Was not done on human subjects
Exposure	5	Did not study the effect of genotypes or alleles of any of the following SNPs of the group specific component (GC) gene; rs2282679, rs7041, or rs4588
Outcome	6	Did not measured the outcomes as mean serum 25 hydroxyvitamin D, or mean serum 1, 25 dihydroxyvitamin D levels for continuous outcomes, or vitamin D insufficiency or deficiency for dichotomous outcomes
	7	Did not have sufficient data for pooling a. For continuous outcome, reported the mean and standard deviation or the beta coefficients and the standard error of serum 25 hydroxyvitamin D or serum 1, 25 dihydroxyvitamin D levels between subjects who had or had not the interested SNPs. b. For categorical outcome; reported the contingency table of interested SNPs and vitamin D insufficiency or deficiency or the risk ratio and the 95% confidence interval or the odds ratio and the 95% confidence interval.

### APPENDIX C

Data Extraction Form

Study ID \_\_\_\_\_

#### Data Extraction Form

**Systematic review and meta-analysis of the association between vitamin D level and the single nucleotide polymorphisms, rs7041, rs4588, and rs2282679 of the group-specific component gene (GC).**

Reviewer (name) .....

Author: \_\_\_\_\_

Journal: \_\_\_\_\_

Year: \_\_\_\_\_ Volume: \_\_\_\_\_ Pages: \_\_\_\_\_

Type of Study Design [ ]

- 1) Cohort study
- 2) Case-control study
- 3) Cross-sectional study
- 8) Other

If other, specify .....

**Study age group** [ ]

- 1) < 18 years
- 2) ≥ 18 years to <50 years
- 3) age ≥ 50 years
- 4) ≥ 18 years

**Ethnicity** [ ]

- 1) White
- 2) Black
- 3) Hispanic
- 4) Mediterranean
- 5) Indian
- 6) Asian
- 7) Mixed
- 8) Other

If other, specify .....

Country study was conducted in .....

Type of studied subjects [ ]

- 1) General subjects
- 2) Disease subjects specify .....

**Study factors**

**Single nucleotide polymorphisms (SNP)**

Type of SNP

- 1) rs4588 [ ] 2) rs7041 [ ] 3) rs2282679 [ ]
- Others \_\_\_\_\_

Method of genotyping

.....

Hardy-Weinberg Equilibrium (HWE) [ ]

- 1) Done
- 2) Not done
- 9) Unknown

Vitamin D supplement [ ]

- 1) Taken
- 2) Not taken
- 9) Unknown

Specify the amount taken, if given in the study

.....

.....

.....

.....

.....

.....

SNP data (tick in the brackets means yes)

- [ ] Genotype data [ ] Haplotype
- [ ] Allele data [ ] Interaction

**Outcome**

Continuous outcome [ ]

- 1) 25 (OH) D, (25, hydroxyvitamin D)
- 2) 1, 25 (OH)<sup>2</sup> D, (1, 25 dihydroxyvitamin D)

Dichotomous outcome [ ]

- 1) Vitamin D deficient      specify cut off  
.....
- 2) Vitamin D insufficient      specify cut off  
.....

Method used to measure vitamin D

.....

<b>Baseline characteristics</b>				
Mean Age				
Mean BMI				
%Male				
%Active Physical activity				
%Alcohol				
% vitamin D supplementation				
Kidney disease				
Liver disease				
Serum vitamin D level				
Sun exposure				
% smoking				

Categorical outcome

Group	Vitamin D deficiency [ ] Vitamin D insufficiency [ ]		OR	95%CI	
	Yes	No			
rs2282679 Genotype data					
TT					
GT					
GG					
Allele data					
T					
G					
rs7041 Genotype data					

GG						
TG						
TT						
Allele data						
G						
T						
rs4588 Genotype data						
CC						
AC						
AA						
Allele data						
C						
A						
*Used to denote the reference group						
group	n	mean	SD	Beta	SE	
rs2282679 Genotype data						
TT						
GT						
GG						

Allele data						
T						
G						
rs7041 Genotype data						
GG						
TG						
TT						
Allele data						
G						
T						
rs4588 Genotype data						

CC						
AC						
AA						
Allele data						
C						
A						

\*Used to denote the reference group

## APPENDIX D

### Risk of bias form

Risk of bias assessment for genetic association studies of association between Vitamin D and the Single Nucleotide Polymorphisms, rs7041, rs4588, and rs2282679 of the group-specific complement gene (GC)

For case control study designs <sup>39</sup>

Domain	Item	How to grade	Low risk of bias
Selection bias	Representativeness of cases		
	A. Consecutive/randomly with a random frame.	Consecutive/randomly selected from cases population with a clearly defined random frame.	Yes
	B. Consecutive/randomly without a random frame.	Consecutive/randomly selected from cases population without clearly defined random frame or with extensive inclusion criteria	Yes
	C. Not described.	If the method of selection is not described.	No
	Representativeness of controls		
	D. Consecutive/randomly from same area	Controls were consecutive/randomly drawn from same area (ward/community) as cases with the same criteria	Yes
	E. Consecutive/randomly from different area	Controls were consecutively/randomly drawn from different areas as cases	No
	F. Not describe	Not describe	No
	Differential participation in case and control		
	A. Non-participant rate <10%	Non-participant rate is small (< 10%) and similar (to the rates) between case and control groups. The non-participant rate did not exceed 10% of the estimated sample size for the study.	Yes
	B. Incomplete participant rates	Incomplete participant rates are different - Refusal or inability to provide data - Refusal or inability to provide biological specimens - Insufficient amount quality of data/ quality of DNA	No
Information bias	Ascertainment of vitamin D level		

	<p>A. Method of vitamin D measurement and definition given.</p> <p>B. Not describe/unclear.</p>	<p>- If the method of vitamin D measurement was clearly given with a clear definition of vitamin D deficiency or insufficiency.</p> <p>- Not describe/unclear definition.</p>	<p>Yes</p> <p>No</p>
	<p>Ascertainment of control</p> <p>A. Method of measurement given.</p> <p>B. Only vitamin D level given.</p> <p>C. Not describe.</p>	<p>- If the method of vitamin D measurement was clearly given.</p> <p>- Just mentioned the vitamin D levels of the controls.</p> <p>- Not describe.</p>	<p>Yes</p> <p>No</p> <p>No</p>
	<p>Ascertainment of genotyping examination</p> <p>A. Genotyping done 'blind' for case and control groups.</p> <p>B. Genotyping error rate &lt; 5%.</p> <p>C. Genotyping error arte not given.</p> <p>D. Quality control procedure if replicate sample.</p> <p>Analysis if replicate sample</p> <p>E. Not blinded.</p> <p>F. Not mention what was done</p> <p>G. No quality control check</p>	<p>- Genotyping done under "blind" condition of case and control group's specimens</p> <p>- Genotyping error rate &lt; 5%.</p> <p>- Genotyping error arte not given.</p> <p>- Quality control procedure e.g., reanalysis of random specimens, using different genotyping methods for analysis.</p> <p>Analysis if replicate sample.</p> <p>- Not blinded</p> <p>- Not mention what was done</p> <p>- No quality control check</p>	<p>Yes</p> <p>Yes</p> <p>No</p> <p>Yes</p> <p>No</p> <p>No</p> <p>No</p>
Confounding bias	<p>Population stratification</p> <p>A. Same ethnicity of case and control</p> <p>B. Cases not related to controls</p> <p>C. Genomic controls</p> <p>D. Not report</p> <p>Other confounding bias</p> <p>E. Confounding variables reported</p> <p>F. Not controlled/not mentioned</p>	<p>- No difference in ethnic origin between cases and controls</p> <p>- Use of controls who were not related to cases</p> <p>- Use of genomic controls</p> <p>- Not report what was done</p> <p>- Controls for at least 2 or 3 confounding variables (e.g., age, gender, obesity, physical activity, kidney disease, liver disease, sun exposure, alcohol intake, vitamin D supplement) in analysis</p> <p>- Not controlled /not mentioned (or, no control/ no mention)</p>	<p>Yes</p> <p>Yes</p> <p>Yes</p> <p>No</p> <p>Yes</p> <p>No</p>
Multiple testing	<p>Multiple polymorphisms studied and multiple comparisons done.</p> <p>A. Adjusted for multiple</p>	<p>- Adjustment for multiple tests such as for e.g., Bonferroni correction, Sidak correction or the Holm-</p>	<p>Yes</p>

	<p>tests.</p> <p>B. No adjustments done.</p> <p>C. Regression analysis was done.</p> <p>D. Multiple pair wise comparison done for genetic model assessment.</p> <p>E. A genetic model was assumed and analyzed for many.</p>	<p>Bonferroni method has been done.</p> <p>- No adjustment for multiple testing was done</p> <p>- Regression analysis was done.</p> <p>- For genetic model assessment a multiple pair wise comparison was done.</p> <p>- A genetic model was assumed but analyzed for many models.</p>	<p>No</p> <p>Yes</p> <p>Yes</p> <p>No</p>
Selective reporting (for replication studies)	<p>A. Report for all polymorphisms.</p> <p>B. Report selective</p>	<p>- Report results of all polymorphisms mentioned in objectives, non-significant or not</p> <p>- Report results of only significant polymorphisms</p>	<p>Yes</p> <p>No</p>
HWE	<p>A. has HWE</p> <p>B. has HWD</p> <p>C. not check HWE</p>	<p>- HWE in control group</p> <p>- HW disequilibrium in control group</p> <p>- Not check HWE</p>	<p>Yes</p> <p>No</p> <p>No</p>
Yes=low/no risk of bias, No = possible/high risk of bias, or information is unclear or not given			

For cohort and cross-sectional study designs <sup>39</sup>				
Domain	Item	How to grade	Low risk of bias	
Selection bias	Representativeness	Consecutive/randomly selected from target population with a clearly defined random frame. Consecutive/randomly selected from target population without clearly defined random frame or with extensive inclusion criteria If the method of selection is not described.	Yes	
	A. Consecutive/randomly with a random frame.			Yes
	B. Consecutive/randomly without a random frame.			No
Information bias	Ascertainment of vitamin D level	- If the method of vitamin D measurement was clearly given with a clear definition of vitamin D deficiency or insufficiency.  - Not describe/unclear definition	Yes	
	A. Method of vitamin D measurement and definition given			No
	B. Not describe/unclear			
	Ascertainment of non-exposure group	- If the method of vitamin D measurement was clearly given	Yes	
	D. Method of measurement given			

	E. Only vitamin D level given F. Not describe	- Just mentioned the vitamin D levels of the controls - Not describe	No No
	Ascertainment of genotyping examination A. Genotyping done 'blind' for exposure and non-exposure groups. B. Genotyping error rate < 5%. C. Genotyping error arte not given. D. Quality control procedure if replicate sample. Analysis if replicate sample A. Not blinded. B. Not mention what was done C. No quality control check	- Genotyping done under "blind" condition of exposure and non-exposure group's specimens  - Genotyping error rate < 5%.  - Genotyping error arte not given.  - Quality control procedure e.g., reanalysis of random specimens, using different genotyping methods for analysis. Analysis if replicate sample. - Not blinded - Not mention what was done  - No quality control check	Yes  Yes  No  Yes  No No No
Confounding bias	Population stratification A. Same ethnicity of exposure and non-exposure groups. B. exposure group subjects not related to controls C. Genomic controls D. Not report  Other confounding bias E. Confounding variables reported  F. Not controlled/not mentioned	- No difference in ethnic origin between exposure and non-exposure groups.  - Use of controls who were not related to exposure group subjects.  - Use of genomic controls - Not report what was done  - Controls for at least 2 or 3 confounding variables (e.g., age, gender, obesity, physical activity, kidney disease, liver disease, sun exposure, alcohol intake, vitamin D supplement) in analysis - Not controlled /not mentioned (or, no control/ no mention)	Yes  Yes  Yes No  Yes  No
Multiple testing	Multiple polymorphisms studied and multiple comparisons done. A. Adjusted for multiple tests. B. No adjustments done. C. Regression analysis was done. D. Multiple pair wise comparison done for	- Adjustment for multiple tests such as for e.g., Bonferroni correction, Sidak correction or the Holm-Bonferroni method has been done. - No adjustment for multiple testing was done	Yes  No Yes

	<p>genetic model assessment. E. A genetic model was assumed and analyzed for many.</p>	<p>- Regression analysis was done.  - For genetic model assessment a multiple pair wise comparison was done.  - A genetic model was assumed but analyzed for many models.</p>	<p>Yes  No</p>
<p>Selective reporting (for replication studies)</p>	<p>A. Report for all polymorphisms. B. Report selective</p>	<p>- Report results of all polymorphisms mentioned in objectives, non-significant or not  - Report results of only significant polymorphisms</p>	<p>Yes  No</p>
<p>HWE</p>	<p>A. has HWE B. has HWD C. not check HWE</p>	<p>- HWE in control group - HW disequilibrium in control group - Not check HWE</p>	<p>Yes No No</p>

Yes=low/no risk of bias, No = possible/high risk of bias, or information is unclear or not given

## BIOGRAPHY

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